

P.B.5818 - Patentlaan 2 2280 HV Rijswijk (ZH) 3 + 31 70 340 2040 TX 31651 epo nl FAX + 31 70 340 3016

Europäisches Patentamt

Zweigstelle in Den Haag Recherchenabteilung

European Patent Office

Branch at The Hague Search division Office européen des brevets

Département à La Haye Division de la recherche

Holdcroft, James Gerald, Dr. Graham Watt & Co.
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7-9 St. Botolph's Road
Sevenoaks, Kent TN13 3AJ
GRANDE BRETAGNE

Datum/Date

2 3. 06. 2003

Zeichen/Ref./Réf. JGH 13973 PCTEP	Anmeldung Nr./Application No./Demande n°./Patent Nr./Patent No./Brevet n°. 98949803.5
Anmelder/Applicant/Demandeur/Patentinhaber/Proprietor/Titulaire CeNeS Pharmaceuticals, Inc.	

COMMUNICATION

RECEIVED

			• ,
The Europe	ean Patent Office herewith transmits		OCT 0 2 2002
	the European search report		OCT 0 3 2003
	the declaration under Rule 45 EPC		TECH CENTER 1600/2900
	the partial European search report und	der Rule 45 EPC	
X		eport concerning the international application under lean patent application. Copies of the documents of	
The followin	ng specifications given by the applicant h	ave been approved by the Search Division:	
	Abstract	Title	Figure
	The abstract was modified by the Search	h Division and the definitive text is attached to this	communication.
	The following figure will be published wi	th the abstract, since the Search Division consider	s that it better characterises

REFUND OF THE SEARCH FEE

the invention than the one indicated by the applicant.

Additional copy(copies) of the documents cited in the European search report.

Figure:

If applicable under Article 10 Rules relating to fees, a separate communication from the Receiving Section on the refund of the search fee will be sent later.



EPO Form 1507 0	2.93		

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SUPPLEMENTARY PARTIAL EUROPEAN SEARCH REPORT

Application Number

which under Rule 45 of the European Patent ConventionEP 98 94 9803 shall be considered, for the purposes of subsequent proceedings, as the European search report

	Citation of document with in	dication, where appropriate,	Relevant	CLASSIFICATION OF THE
ategory	of relevant pass		to claim	APPLICATION (Int.Cl.6)
X	WO 96 15812 A (MARCI	HIONNI MARK A ; MCBURNEY	/ 1-4	A61K38/18
•	ROBERT N (US); GWYN	NE DAVID I (US); BE)		C07K14/475
	30 May 1996 (1996-09	5-30)		A61K48/00
,	* see claims 32-35,	fig 11, 13.0, 14-16,	1-4	A61P25/00
	24-25, page 18 and	page 30 line 1-13 *		
•	UO OC 20402 A / DEDM	 INGHAM MCDONOGH OLIVIA	1-4	
Ĭ.	WO 96 30403 A (BENIII)	(US); GWYNNE DAVID I)		
	3 October 1996 (199	6-10-03)		
	* see claims 45-46	and 58, fig. 13.0 and	1-4	
	page 30 *			
,	MADOUTONNT M A FT	 AL.: "Neuregulins as	1-4	
•	potential drugs for	neurological		
	disorders."			
	COLD SPRING HARBOR			
	QUANTITATIVE BIOLOG	r., s 459-472, XP009007573		
	* see the whole doc	ument *		TECHNICAL FIELDS SEARCHED (Int.CI.6)
	·			A61K C07K
				A61P
The s	upplementary search report has b	een based on the last set of claims varch.	alid ——————	4
	MPLETE SEARCH			
not come	sty with the EPC to such an extent that	application, or some or all of its claims, doe a meaningful search into the state of the a	es/do rt cannot	
be carrie	d out, or can only be carried out partial	ly, for the following claims:		
Claims s	earched completely :			
Claims s	earched incompletely :			
o				
	ot searched:			
5 Reason	for the limitation of the search:			
Cla	im 5 does not contai	n any additional limit	ing	
t.ec	hnical featur as com	pared to the preceding	claims.	
Def	ining a product by i oit the scope of a cl	ts desired effect does	กดเ	
110	it the scope of a ci			
	Place of search	Date of completion of the search		Examiner
	MUNICH	17 March 2003	Me	rckling, V
			rinte underlying th	e invention
	CATEGORY OF CITED DOCUMENTS	T : theory or princ E : earlier patent	document, but pu	blished on, or
X:pa	rticularly relevant if taken alone	E : earlier patent after the filing	document, but pu date	blished on, or
X:pa Y:pa doo		E : earliër patent after the filling ther D: document cite L : document cite	document, but pu date ed in the applicatio	blished on, or on s

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Application Number

EP 98 94 9803

The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:

1. Claims: 1-4 (part)

European Patent

Office

Use of the neuregulin comprising aminoacid Seq. ID No.2,n or encoded by nucleic acid Seq. ID No.1, for the manufacture of a medicament for treating a mammal suffering from one of the diseases listed in claim 1.

2. Claims: 1-4 (part)

Inventions 2-X: Use of the neuregulin comprising one of the aminoacid sequences as defined in claim 2 (or of a gene encoding such a polypeptide), for the manufacture of a medicament for treating a mammal suffering from one of the diseases listed in claim 1.

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CLAIMS INCURRING FEES
The present European patent application comprised at the time of filing more than ten claims.
Only part of the claims have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims and for those claims for which claims fees have been paid, namely claim(s):
No claims fees have been paid within the prescribed time limit. The present European search report has been drawn up for the first ten claims.
LACK OF UNITY OF INVENTION
The Search Division considers that the present European patent application does not comply with the requirements of unity of invention and relates to several inventions or groups of inventions, namely:
see sheet B
1
All further search fees have been paid within the fixed time limit. The present European search report has been drawn up for all claims.
As all searchable claims could be searched without effort justifying an additional fee, the Search Division did not invite payment of any additional fee.
Only part of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the inventions in respect of which search fees have been paid, namely claims:
None of the further search fees have been paid within the fixed time limit. The present European search report has been drawn up for those parts of the European patent application which relate to the invention first mentioned in the claims, namely claims: 1-4 (part.)

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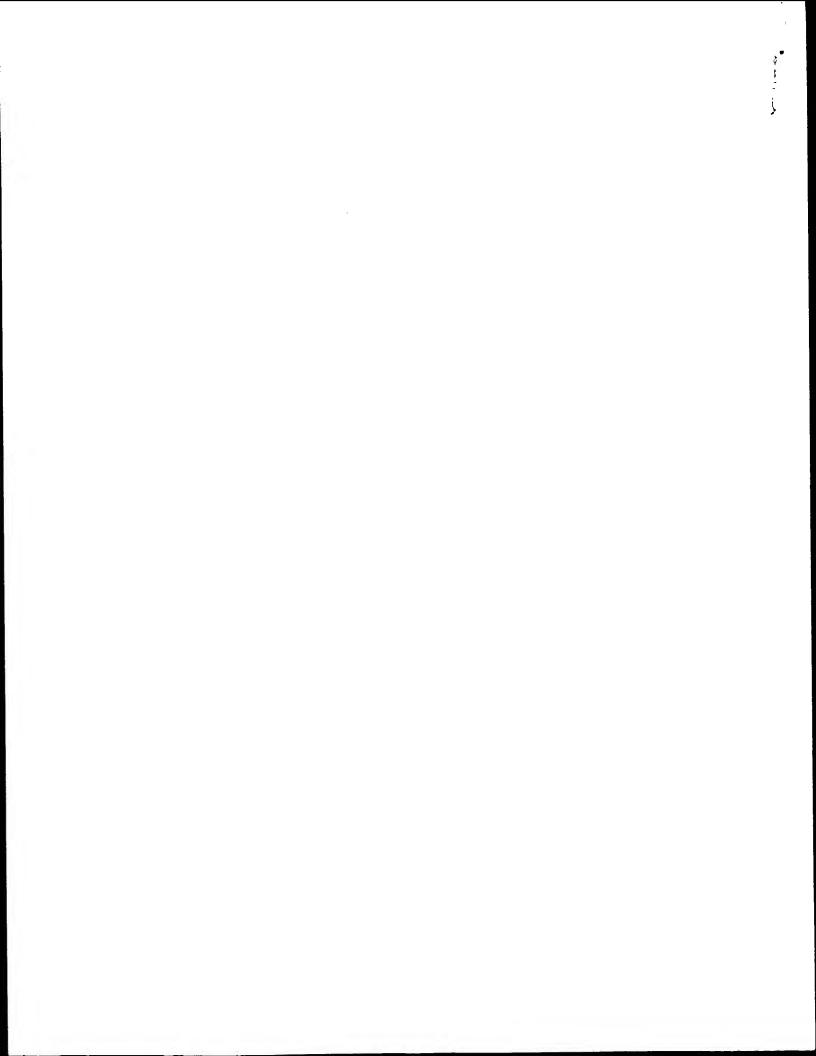
ANNEX TO THE EUROPEAN SEARCH REPO ON EUROPEAN PATENT APPLICATION NO.

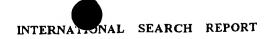
EP 98 94 9803

This annex lists the patent family members relating to the patent documents cited in the above-mentioned European search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

17-03-2003

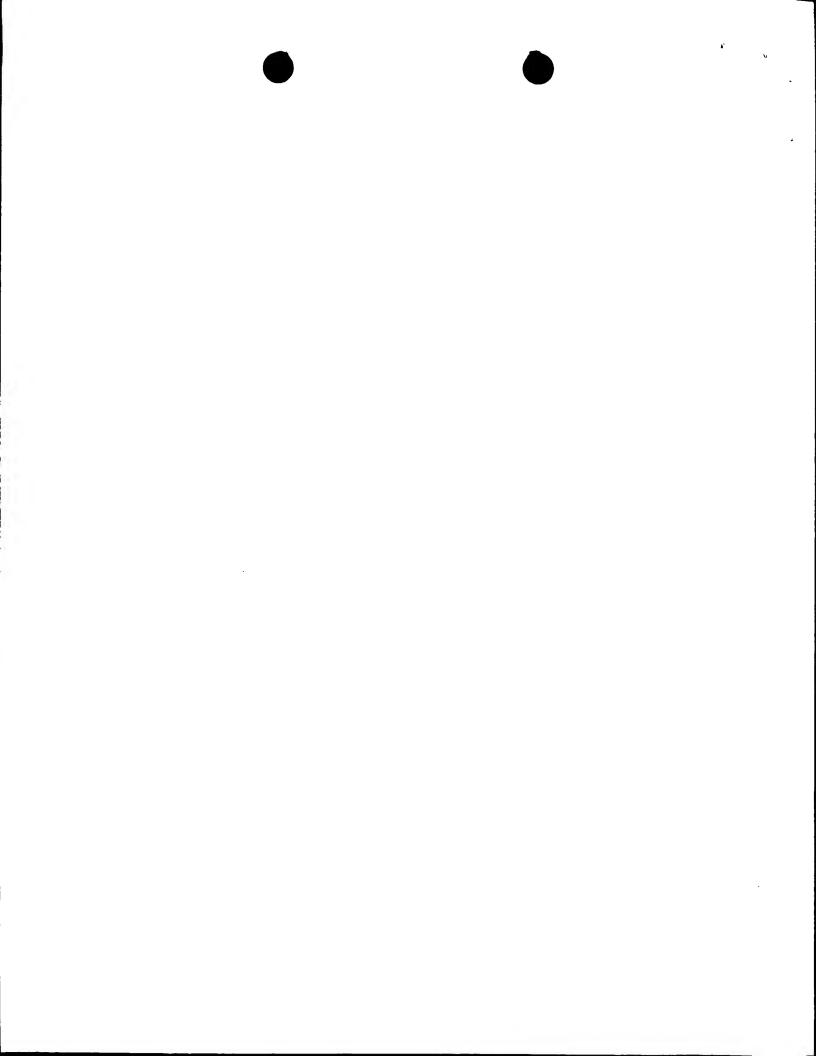
Patent document cited in search repo		Publication date	Patent family member(s)	Publication date
WO 9615812	A	30-05-1996	US 6087323 A AU 707599 B AU 4238496 A CA 2204850 A EP 0784488 A JP 10509717 T US 2003040465 A	11-07-2000 15-07-1999 17-06-1996 30-05-1996 23-07-1997 22-09-1998 27-02-2003
WO 9630403	Α	03-10-1996	AU 713384 B AU 5376696 A CA 2215330 A EP 0820467 A JP 11509831 T	02-12-1999 16-10-1996 03-10-1996 28-01-1998 31-08-1999





International application No.
PCT/US98/21349

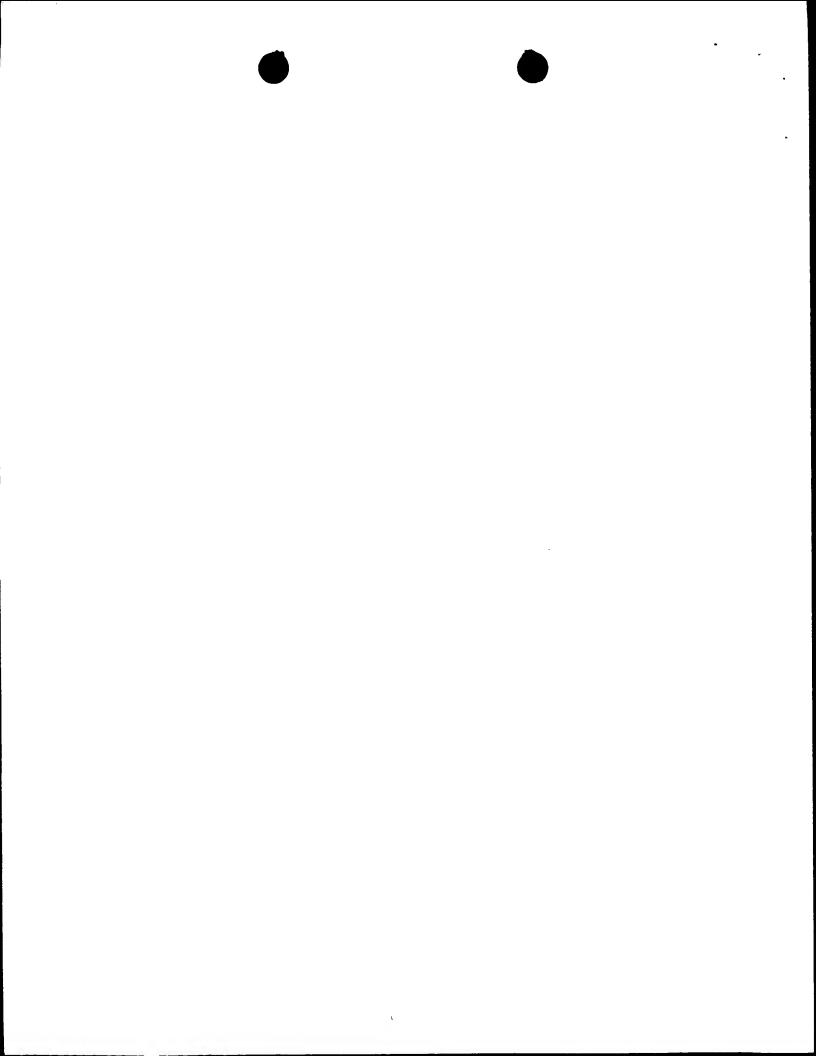
A. CLAS	A. CLASSIFICATION OF SUBJECT MATTER					
	A61K 31/70, 38/00, 38/02, 38/18					
US CL :	514/2, 12, 44, 903, 907 International Patent Classification (IPC) or to both n	ational classification and IPC				
	DS SEARCHED					
	cumentation searched (classification system followed	by classification symbols)				
	14/2, 12, 44, 903, 907					
Documentati	on searched other than minimum documentation to the	extent that such documents are included	in the fields searched			
		me of data base and where practicable	search terms used)			
	ata base consulted during the international search (nat	me of data base and, whole processes	, 202.0 101.110 11200)			
Please Sec	Extra Sheet.					
C. DOC	UMENTS CONSIDERED TO BE RELEVANT					
Category*	Citation of document, with indication, where app	propriate, of the relevant passages	Relevant to claim No.			
Х	US 5,530,109 A (GOODEARL et al.)	25 June 1996, column 3, line	1-34			
^	3 to column 6, line 53 and column 11,	line 1 to column 12, line 59.				
	<i>5</i> to column o, 11110					
		•				
			•			
			•			
			<u> </u>			
Furt	her documents are listed in the continuation of Box C					
-	pecial categories of cited documents:	"T" later document published after the int date and not in conflict with the app	lication but cited to understand			
to	comment defining the general state of the art which is not considered be of particular relevance	the principle or theory underlying the "X" document of particular relevance; the	ne claimed invention cannot be			
	arlier document published on or after the international filing date	considered novel or cannot be considered when the document is taken alone	ered to involve an inventive step			
l ci	ocument which may throw doubts on priority claim(s) or which is ted to establish the publication date of another citation or other	eve document of particular relevance; the	ne claimed invention cannot be			
at.	ecial reason (as specified) comment referring to an oral disclosure, use, exhibition or other	considered to involve an inventive combined with one or more other suc	e step when the document is the documents, such combination			
m m	ceans ocument published prior to the international filing date but later than	being obvious to a person skilled in	the art			
th:	e priority date claimed					
Date of the	actual completion of the international search	Date of mailing of the international se				
17 DECE	17 DECEMBER 1998 Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Authorized officer STEPHEN GUCKER STEPHEN GUCKER					
Name and mailing address of the ISA/US Authorized officer						
Commissioner of Patents and Trademarks Box PCT STEPHEN GUCKER						
	on, D.C. 20231 No. (703) 305-3230	Telephone No. (703) 308-0196	<i>/-</i>			
Facsimile 1	No. (703) 305-3230	· · · · · · · · · · · · · · · · · · ·				





International application No.
PCT/US98/21349

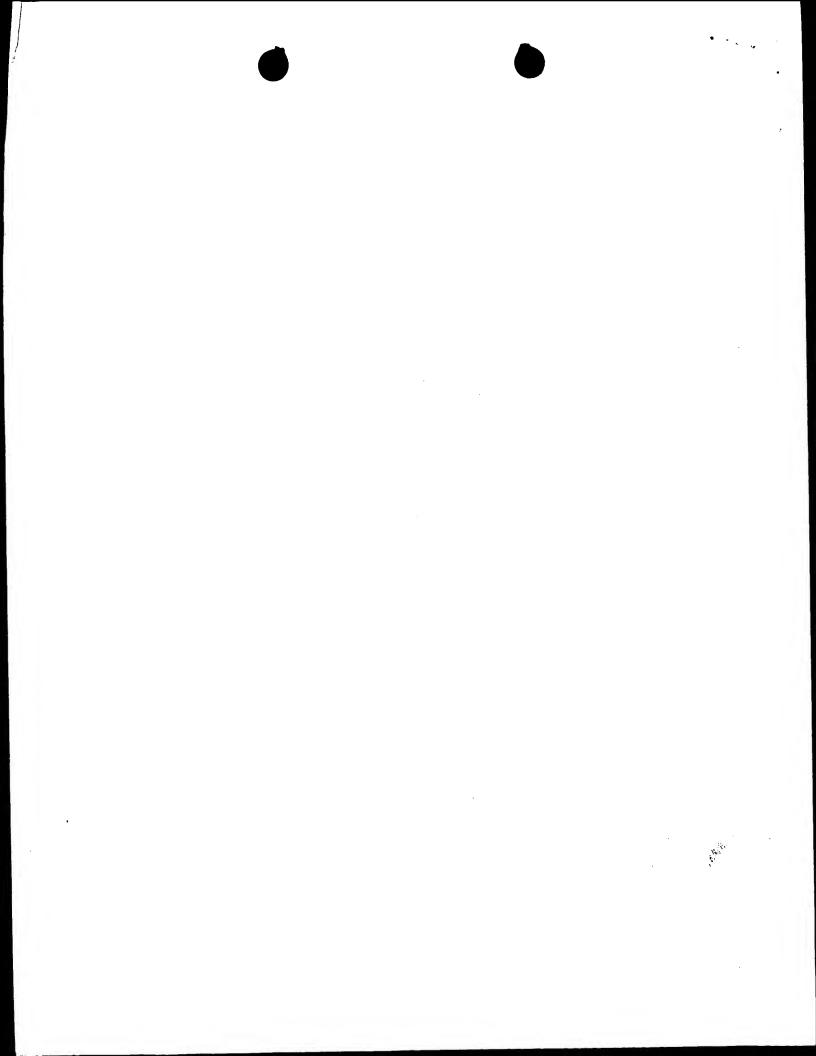
Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)				
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:				
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:				
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:				
Claims Nos.: 35-36 because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).				
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)				
This International Searching Authority found multiple inventions in this international application, as follows:				
·				
As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.				
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.				
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:				
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:				
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees				
No protest accompanied the payment of additional search fees.				



B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, MEDLINE, SCISEARCH, EMBASE, BIOSIS, CAPLUS, WPIDS, BIOTECHDS, CONFSCI, LIFESCI neuregulin#, glia#, heregulin#, GGF#, ischemia#, dementia#, Parkinson#, Huntington#, Alzheimer#, infarct#, amyotrophic, Down#, Korsakoff#, heart#, cardiac, spinal



PATENT COOPERATION TREATY

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INTERNATIONAL PRELIMINARY EXAMINATION REPORT

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(PCT Article 36 and Rule 70)

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Applicant's or agent's file reference 47440-PCT	FOR FURTHER ACTION Sec No Prelimin	TECH CENTER : 00 tification of Transmittal of International (Natural Control of Transmittal of International (Natural Control of Transmittal of International Control of Transmittal (Natural Control of Transmittal Control of Transmittal (Natural Control of Transmittal Control of Transmittal Control of Transmittal (Natural Control of Transmittal Control of Transmittal Control of Transmittal (Natural Control of Transmittal Control of T
International application No.	International filing date (day/month/year)	
PCT/US98/21349	08 OCTOBER 1998	14 OCTOBER 1997
International Patent Classification (IPC) IPC(7): A61K 31/70, 38/00, 38/02, 38/	or national classification and IPC /18 and US Cl.: 514/2, 12, 44, 903, 907	
Applicant CAMBRIDGE NEUROSCIENCE, INC.		Anthri
This international prelimination Examining Authority and is	ary examination report has been prepared transmitted to the applicant according	pared by this International Preliminary to Article 36.
2. This REPORT consists of a	total ofsheets.	
(see Rule 70.16 and Sect	ion 607 of the Administrative Instructions	escription, claims and/or drawings which have ning rectifications made before this Authority. s under the PCT).
These annexes consist of a to		
3. This report contains indications	s relating to the following items:	
I X Basis of the repor	t	
II Priority		
III X Non-establishment	t of report with regard to novelty, inve	ntive step or industrial applicability
IV Lack of unity of i		·
V X Reasoned statement citations and explan	t under Article 35(2) with regard to nove actions supporting such statement	lty, inventive step or industrial applicability;
VI Certain documents of	pited	
VII Certain defects in th	e international application	
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22 APRIL 1999	26 MAY 2000	0
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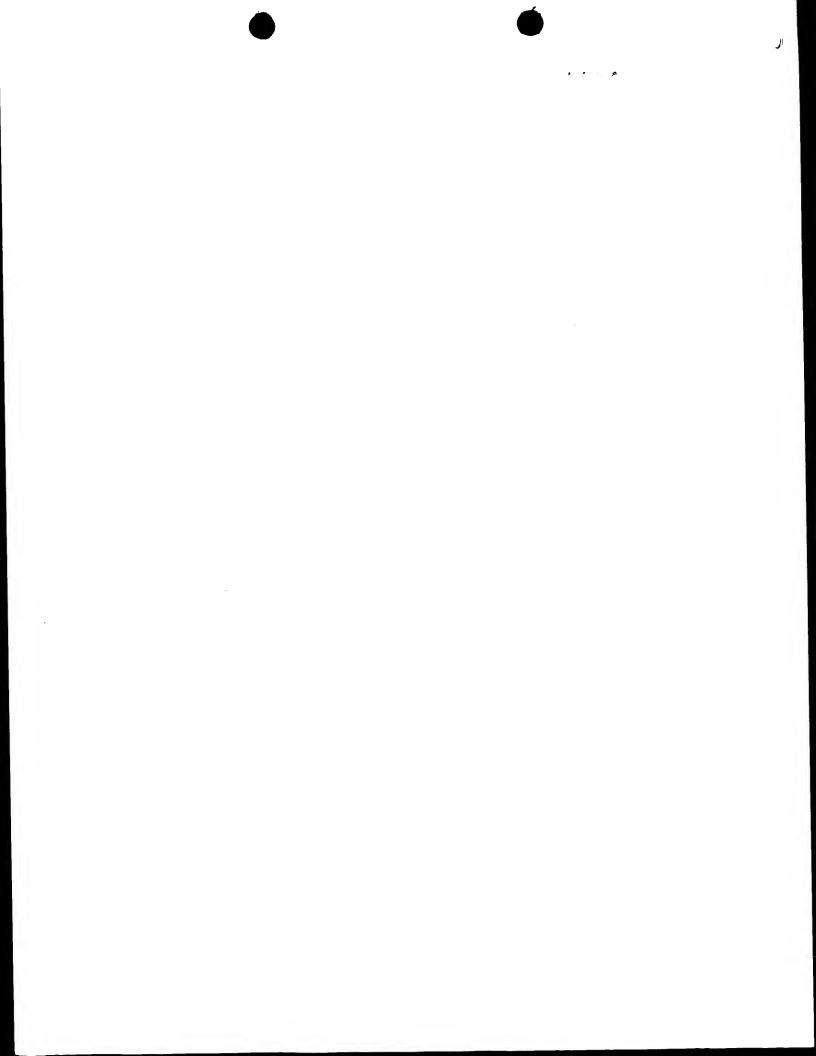
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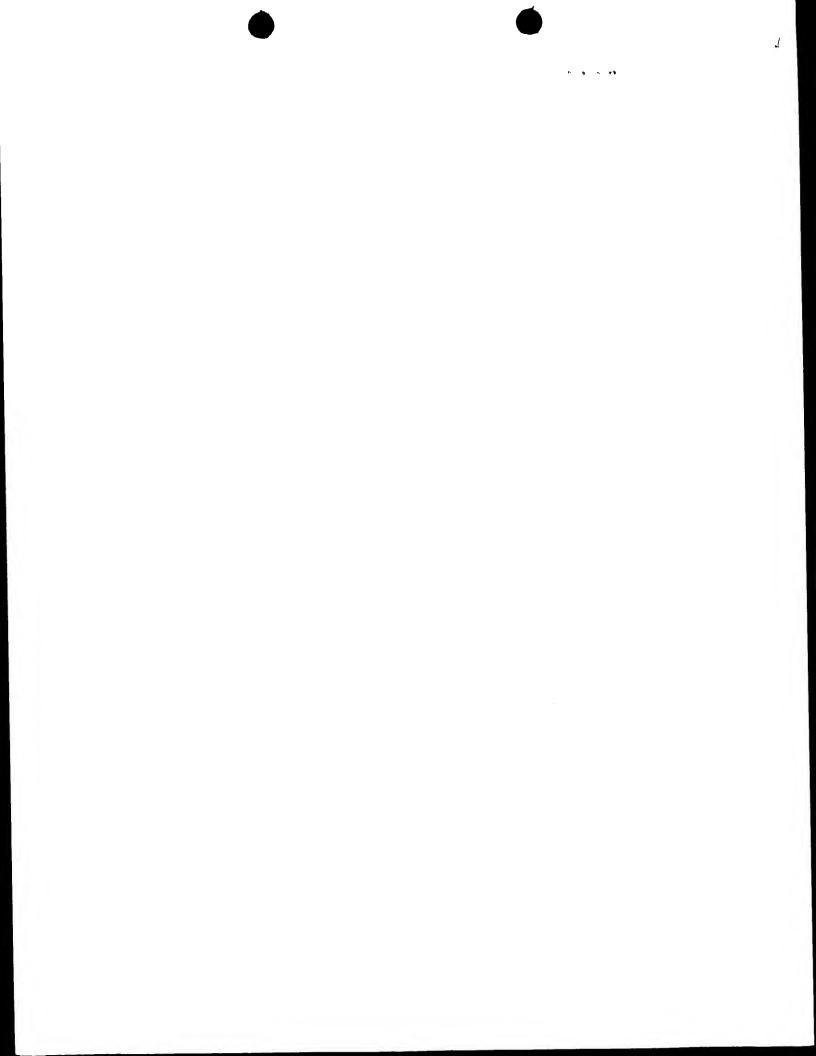
PCT/US98/21349

I.	Basis c	of the report				
1. V	With regar	rd to the elements of the inter	— tional applia			
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Ŀ	لت	claims:				
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- P	Vith regard	rd to any nucleotide and/ory examination was carried	or amino acid d out on the b	sequence disclosed in passis of the sequence	in the international o	application, the international
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	X.	the description, pages	NONE			
		the claims, Nos.	NONE			
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and	70.17).	ongnamy jace and	are not unnext	ea to trus report since	they do not contain	der Article 14 are referred to n amendments (Rules 70.16
**Any	y replace:	ment sheet containing such	ı amendments i	must be referred to un	nder item 1 and ann	aved to this penses



International application No. PCT/US98/21349

m.	Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
1. T.	he questions whether the claimed invention appears to be novel, to involve an inventive step (to be non obvious), or to be adustrially applicable have not been and will not be examined in respect of:
	the entire international application.
×	claims Nos. <u>35-36</u>
	because:
	the said international application, or the said claim Nos. relate to the following subject matter which does not require international preliminary examination (specify).
X	the description, claims or drawings (indicate particular elements below) or said claims Nos. 35-36 are so unclear that no meaningful opinion could be formed (specify).
beca	ause they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
	the claims, or said claims Nos are so inadequately supported by the description that no meaningful opinion could be formed.
	no international search report has been established for said claims Nos
z. A. m. sequa	eaningful international preliminary examination cannot be carried out due to the failure of the nucleotide and/or amino acid ence listing to comply with the standard provided for in Annex C of the Administrative Instructions:
	the written form has not been furnished or does not comply with the standard.
Ш	the computer readable form has not been furnished or does not comply with the standard.



International application No.

PCT/US98/21349

Novelty (N)			
()	Claims	NONE	VD
	Claims	1-34	YE
Inventive Step (IS)	Claims	NONE	
	Claims	1-34	YES NO
Industrial Applicability (IA)	Claims	1-34	VES
	Claims	NONE	YES
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International application No. PCT/US98/21349

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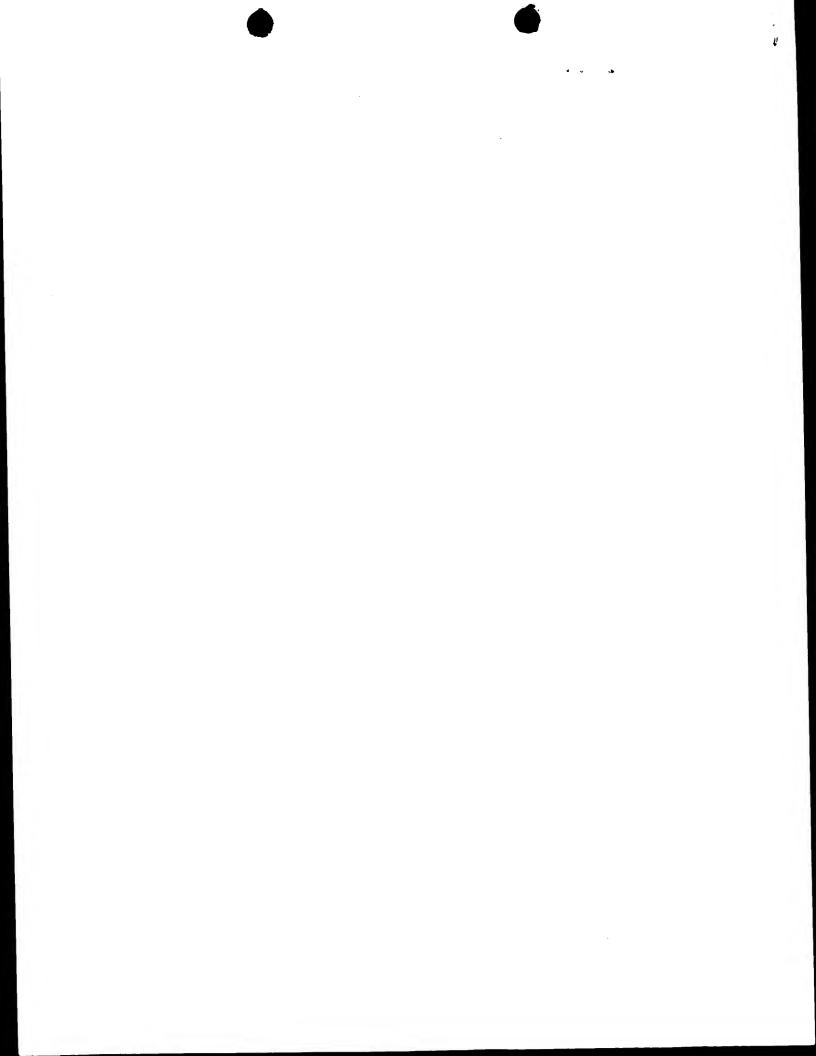
(To be used when the space in any of the preceding boxes is not sufficient)

Continuation of: Boxes I - VIII

Sheet 10

- I. BASIS OF REPORT:
- 5. (Some) amendments are considered to go beyond the disclosure as filed: NONE

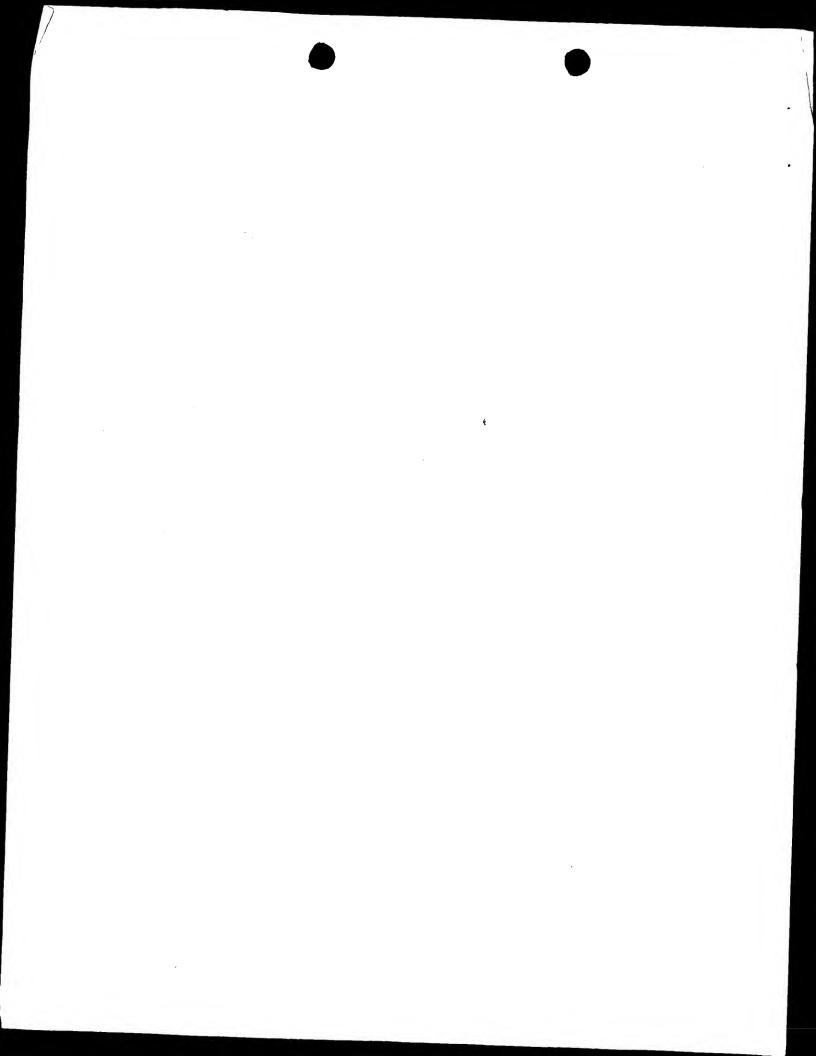
Form PCT/IPEA/409 (Supplemental Box) (July 1998)★



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			ES	Spain	LT	Lithuania	SN	Senegal
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1	BA	Bosnia and Herzegovina	GH	Ghana	MK	The former Yugoslav	TR	Turkey
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١	BE	Belgium	GR	Greece	ML	Mali	UA	Ukraine
1	BF	Burkina Faso	HU	Hungary	MN	Mongolia	UG	Uganda
1	BG	Bulgaria	IE	Ireland	MR	Mauritania	US	United States of America
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١	CF		KE	Kenya	NO	Norway		
١	CG	Congo	KG	Kyrgyzstan	NZ	New Zealand		
١	CH	Switzerland	KP	Democratic People's	PL	Poland		
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PCT





INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶: A61K 31/70, 38/00, 38/02, 38/18

A1 (11) International Publication Number:

WO 99/18976

US

(43) International Publication Date:

22 April 1999 (22.04.99)

(21) International Application Number:

PCT/US98/21349

(22) International Filing Date:

8 October 1998 (08.10.98)

(30) Priority Data:

60/062,109

14 October 1997 (14.10.97)

(74) Agents: CONLIN, David, G. et al.; Dike, Bronstein, Roberts & Cushman, LLP, 130 Water Street, Boston, MA 02109 (US).

(63) Related by Continuation (CON) or Continuation-in-Part (CIP) to Earlier Application

Filed on

60/062,109 (CON) 14 October 1997 (14.10.97)

(71) Applicant (for all designated States except US): CAMBRIDGE NEUROSCIENCE, INC. [US/US]; Building 700, One Kendall Square, Cambridge, MA 02139 (US).

(72) Inventors; and

(75) Inventors/Applicants (for US only): MCBURNEY, Robert, N. [US/US]; 20 Leslie Road, Newton, MA 02166 (US). HOLT, William [US/US]; 162 A East Central Street, Natick, MA 01760 (US). GWYNNE, David, I. [US/US]; 77 Grover Street, Beverly, MA 01915 (US). MARCHIONNI, Mark [US/US]; 24 Twin Circle Drive, Arlington, MA 02174 (US).

(81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, HR, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

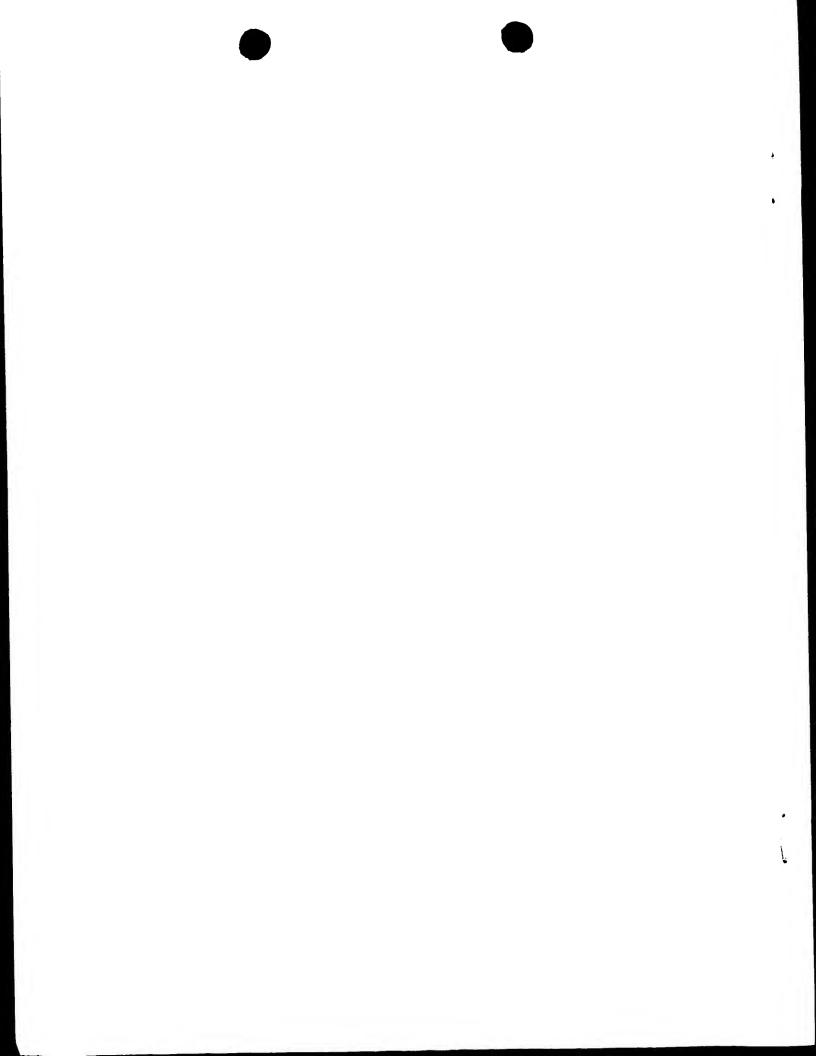
Published

With international search report.

(54) Title: THERAPEUTIC METHODS COMPRISING USE OF A NEUREGULIN

(57) Abstract

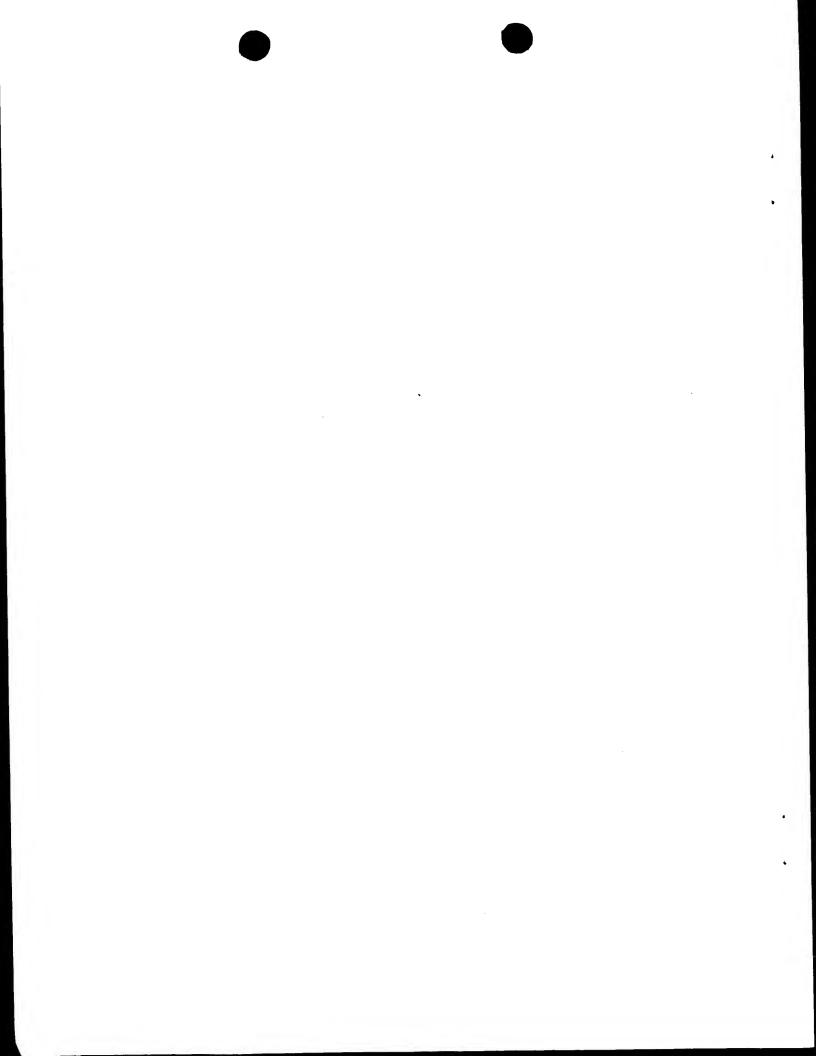
The invention provides methods for treatment and/or prophylaxis of certain neurological-related disorders, particularly treatment or prophylaxis of the effects of stroke, brain or spinal cord injury or ischemia, heart attack, optic nerve and retinal injury and ischemia and other acute-type conditions disclosed herein as well as chronic-type conditions, specifically epilepsy, Alzheimer's disease, Parkinson's disease, Huntington's disease, Amyotrophic Lateral Sclerosis, Down's Syndrome, Korsakoff's disease, cerebral palsy and/or age-dependent dementia. The methods of the invention comprise administration of a neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a neuregulin fragment or derivative, to a patient suffering from or susceptible to such conditions.



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THERAPEUTIC METHODS COMPRISING USE OF A NEUREGULIN

BACKGROUND OF THE INVENTION

1. Field of the Invention

The present invention relates to methods for treatment of certain neurologicalrelated injuries and disorders comprising use of a neuregulin, or a fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or neuregulin fragment or derivative.

2. Background

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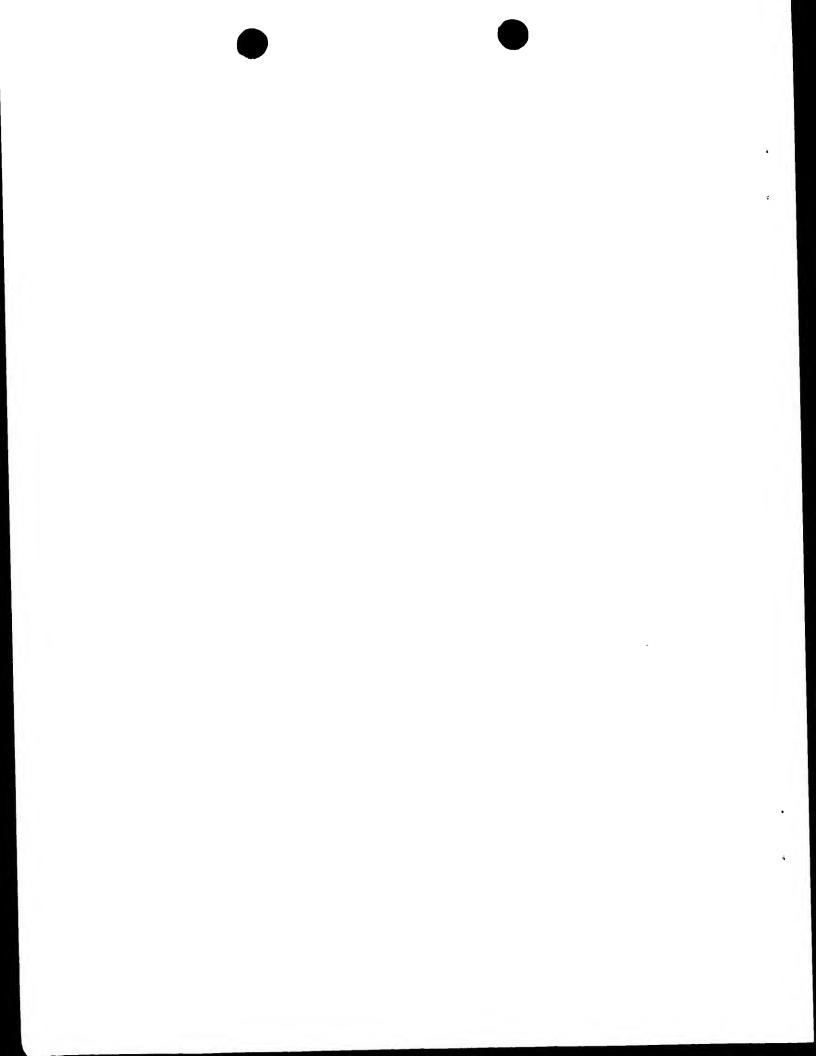
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Nerve cell death (degeneration) can cause potentially devastating and irreversible effects for an individual and may occur e.g. as a result of stroke, heart attack or other brain or spinal chord ischemia or trauma. Additionally, neurodegenerative disorders involve nerve cell death (degeneration) such as Alzheimer's disease, Parkinson's disease, Huntington's disease, Amyotrophic Lateral Sclerosis, Down's Syndrome and Korsakoff's disease.

Therapies have been investigated to treat nerve cell degeneration and related disorders, e.g., by limiting the extent of nerve cell death that may otherwise occur to an individual as well as promoting repair, remodeling and reprogramming after stroke or other neuronal injury. See, e.g., F. Seil, *Curr Opin Neuro*, 10:49-51 (1997); N. L. Reddy et al., *J Med Chem*, 37:260-267 (1994); and WO 95/20950.

Certain growth factors have been reported to exhibit neuroprotective properties. In particular, nerve growth factor (NGF) has been evaluated in certain neuroprotective models. See, for example, G. Sinson et al., *J Neurosurg*, 86(3):511-518 (1997); and G. Sinson et al., *J Neurochem*, 65(5):2209-2216 (1995). Osteogenic protein-1 (OP-1) has been evaluated in a rat model of cerebral hypoxia/ischemia for neuroprotective activity. G. Perides, *Neurosci Lett*, 1871):21-24 (1995). Glial cell line-derived neurotrophic factor (GDNF) was reported to exhibit trophic activity on certain populations of central neurons. Y. Wang et al., *J Neurosci*, 17(11):4341-4348 (1997). Small molecules also have been investigated as neuroprotective agents, such



as MK-801. See B. Meldrum, Cereb Brain Metab Rev, 2:27-57 (1990); D. Choi, Cereb Brain Metab Rev, 2:27-57 (1990).

However, no effective pharmacotherapies are in regular clinical use for ischemia-induced brain injury or other such injuries and disorders. See, for example, Y. Wang et al., *supra*; G. Sinson et al., *J Neurochem*, *J Neurochem*, 65(5):2209 (1995).

It thus would be highly desirable to have new neuroprotective agents, particularly agents to limit the extent or otherwise treat nerve cell death (degeneration) that occur with stroke, heart attack or brain or spinal cord trauma, or to treat Alzheimer's disease, Parkinson's disease, Huntington's disease, Amyotrophic Lateral Sclerosis, Down's Syndrome and Korsakoff's disease. It also would be desirable to have agents that promote repair, remodeling or reprogramming after stroke or other neuronal injury.

SUMMARY OF THE INVENTION

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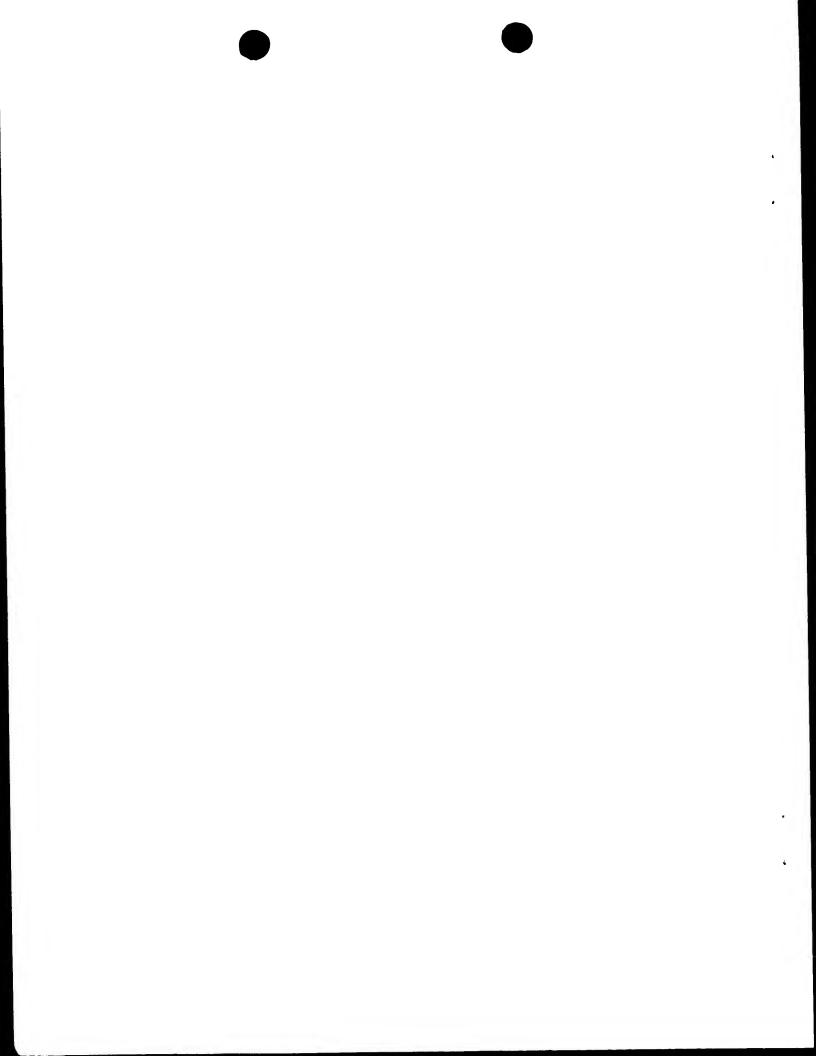
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The present invention provides methods for treatment and/or prophylaxis of certain neurological-related disorders, particularly treatment or prophylaxis of the effects of stroke, brain or spinal cord injury or ischemia, heart attack, optic nerve and retinal injury and ischemia and other acute-type conditions disclosed herein as well as chronic-type conditions, specifically epilepsy, Alzheimer's disease, Parkinson's disease, Huntington's disease, Amyotrophic Lateral Sclerosis, Down's Syndrome, Korsakoff's disease, cerebral palsy and/or age-dependent dementia. Methods of the invention also include therapies for promoting repair, remodeling or reprogramming after stroke or other neuronal injury.

The methods of the invention comprise administration of an effective amount of neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a neuregulin fragment or derivative (i.e. gene therapy), to a patient suffering from or susceptible to such conditions.

Neuregulins are members of the epidermal growth factor (EGF) superfamily and include glial growth factor (GGF), acetylcholine receptor-inducing activity (ARIA), neu differentiation factor (NDF) and heregulins (HRF). See D. E. Wen et al., Cell, 69:559-572 (1992); W.E. Holmes et al., Science, 256:1205-1210 (1992); M.A. Marchionni et al., Nature, 362:312-318 (1993); and D.L. Falls, Cell, 72:801-815



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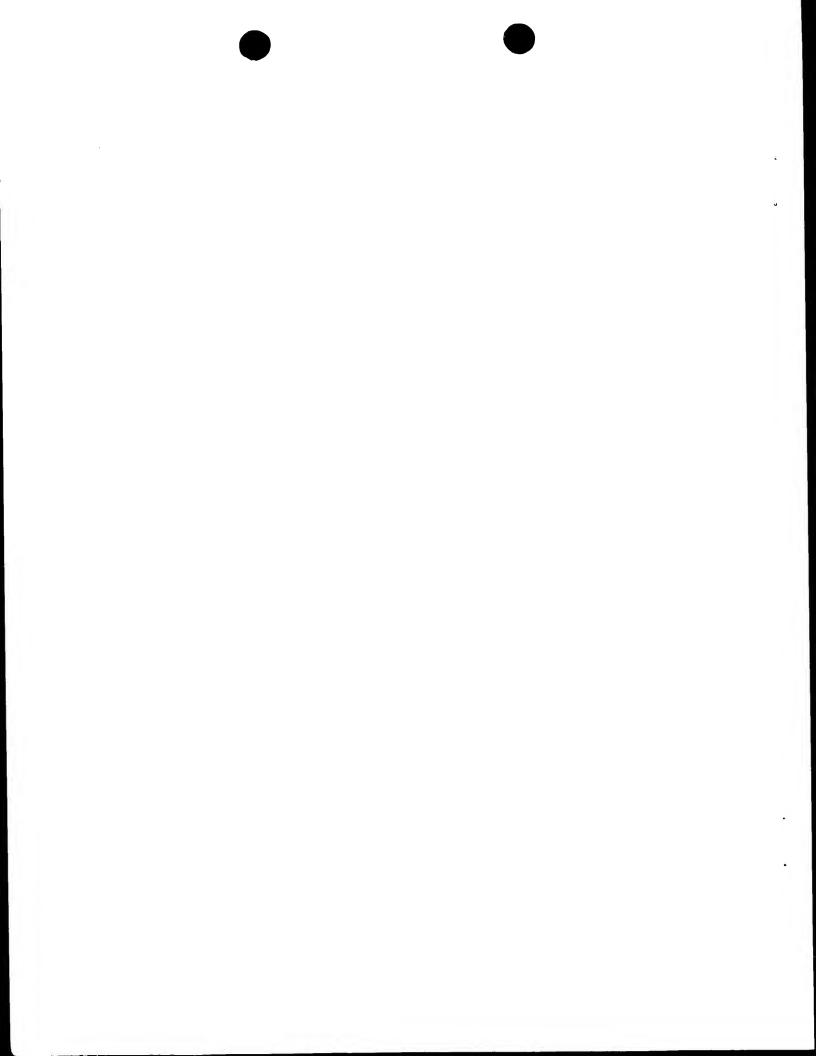
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(1993). A variety of neuregulins and fragments and derivatives thereof can be employed in the methods of the invention. For example, suitable agents have been disclosed in U.S. Patent 5,530,109 and PCT/US93/07491. Neuregulins also have been reported in U.S. Patent 5,367,060. Preferred neuregulins include regions shown in FIGS. 1-2 (SEQ ID NOS. 2 and 4), also known as the E sequence. Preferred neuregulins or fragments or derivatives also include those that contain the C, C/D or C/D' sequences as shown in Figures 7, 8 and 9 respectively of the drawings, or those neuregulins or fragments or derivatives that have substantial homology to the peptide sequences shown in Figures 7, 8 or 9, e.g. at least about 70 percent homology, or at least about 80 percent homology, or more preferably at least about 90 or 95 percent homology to the peptide sequences shown in Figures 7, 8 or 9. Preferred nucleic acids and fragments and derivatives for use in the methods of the invention include those nucleic acids that include one or more nucleic acids sequences shown in Figures 7, 8 and 9 of the drawings, or those nucleic acids that that have substantial homology to the nucleic acid sequences shown in Figures 7, 8 or 9, e.g. at least about 70, 80, 90 or 95 percent homology to the nucleic acid sequences shown in Figures 7, 8 or 9. A particularly preferred neuregulin is encoded by DNA obtainable from the clone pGGF2HBS11 (ATCC Deposit No. 75347). Also preferred are neuregulins encoded by DNA obtainable from GGF2BPP5, GGF2BPP2 and GGF2BPP4.

Typical patients that may be treated in accordance with the methods of the invention are persons suffering from brain or spinal cord trauma or ischemia, stroke, heart attack, hypoxia, hypoglycemia, post-surgical neurological deficits, decreased blood flow or nutrient supply to retinal tissue or optic nerve, retinal trauma or ischemia or optic nerve injury. Patients suffering from chronic-type conditions also may be treated in accordance with the invention, specifically subjects suffering from or susceptible to epilepsy, Parkinson's disease, Huntington's disease, Amyotrophic Lateral Sclerosis, Alzheimer's disease, Down's Syndrome, Korsakoff's disease, cerebral palsy and/or age-dependent dementia.

Also, as discussed above, a neuregulin or fragment or derivative thereof or nucleic acid encoding same, may be administered to promote repair, remodeling or reprogramming to a subject that has suffered stroke or other neuronal injury such as traumatic brain or spinal cord injury. In such cases, the therapeutic agent may be



suitably administered to the subject over an extended period following the injury, e.g. at least about 1, 2, 3, 4, 6, 8, 12 or 16 weeks following the injury.

Other aspects of the invention are disclosed infra.

BRIEF DESCRIPTION OF THE DRAWINGS

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FIG. 1 shows a nucleotide sequence (SEQ ID NO:1) encoding a preferred neuregulin region (E segment of human GGF) and the amino acid sequence (SEQ ID NO:2) of that preferred region.

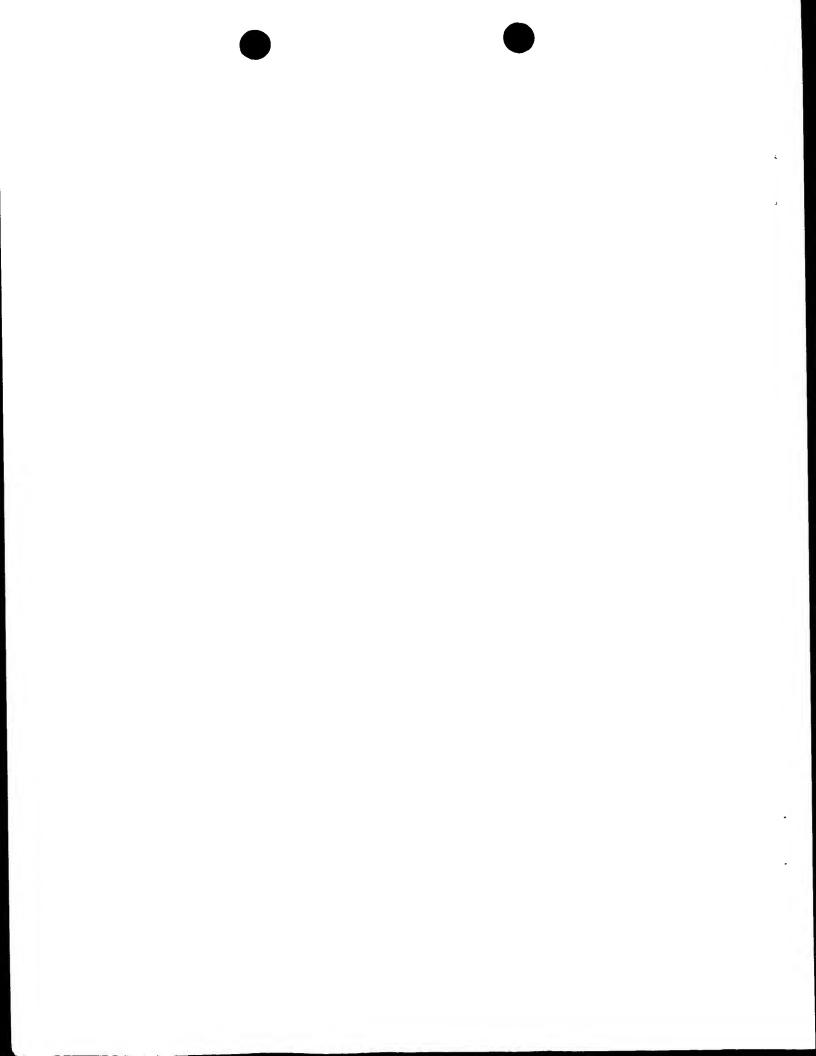
FIG. 2 shows a nucleotide sequence (SEQ ID NO:3) encoding a preferred neuregulin region (E segment of bovine GGF) and the amino acid sequence (SEQ ID NO:4) of that preferred region.

FIG. 3 shows nucleotide sequences (SEQ ID NOS:6-7) encoding further neuregulin regions (B segment of human and bovine GGF) and amino acid sequences (SEQ ID NOS:5 and 8) of those regions. Line 1 is the predicted amino acid sequence of bovine B segment, line 2 is a nucleotide sequence of bovine B segment, line 3 is a nucleotide sequence of human B segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human B segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

FIG. 4 shows nucleotide sequences (SEQ ID NOS:10-11) encoding further neuregulin regions (A segment of human and bovine GGF) and amino acid sequences (SEQ ID NOS:9 and 12) of those regions. Line 1 is the predicted amino acid sequence of bovine A segment, line 2 is a nucleotide sequence of bovine A segment, line 3 is a nucleotide sequence of human A segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human A segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

FIG. 5 shows a nucleotide sequence (SEQ ID NO:13) encoding a further neuregulin region (A' segment of bovine GGF) and the predicted amino acid sequence (SEQ ID NO:14) of that region.

FIG. 6 shows nucleotide sequences (SEQ ID NOS:16-17) encoding further neuregulin regions (G segment of bovine and human GGF) and amino acid sequences (SEQ ID NOS:15 and 18) of that region. Line 1 is the predicted amino acid sequence of bovine G segment, line 2 is a nucleotide sequence of bovine G segment, line 3 is a



nucleotide sequence of human G segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human G segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

FIG. 7 shows nucleotide sequences (SEQ ID NOS:20-21) encoding further neuregulin regions (C segment of bovine and human GGF) and amino acid sequences (SEQ ID NOS:19 and 22) of those regions. Line 1 is the predicted amino acid sequence of bovine C segment, line 2 is a nucleotide sequence of bovine C segment, line 3 is a nucleotide sequence of human C segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human C segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

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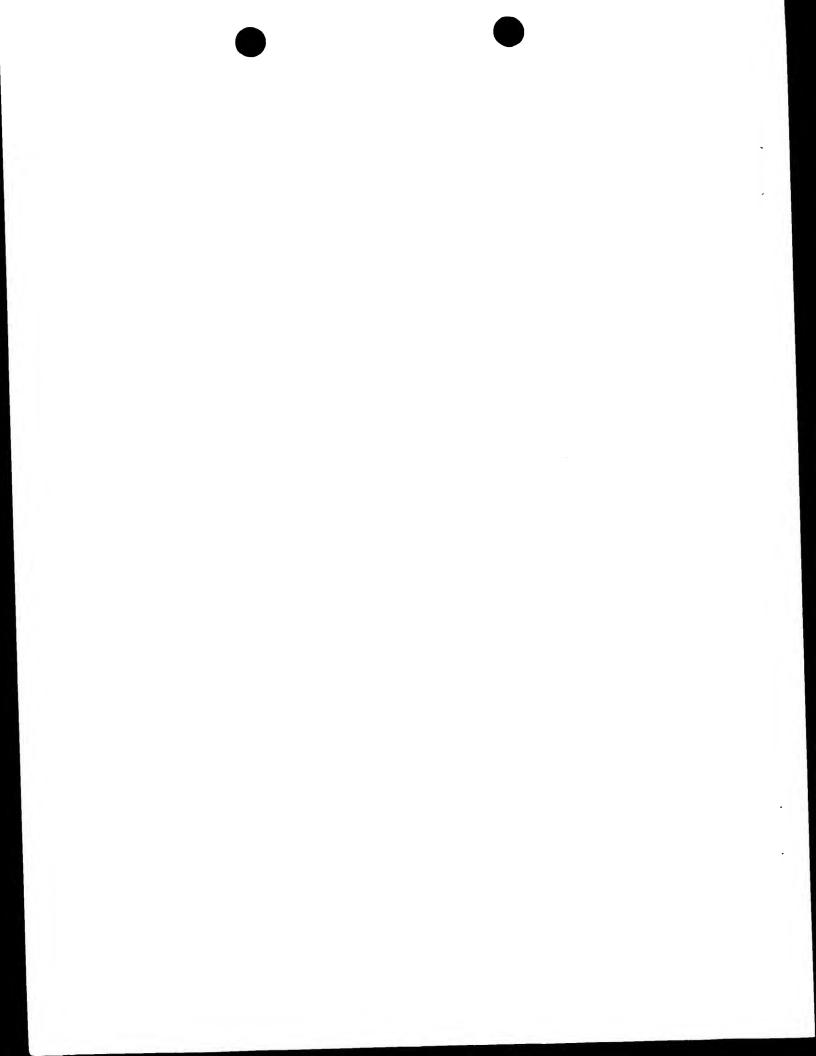
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FIG. 8 shows nucleotide sequences (SEQ ID NOS:24-25) encoding further neuregulin regions (C/D segment of human and bovine GGF) and amino acid sequences (SEQ ID NOS:23 and 26) of those regions. Line 1 is the predicted amino acid sequence of bovine C/D segment, line 2 is a nucleotide sequence of bovine C/D segment, line 3 is a nucleotide sequence of human C/D segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human C/D segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

FIG. 9 shows nucleotide sequences (SEQ ID NOS:28-29) encoding a further neuregulin region (C/D' segment of the human and bovine GGF) and the amino acid sequence (SEQ ID NO:27) of that region. Line 1 is the predicted amino acid sequence of the C/D' segment, line 2 is a nucleotide sequence of bovine C/D' segment and line 3 is a nucleotide sequence of human C/D' segment (nucleotide base matches are indicated with a vertical line).

FIG. 10 shows nucleotide sequences (SEQ ID NOS:31-32) encoding a further neuregulin region (D segment of the human and bovine GGF) and the amino acid sequence (SEQ ID NO:30) of that region. Line 1 is the predicted amino acid sequence of the D segment, line 2 is a nucleotide sequence of bovine D segment and line 3 is a nucleotide sequence of human D segment (nucleotide base matches are indicated with a vertical line).



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FIG. 11 shows nucleotide sequence (SEQ ID NO:34) encoding a further neuregulin region (D' segment of bovine GGF) and the amino acid sequence (SEQ ID NO:33) of that region.

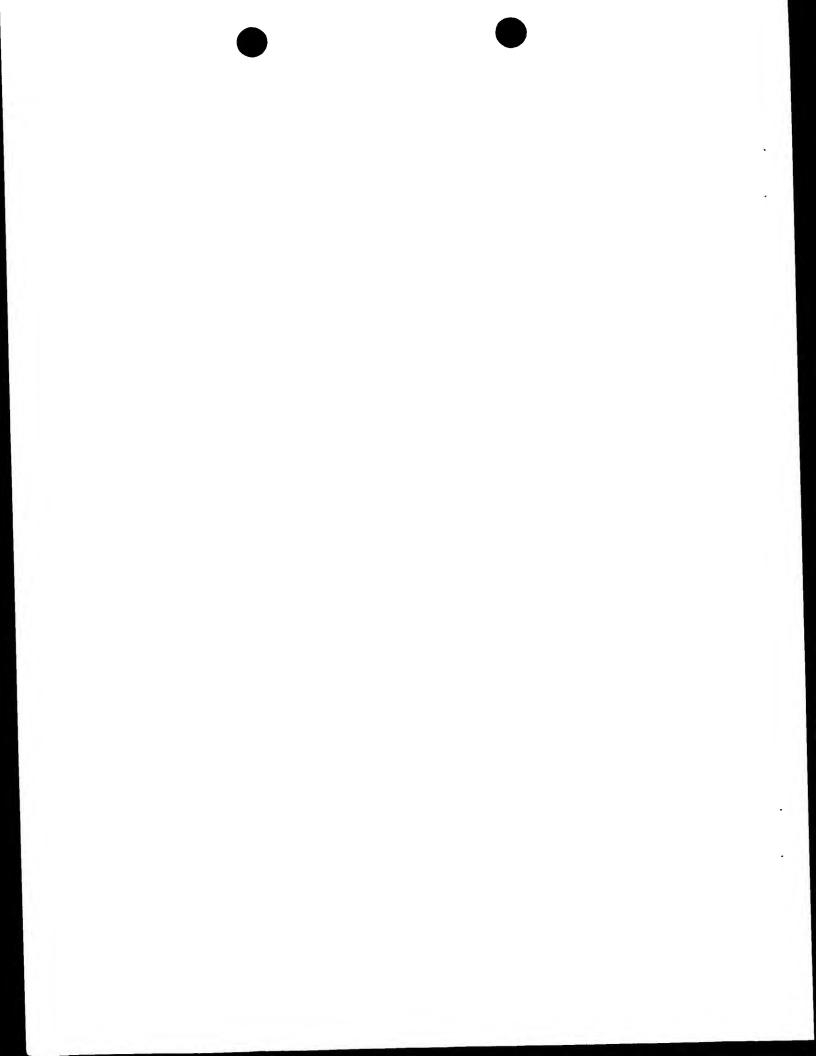
FIGS. 12A-12B show nucleotide sequences (SEQ ID NOS:36-37) encoding further neuregulin regions (H segment of human and bovine GGF) and amino acid sequences (SEQ ID NO:35 and 38) of that region. Line 1 is the predicted amino acid sequence of bovine H segment, line 2 is a nucleotide sequence of bovine H segment, line 3 is a nucleotide sequence of human H segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human H segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

FIG. 13 shows a nucleotide sequence (SEQ ID NO:40) encoding a further neuregulin region (K segment of bovine GGF) and the amino acid sequence (SEQ ID NO:39) of that region.

FIGS. 14A-14C show nucleotide sequences (SEQ ID NOS:42-43) encoding a further neuregulin region (L segment of bovine and human GGF) and amino acid sequences (SEQ ID NO:41 and 44) of that region. Line 1 is the predicted amino acid sequence of bovine L segment, line 2 is a nucleotide sequence of bovine L segment, line 3 is a nucleotide sequence of human L segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human L segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

FIG. 15 shows nucleotide sequences (SEQ ID NOS:46-47) encoding further neuregulin regions (F segment of bovine and human GGF) and amino acid sequences (SEQ ID NOS:45 and 48) of that region. Line 1 is the predicted amino acid sequence of bovine F segment, line 2 is a nucleotide sequence of bovine F segment, line 3 is a nucleotide sequence of human F segment (nucleotide base matches are indicated with a vertical line), and line 4 is the predicted amino acid sequence of human F segment shown where it differs from the bovine sequence set forth in line 1 of the figure.

FIGS. 16A-16C show the nucleotide sequence (SEQ ID NO:49) and deduced amino acid sequence (SEQ ID NO:50) of GGF2BPP4.



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FIGS. 17A-17B show the nucleotide sequence (SEQ ID NO:51) and deduced amino acid sequence (SEQ ID NO:52) of GGF2BPP2.

FIGS. 18A-18B show the nucleotide sequence (SEQ ID NO:53) and deduced amino acid sequence (SEQ ID NO:54) of GGF2BPP5.

5 DETAILED DESCRIPTION OF THE INVENTION

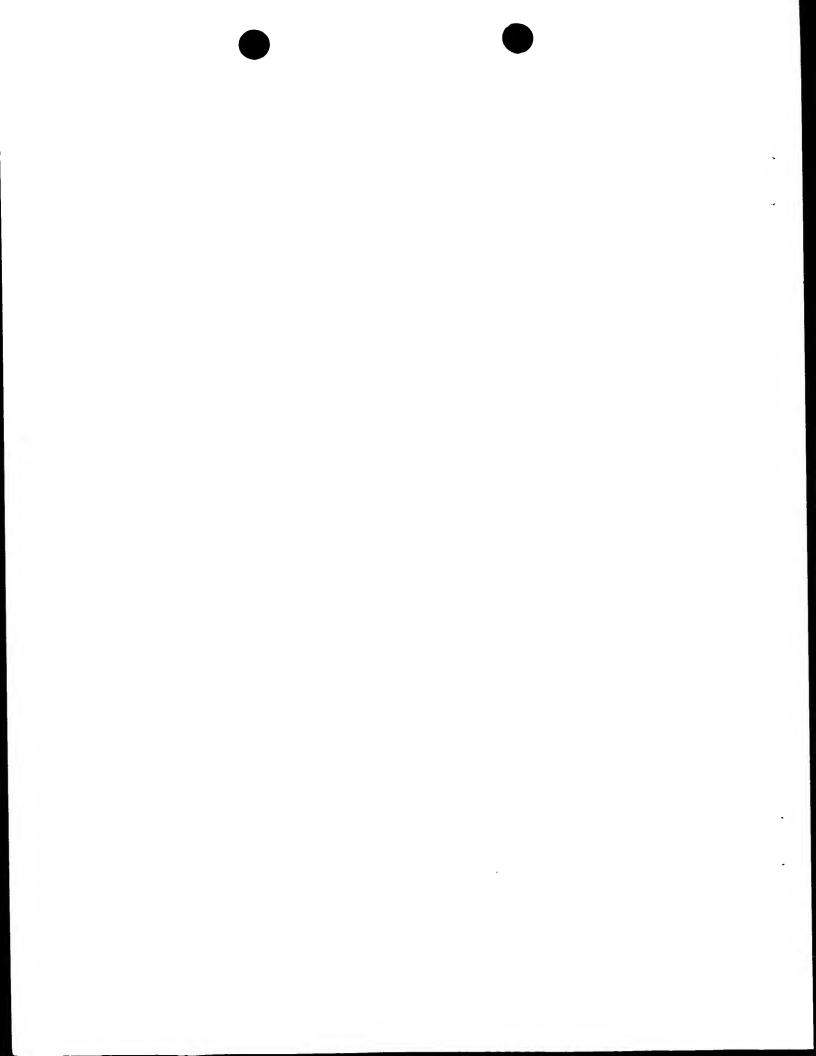
As discussed above, preferred neuregulins for use in the therapeutic methods of the present invention include those disclosed in U.S. Patent 5,530,109 and PCT/US93/07491, incorporated herein by reference. Particularly preferred neuregulins comprise an amino acid sequence of the following formula:

WYBAZCX

wherein WYBAZCX is composed of amino acid sequences that include one or more sequences shown in FIGS. 1 through 15 (which includes SEQ ID NOS:2, 4, 5, 8, 9, 12, 14, 15, 18, 19, 22, 23, 26, 27, 30, 33, 35, 38, 39, 41, 44, 45 and 48), wherein W comprises the polypeptide segment F, or is absent; wherein Y comprises the polypeptide segment E, or is absent; wherein Z comprises the polypeptide segment G or is absent; and wherein X comprise a polypeptide segment selected from the group consisting of C/D HKL, C/D H, C/D HL, C/D D, C/D' HL, C/D' HKL, C/D' H, C/D' D, C/D C/D' HKL, C/D C/D' H, C/D C/D' HL, C/D D' HKL, C/D C/D' D' HKL, And preferably that either

- a) at least one of F, Y, B, A, Z, C or X is of bovine origin; or
- b) Y comprises the polypeptide segment E; or
- c) X comprises the polypeptide segments C/D HKL, C/D D, C/D' HKL,
 C/D C/D' HKL, C/D C/D' D, C/D D' H, C/D D' HL, C/D D' HKL, C/D' D' H, C/D'
 25 D' HKL, C/D C/D' D'H, C/D C/D' D HL, C/D C/D' D' HKL, C/D'H, C/D C/D' H or
 C/D C/D' HL.

Particularly preferred neuregulins also include those polypeptides that include the segments FB polypeptides that include the segments FBA' (i.e. the groups F, B and A' as defined herein including in the drawings); polypeptides that include the segments EBA (i.e. the groups E, B and A as defined herein including in the drawings); polypeptides that include the segments EBA' (i.e. the groups E, B and A' as defined herein including in the drawings); A (i.e. the group A as defined herein



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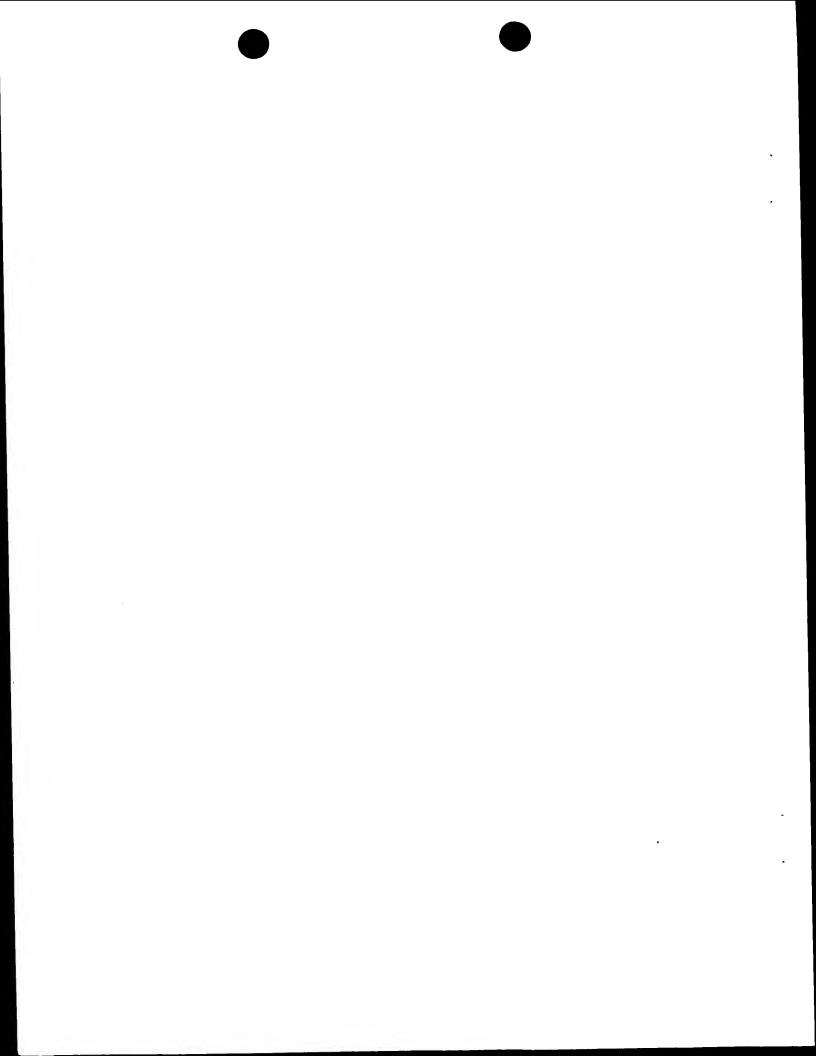
including in the drawings); polypeptides that include the segments FEBA (i.e. the groups F, E, B and A as defined herein including in the drawings); polypeptides that include the segments FBA' (i.e. the groups F, B and A' as defined herein including in the drawings); and polypeptides that include the segments FEBA' (i.e. the groups F, E, B and A' as defined herein including in the drawings).

Also preferred are nucleic acids that code for the above preferred polypeptides.

A "fragment" or "derivative" of a neuregulin refers to herein 1) a peptide in which one or more amino acid residues are with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) a peptide in which one or more of the amino acid residues includes a substituent group, or (iii) a peptide in which the mature protein is fused with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol). Thus, a fragment or derivative for use in accordance with the methods of the invention includes a proprotein, which can be activated by cleavage of the proprotein portion to produce an active mature polypeptide.

The polypeptide fragments and derivatives of the invention are of a sufficient length to uniquely identify a region of a neuregulin. Neuregulin fragments thus preferably comprise at least 8 amino acids, usually at least about 12 amino acids, more usually at least about 15 amino acids, still more typically at least about 30 amino acids, even more typically at least about 50 or 70 amino acids. Preferred fragments or derivatives for use in the methods of the invention include those that have at least about 70 percent homology (sequence identity) to any of the preferred sequences mentioned above, more preferably about 80 percent or more homology to any of the preferred sequences mentioned above, still more preferably about 85 to 90 percent or more homology to any of the preferred sequences mentioned above. Sequence identity or homology with respect to a neuregulin as referred to herein is the percentage of amino acid sequences of a neuregulin protein or fragment or derivative thereof that are identical with a specified sequence, after introducing any gaps necessary to achieve the maximum percent homology.

The neuregulin fragments and derivatives for use in the methods of the invention preferably exhibit good activity in standard neuroprotective assays such as



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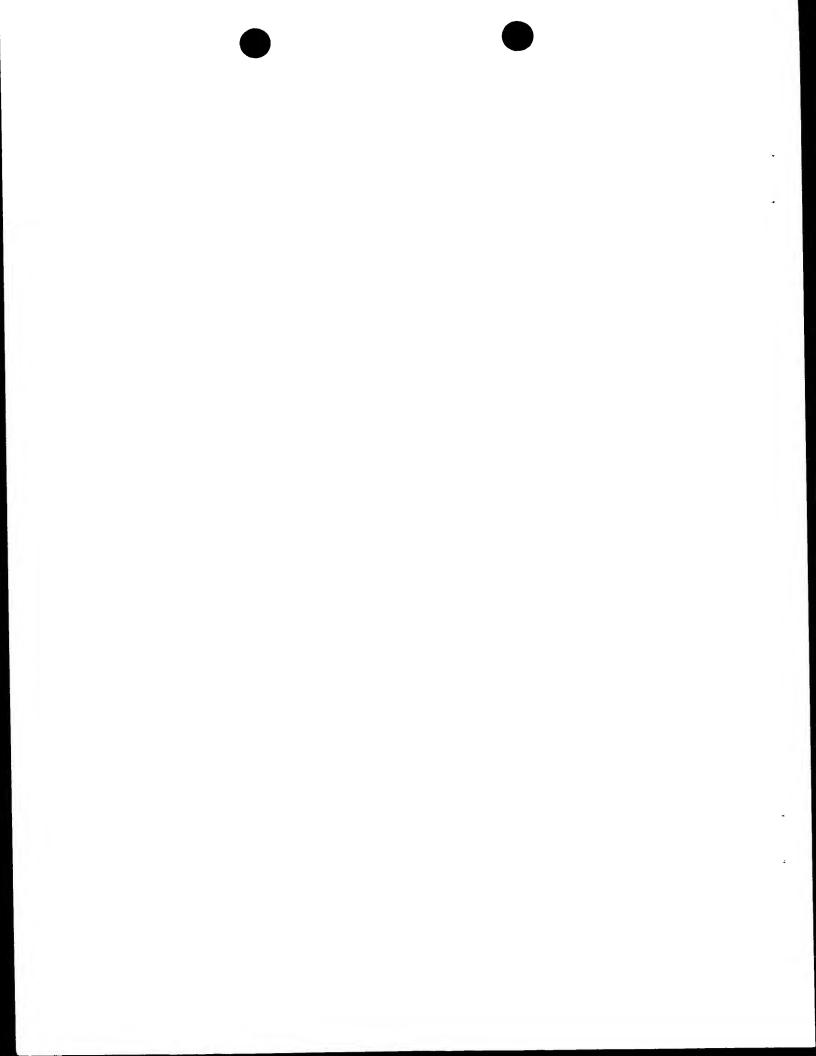
the *in vivo* cerebral ischemia assay of Example 1, which follows. That assay includes the following steps: a) continuous intraventricular infusion of the protein fragment or derivative or vehicle alone to test rats for three days prior to inducing focal ischemic infarcts in right lateral cerebral cortex; and b) twenty-four hours after inducing ischemic infarcts, infarct volume in each test animal is determined by image analysis. Preferably, a protein fragment or derivative of the invention provides at least about a 10% reduction in infarct volume relative to vehicle-treated animals, more preferably about a 20% reduction in infarct volume, still more preferably about a 25% reduction in infarct volume relative to vehicle-treated animals in such an assay. References herein to *in vivo* cerebral ischemia assay are intended to refer to an assay of the above

steps a) and b), which are more fully described in Example 1 which follows.

As discussed above, neuregulin nucleic acid fragments and derivatives are also provided for use in the methods of the invention. Those fragments and derivatives typically are of a length sufficient to bind to a sequence of any of the nucleic acid sequences shown in Figures 1-15 of the drawings, including SEQ ID NOS:1, 3, 6, 7, 10, 11, 13, 16, 17, 20, 21, 24, 25, 28, 29, 31, 32, 34, 36, 37, 40, 42 and 43 under the following moderately stringent conditions (referred to herein as "normal stringency" conditions): use of a hybridization buffer comprising 20% formamide in 0.8M saline/0.08M sodium citrate (SSC) buffer at a temperature of 37°C and remaining bound when subject to washing once with that SSC buffer at 37°C.

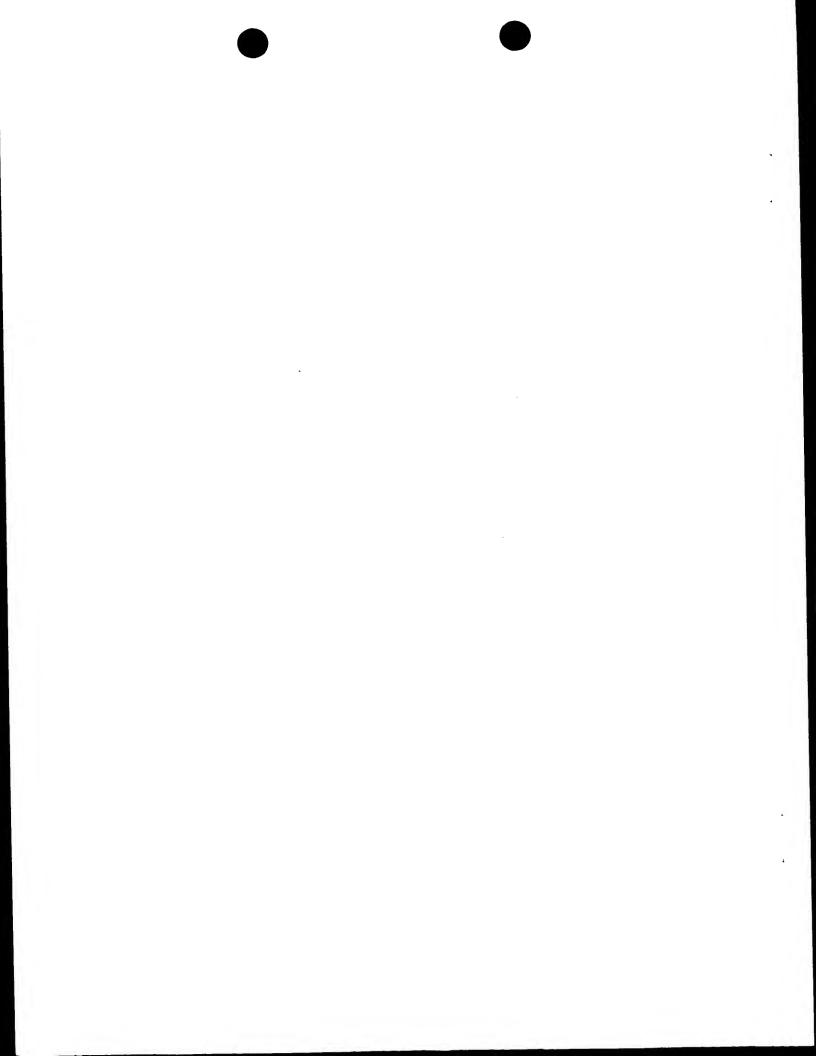
Preferred neuregulin nucleic acid fragments and derivatives of the invention will bind to a sequence of any of the nucleic acid sequences shown in Figures 1-15 of the drawings, including SEQ ID NOS:1, 3, 6, 7, 10, 11, 13, 16, 17, 20, 21, 24, 25, 28, 29, 31, 32, 34, 36, 37, 40, 42 and 43 under the following highly stringent conditions (referred to herein as "high stringency" conditions): use of a hybridization buffer comprising 20% formamide in 0.9M saline/0.09M sodium citrate (SSC) buffer at a temperature of 42°C and remaining bound when subject to washing twice with that SSC buffer at 42°C.

The neuregulin nucleic acid fragments and derivatives preferably should comprise at least 20 base pairs, more preferably at least about 50 base pairs, and still more preferably a nucleic acid fragment or derivative of the invention comprises at least about 100, 200, 300 or 400 base pairs. In some preferred embodiments, the



nucleic acid fragment or derivative is bound to some moiety which permits ready identification such as a radionucleotide, fluorescent or other chemical identifier.

Isolated neuregulin and peptide fragments or derivatives of the invention are preferably produced by recombinant methods, although suitable neuregulins also can 5 be isolated from various sources. See the procedures disclosed U.S. Patent 5,530,109; U.S. Patent 5,367,060; and PCT/US93/07491, incorporated herein by reference. A wide variety of molecular and biochemical methods are available for generating and expressing neuregulin; see e.g. the procedures disclosed in Molecular Cloning, A Laboratory Manual (2nd Ed., Sambrook, Fritsch and Maniatis, Cold Spring Harbor), 10 Current Protocols in Molecular Biology (Eds. Aufubel, Brent, Kingston, More, Feidman, Smith and Stuhl, Greene Publ. Assoc., Wiley-Interscience, NY, N.Y. 1992) or other procedures that are otherwise known in the art. For example, neuregulin or fragments or derivatives thereof may be obtained by chemical synthesis, or more preferably by expression in bacteria such as E coli and eukaryotes such as yeast, 15 baculovirus, or mammalian cell-based expression systems, etc., depending on the size, nature and quantity of neuregulin or fragment or derivative thereof. More particularly, a recombinant DNA molecule comprising a vector and a DNA segment encoding neuregulin, or a fragment or derivative thereof, can be constructed. Suitable vectors include e.g. baculovirus-derived vectors for expression in insect cells (see 20 Pennock et al., Mol. Cell. Biol., 4:399-406 (1984)), T7-based expression vector for expression in bacteria (see Rosenberg et al., Gene, 56:125-135 (1987)) and the pMSXND expression vector for expression in mammalian cells (Lee and Nathans, J. Biol. Chem., 263:3521-3527 (1988)). The DNA segment can be present in the vector operably linked to regulatory elements, e.g., a promoter (e.g., polyhedron, T7 or 25 metallothionein (Mt-I) promoters), or a leader sequence to provide for secretory expression of the polypeptide. The recombinant DNA molecule containing the DNA coding for a neuregulin or a fragment or derivative thereof can be introduced into appropriate host cells by known methods. Suitable host cells include e.g. prokaryotes such as E. coli, Bacillus subtilus, etc., and eukaryote such as animal cells and yeast 30 strains, e.g., S. cerevisiae. Mammalian cells may be preferred such as J558, NSO, SP2-O or CHO. In general, conventional culturing conditions can be employed. See Sambrook, supra. Stable transformed or transfected cell lines can then be selected.



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The expressed neuregulin or fragment or derivative thereof then can be isolated and purified by known methods. Typically the culture medium is centrifuged and the supernatant purified by affinity or immunoaffinity chromatography, e.g. Protein-A or Protein-G affinity chromatography or an immunoaffinity protocol comprising use of monoclonal antibodies that bind neuregulins.

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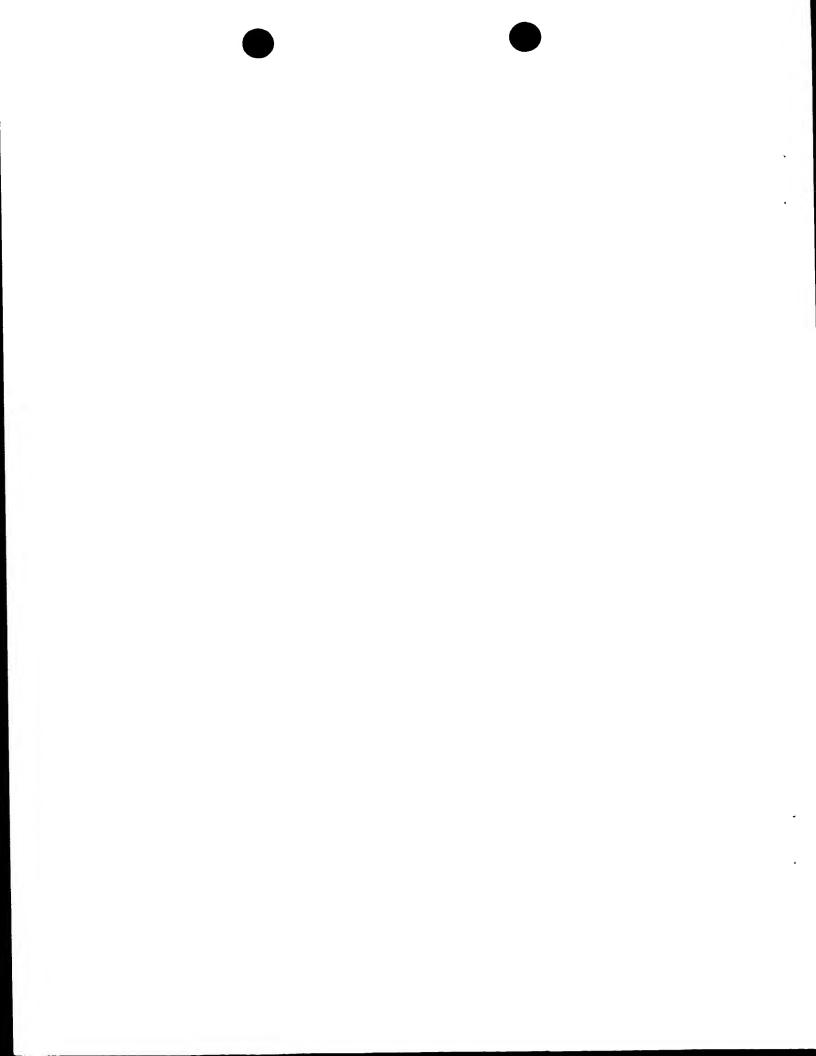
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Neuregulin nucleic acids used in the methods of the invention are typically isolated, meaning the nucleic acids comprise a sequence joined to a nucleotide other than that which it is joined to on a natural chromosome and usually constitute at least about 0.5%, preferably at least about 2%, and more preferably at least about 5% by weight of total nucleic acid present in a given fraction. A partially pure nucleic acid constitutes at least about 10%, preferably at least about 30%, and more preferably at least about 60% by weight of total nucleic acid present in a given fraction. A pure nucleic acid constitutes at least about 80%, preferably at least about 90%, and more preferably at least about 95% by weight of total nucleic acid present in a given fraction.

As discussed above, the present invention includes methods for treating and preventing certain neurological-related injuries and disorders, comprising the administration of an effective amount of a neuregulin or fragment or derivative thereof, or nucleic acid encoding same, to a subject including a mammal, particularly a human, in need of such treatment.

In particular, the invention provides methods for treatment and/or prophylaxis of nerve cell death (degeneration) resulting from hypoxia, hypoglycemia, brain or spinal cord ischemia, brain or spinal cord trauma, stroke, heart attack or drowning. Typical candidates for treatment include e.g. heart attack, stroke and/or persons suffering from cardiac arrest neurological deficits, brain or spinal cord injury patients, patients undergoing major surgery such as heart surgery where brain ischemia is a potential complication and patients such as divers suffering from decompression sickness due to gas emboli in the blood stream. Candidates for treatment also will include those patients undergoing a surgical procedure involving extra-corporal circulation such as e.g. a bypass procedure.

The invention also provides methods for treatment which comprise administration of a neuregulin or fragment or derivative thereof, or nucleic acid



encoding same, to a patient that is undergoing surgery or other procedure where brain or spinal cord ischemia is a potential risk. For example, carotid endarterectomy is a surgical procedure employed to correct atherosclerosis of the carotid arteries. Major risks associated with the procedure include intraoperative embolization and the danger of hypertension in the brain following increased cerebral blood flow, which may result in aneurysm or hemorrhage. Thus, an effective amount of a neuregulin or fragment or derivative thereof, or nucleic acid encoding same, could be administered pre-operatively or peri-operatively to reduce such risks associated with carotid endarterectomy, or other post-surgical neurological deficits.

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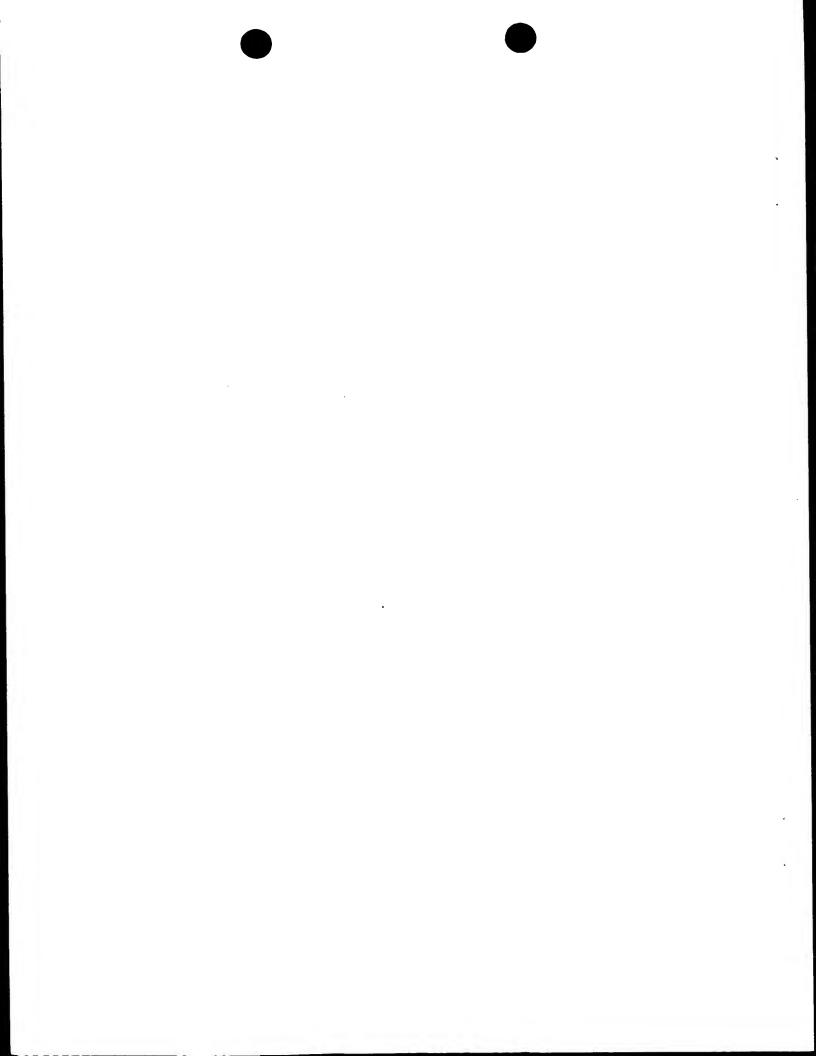
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The invention also is effective to promote and enhance recovery from acute nerve cell death and neurological conditions. Thus, for example, a neuregulin or fragment or derivative thereof, or nucleic acid encoding same, could be administered to promote repair, remodeling or reprogramming to a patient that has suffered from stroke or other neuronal injury, suitably for an extended period as discussed above. A therapeutic agent of the invention also could be administered post-operatively to promote recovery from any neurological deficits that may have occurred to a patient that has undergone surgery.

The invention further includes methods for prophylaxis against neurological deficits resulting from e.g. coronary artery bypass graft surgery and aortic valve replacement surgery, or other procedure involving extra-corporal circulation. Those methods will comprise administering to a patient undergoing such surgical procedures an effective amount of a neuregulin or fragment or derivative thereof, or nucleic acid encoding same, typically either pre-operatively or peri-operatively.

The invention also provides methods for prophylaxis and treatment against neurological injury for patients undergoing myocardial infarction, a procedure that can result in ischemic insult to the patient. Such methods will comprise administering to a patient undergoing such surgical procedure an effective amount of a neuregulin or fragment or derivative thereof, or nucleic acid encoding same, typically either preoperatively or peri-operatively.

Also provided are methods for treating or preventing neuropathic pain such as may be experienced by cancer patients, persons having diabetes, amputees and other persons who may experience neuropathic pain. These methods for treatment comprise



administration of an effective amount of a neuregulin or fragment or derivative thereof, or nucleic acid encoding same, to a patient in need of such treatment.

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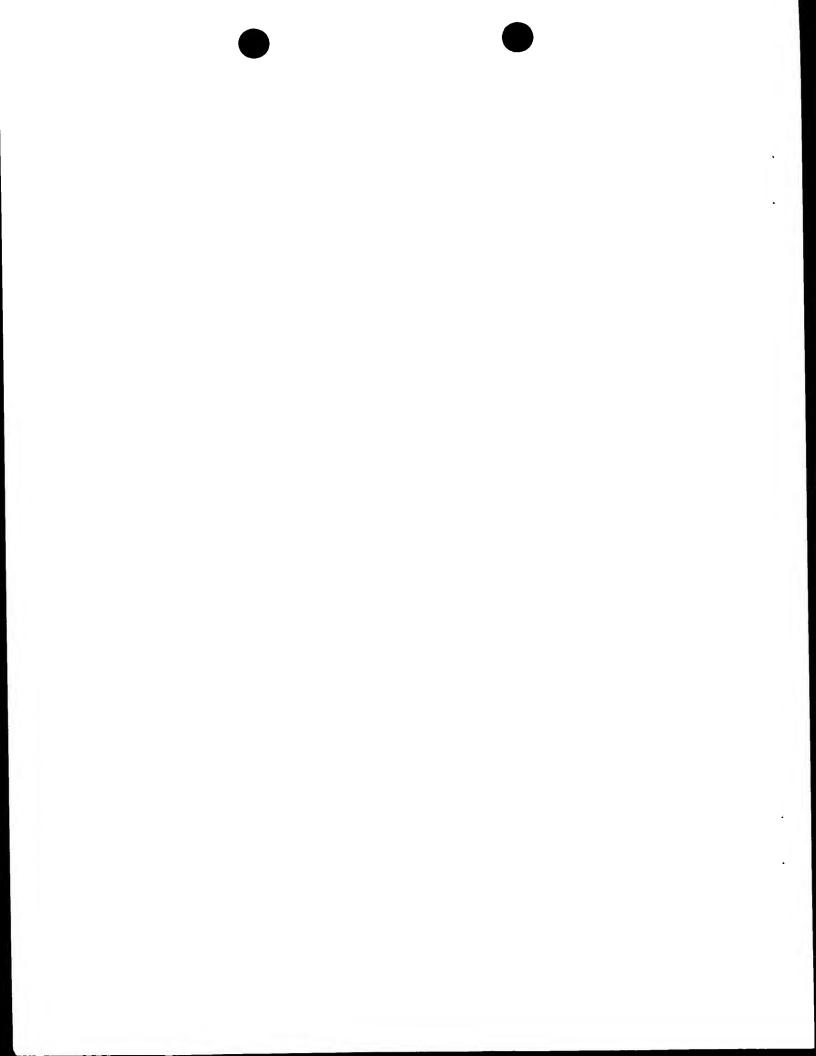
The invention also provides methods for treatment and prophylaxis against retinal ischemia or degeneration and resulting visual loss. For example, a neuregulin or fragment or derivative thereof, can be administered parenterally or by other procedure as described herein to a subject a suffering from or susceptible to ischemic insult that may adversely affect retinal function, e.g., significantly elevated intraocular pressures, diseases such as retinal artery or vein occlusion, diabetes or other ischemic ocular-related diseases. Post-ischemic administration also may limit retinal damage. The invention also includes methods for treating and prophylaxis against decreased blood flow or nutrient supply to retinal tissue or optic nerve, or treatment or prophylaxis against retinal trauma or optic nerve injury. Subjects for treatment according to such therapeutic methods of the invention may be suffering or susceptible to retinal ischemia that is associated with atherosclerosis, venous capillary insufficiency, obstructive arterial or venous retinopathies, senile macular degeneration, cystoid macular edema or glaucoma, or the retinal ischemia may be associated with a tumor or injury to the mammal. Intravitreal injection also may be a preferred administration route to provide more direct treatment to the ischemic retina.

The invention further provides a method of treating Korsakoff's disease, a chronic alcoholism-induced condition, comprising administering to a subject including a mammal, particularly a human, an effective amount of a neuregulin or fragment or derivative thereof, in an amount effective to treat the disease.

Compounds of the invention are anticipated to have utility for the attenuation of cell loss, hemorrhages and/or amino acid changes associated with Korsakoff's disease.

The invention further includes methods for treating a person suffering from or susceptible to epilepsy, emesis, narcotic withdrawal symptoms and age-dependent dementia, comprising administering to a subject including a mammal, particularly a human, an effective amount of a neuregulin or fragment or derivative thereof, in an amount effective to treat the condition.

It will be appreciated that in some instances a neuregulin or a fragment or derivative thereof will be preferably administered to a subject rather than a neuregulin nucleic acid, particularly where a patient is suffering from or susceptible to an acute



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neurological injury that demands immediate therapy. For example, administration of a neuregulin polypeptide may be preferred to a patient suffering from stroke, heart attack, traumatic brain injury and the like where it is desired to deliver the active therapeutic as quickly as possible.

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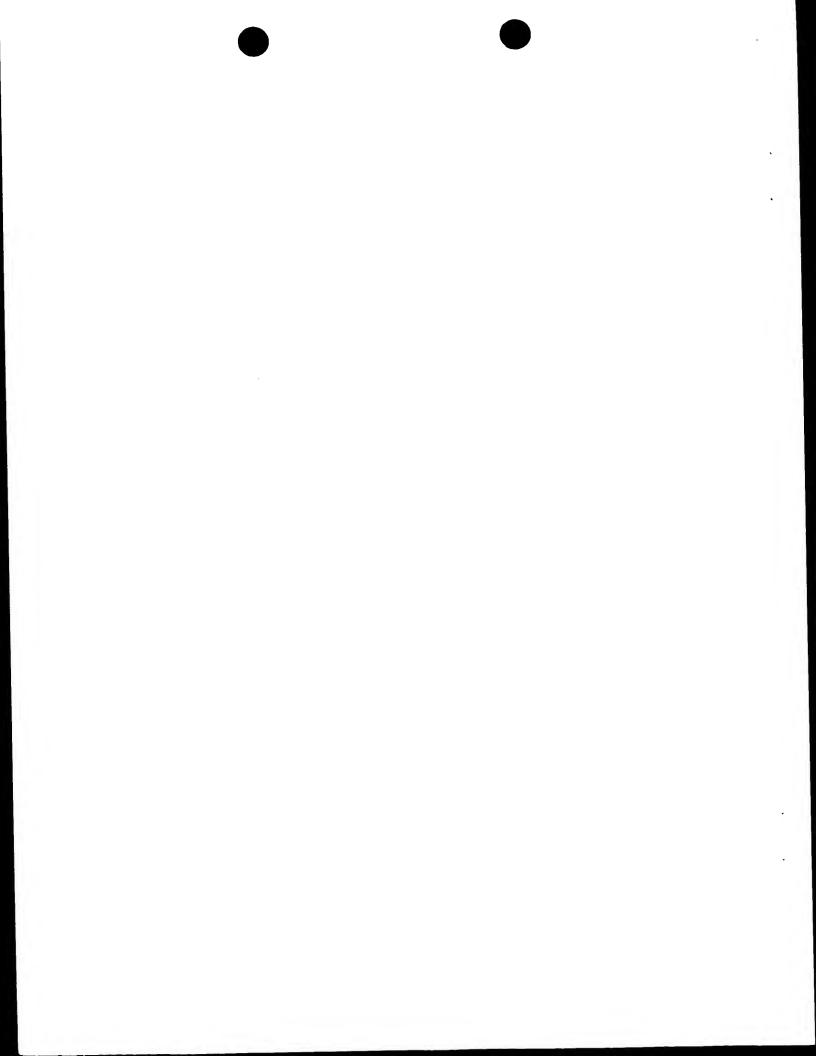
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In the therapeutic methods of the invention, neuregulin peptides and nucleic acids may be suitably administered to a subject such as a mammal, particularly a human, by any of a number of routes including parenteral (including subcutaneous, intramuscular, intravenous and intradermal), oral, rectal, nasal, vaginal and optical (including buccal and sublingual) administration. A neuregulin protein or nucleic acid or fragment or derivative thereof may be administered to a subject alone or as part of a pharmaceutical composition, comprising the peptide or nucleic acid together with one or more acceptable carriers and optionally other therapeutic ingredients. The carriers should be "acceptable" in the sense of being compatible with the other ingredients of the formulation and not deleterious to the recipient thereof.

Nucleic acids encoding a neuregulin or a neuregulin fragment or derivative can be administered to a patient by generally known gene therapy procedures. See, for example, WO 90/11092 and WO 93/00051. Thus, for instance, the nucleic acids may be introduced into target cells by any method which will result in the uptake and expression of the nucleic acid by the target cells. These methods can include vectors, liposomes, naked DNA, adjuvant-assisted DNA, catheters, etc. Preferably, the administered nucleic acid codes for an appropriate secretory sequence to promote expression upon administration. Suitable vectors for administering a nucleic acid in accordance with the invention include chemical conjugates such as described in WO 93/04701, which has targeting moiety (e.g. a ligand to a cellular surface receptor), and a nucleic acid binding moiety (e.g. polylysine), viral vector (e.g. a DNA or RNA viral vector), fusion proteins such as described in PCT/US 95/02140 (WO 95/22618) which is a fusion protein containing a target moiety (e.g. an antibody specific for a target cell) and a nucleic acid binding moiety (e.g. a protamine), plasmids, phage, etc. The vectors can be chromosomal, non-chromosomal or synthetic.

Preferred vectors include viral vectors, fusion proteins and chemical conjugates. Retroviral vectors include moloney murine leukemia viruses. DNA viral vectors are preferred. These vectors include pox vectors such as orthopox or avipox



vectors, herpes virus vectors such as a herpes simplex I virus (HSV) vector [A.I. Geller et al., J. Neurochem, 64:487 (1995); F. Lim et al., in DNA Cloning:

Mammalian Systems, D. Glover, Ed. (Oxford Univ. Press, Oxford England) (1995);

A.I. Geller et al., Proc Natl. Acad. Sci. U.S.A.:90 7603 (1993); A.I. Geller et al., Proc Natl. Acad. Sci USA, 87:1149 (1990)], Adenovirus Vectors [LeGal LaSalle et al., Science, 259:988 (1993); Davidson, et al., Nat. Genet., 3:219 (1993); Yang et al., J. Virol., 69:2004 (1995)] and Adeno-associated Virus Vectors [Kaplitt, M.G., et al., Nat. Genet., 8:148 (1994)].

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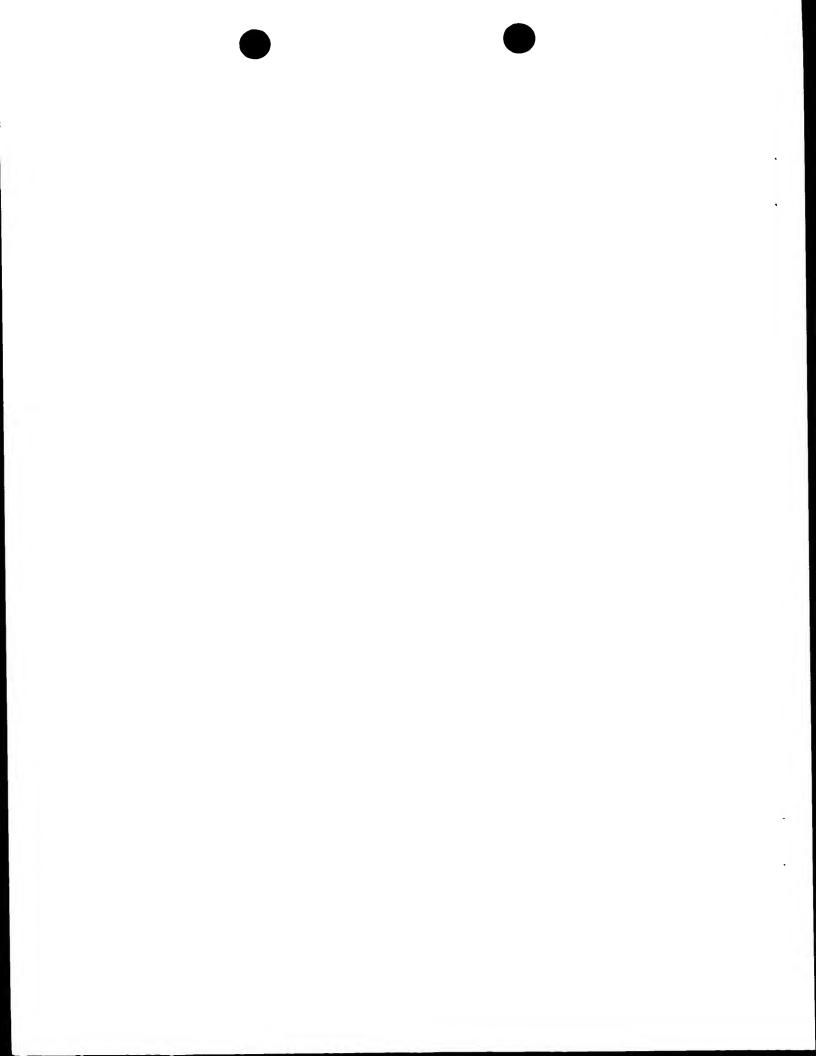
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Pox viral vectors introduce the gene into the cell cytoplasm. Avipox virus vectors result in only a short-term expression of the nucleic acid. Adenovirus vectors, adeno-associated virus vectors and herpes simplex virus (HSV) vectors are preferred for introducing the nucleic acid into neural cells. The adenovirus vector results in a shorter term expression (about 2 months) than adeno-associated virus (about 4 months), which in turn is shorter than HSV vectors. The particular vector chosen will depend upon the target cell and the specific condition being treated. The introduction can be by standard techniques, e.g. infection, transfection, transduction or transformation. Examples of modes of gene transfer include e.g., naked DNA, Ca₃(PO₄)₂ precipitation, DEAE dextran, electroporation, protoplast fusion, lipofecton, cell microinjection, and viral vectors.

A vector can be employed to target essentially any desired target cell. For example, stereotaxic injection can be used to direct the vectors (e.g. adenovirus, HSV) to a desired location. Additionally, the particles can be delivered by intracerebroventricular (icv) infusion using a minipump infusion system, such as a SynchroMed Infusion System. A method based on bulk flow, termed convection, has also proven effective at delivering large molecules to extended areas of the brain and may be useful in delivering the vector to the target cell (Bobo et al., *Proc. Natl. Acad. Sci. USA*, 91:2076-2080 (1994); Morrison et al., *Am. J. Physiol.*, 266:292-305 (1994)). Other methods that can be used include catheters, intravenous, parenteral, intraperitoneal and subcutaneous injection, and oral or other known routes of administration.

Parenteral formulations for administration of a neuregulin or a fragment or derivative thereof may be in the form of liquid solutions or suspensions; for oral



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administration, formulations may be in the form of tablets or capsules; and for intranasal formulations, in the form of powders, nasal drops, or aerosols.

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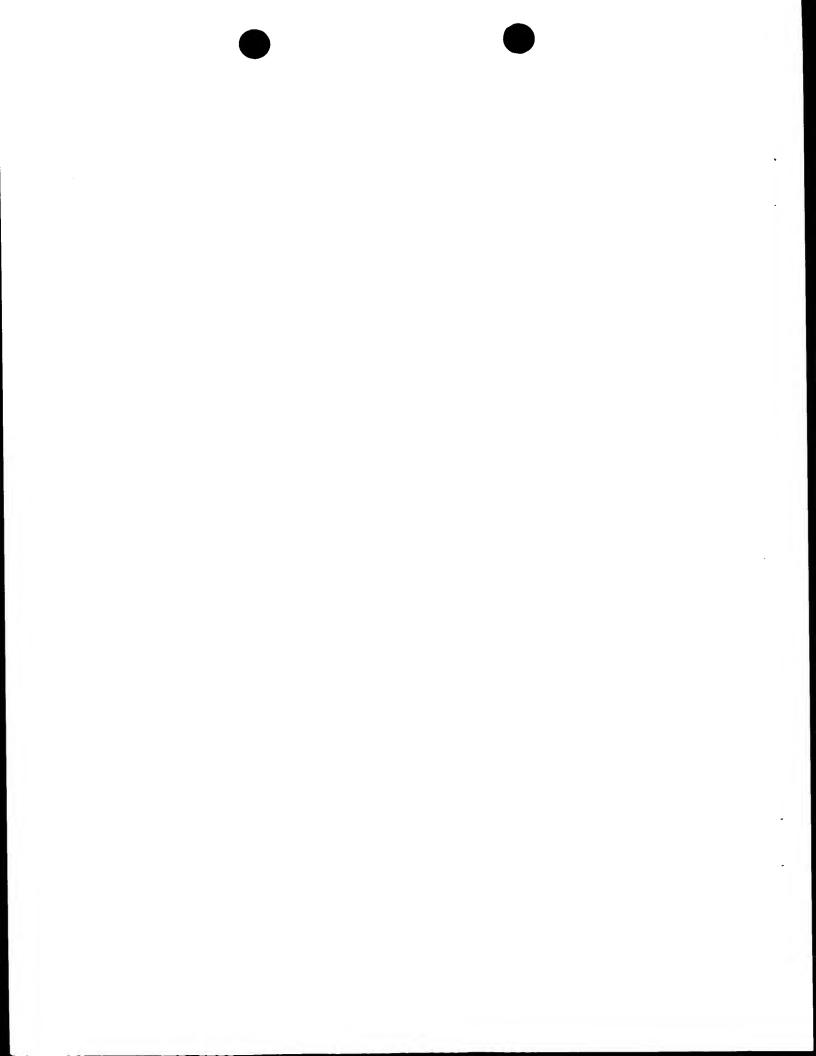
Methods well known in the art for making formulations are found in, for example, "Remington's Pharmaceutical Sciences". Formulations for parenteral administration may, for example, contain as excipients sterile water or saline, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin, or hydrogenated naphthalenes, biocompatible, biodegradable lactide polymer, or polyoxyethylene-polyoxypropylene copolymers may be used to control the release of the present factors. Other potentially useful parenteral delivery systems for a neuregulin or fragments or derivatives thereof include ethylene-vinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes. Formulations for inhalation may contain as excipients, for example, lactose, or may be aqueous solutions containing, for example, polyoxyethylene-9-lauryl ether, glycocholate and deoxycholate, or may be oily solutions for administration in the form of nasal drops, or as a gel to be applied intranasally. Formulations for parenteral administration may also include glycocholate for buccal administration, methoxysalicylate for rectal administration, or citric acid for vaginal administration.

The concentration of a neuregulin or a fragment or derivative thereof, or nucleic acid encoding such polypeptides, administered to a particular subject will vary depending upon a number of issues, including the condition being treated, the mode and site of administration, the age, weight sex and general health of the subject, and other such factors that are recognized by those skilled in the art. Optimal administration rates for a given protocol of administration can be readily determined by those skilled in the art.

All documents mentioned herein are incorporated herein by reference in their entirety. The invention is further illustrated by the following non-limiting Examples. Example 1 -- In vivo neuroprotection assay

Neuregulins and neuregulin fragments and derivatives can be assessed for neuroprotective efficacy pursuant to the following assay.

Mature male Long-Evans rats (Charles River, 250-350g) are allowed food and water *ad libitum*. Animals are anesthetized with sodium pentobarbital (60 mg/kg, i.p.) and placed in a stereotaxic head holder (David Kopf Instruments, Tujunga, CA). The



dorsal surface of the skull is exposed by midline incision, and a small burr hole (2 mm diameter) is drilled over the right lateral ventricle, 1.6 mm lateral and 0.9 mm posterior to bregma. A stainless steel cannula (I.D. 0.020", O.D. 0.028", 2 cm long) is then inserted stereotaxically into the ventricle to a depth of 4.4 mm beneath the surface of the skull. The tubing is suitably bent at a 90° angle 1-1.6 cm from its tip and connected to polyethylene tubing (I.D. 0.76 mm, O.D. 1.22 mm, 10 cm long) that is connected (by glue) to a mini-osmotic pump (Alzet 1007D, 100 µl fill volume, pump rate = 0.5 µl/hr; Alza Corp., Palo Alto, CA) implanted subcutaneously in the back. The cannula can be suitably fixed to the skull by orthodontic resin (L.D. Culk Co., Milford, DE) bonded to two small machine screws (1/8" stainless steel slotted) inserted in the skull. The pump, tubing, and cannula are primed before insertion with infusate solutions; a 3-0 nylon suture is inserted into the cannula during implantation to prevent obstruction by brain tissue. The wound is closed with 3-0 silk suture and cefazolin (10 mg, i.m.) is administered. After surgery animals are suitably kept in individual cages and fed soft food.

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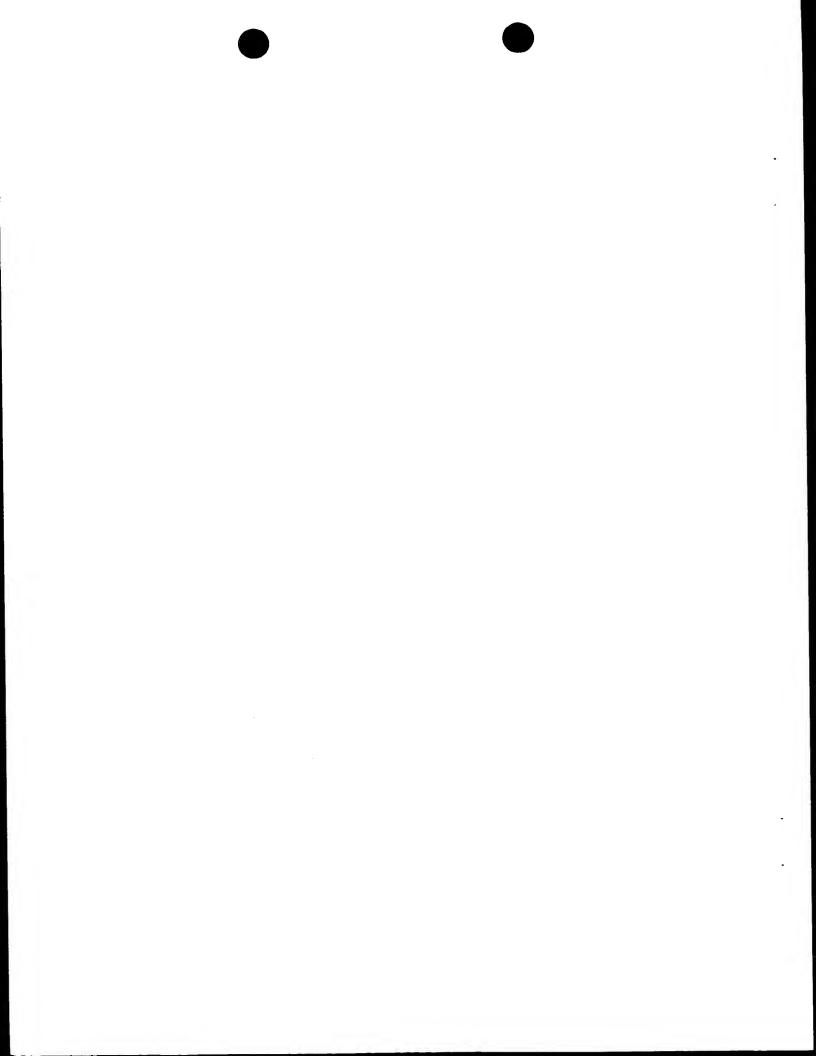
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Pumps are filled with vehicle alone (containing 127 mM NaCl, 2.6 mM KCl, 1.2 mM CaCl₂, 0.9 mM MgCl₂, 4.14 mM HEPES, 3 mM glycerin, 0.001% bovine serum albumin [BSA], and 0.01% fast green), or vehicle neuregulin or fragment or derivative thereof (100 μgm/ml). Heparin can be suitably used at relatively low doses, e.g. about 0.8 units/kg/day which is approximately 250-500 times less than a standard anticoagulant dose.

Three days after cannula implantation, animals are reanesthetized with 2% halothane and given atropine (0.15 mg/kg, i.p.). Animals are then intubated and connected to a ventilator (SAR-830; CWE Inc., Ardmore, PA) delivering 1% halothane/70% nitrous oxide in oxygen. The right femoral artery and vein are cannulated for monitoring of mean arterial blood pressure (MABP; Gould RS3200 Blood Pressure Monitor, Gould Inc., Valley View, OH), and blood sampling. Animals are then paralyzed with pancuronium bromide (0.5 mg/kg, i.v.). Arterial blood gasses (Corning 178 Blood Gas Analyzer, Ciba Corning Diagnostic Corp., Medford, MA), blood glucose (Accu-Check Blood Glucose Analyzer, Boehringer Mannheim, Indianapolis, IN), and hematocrit are measured at least twice during surgery and the immediate post-operative period. The stroke volume and rate of the



ventilator are adjusted to maintain PaO₂ between 100-200 mm Hg and PaCO₂ between 30-40 mm Hg. Core body temperature may be monitored by rectal thermocouple (e.g. Model 73ATA, Yellow Springs Instrument Co., Yellow Springs, OH) and maintained between 36-37°C with a homeothermic blanket control unit (Harvard Bioscience, South Natick, MA).

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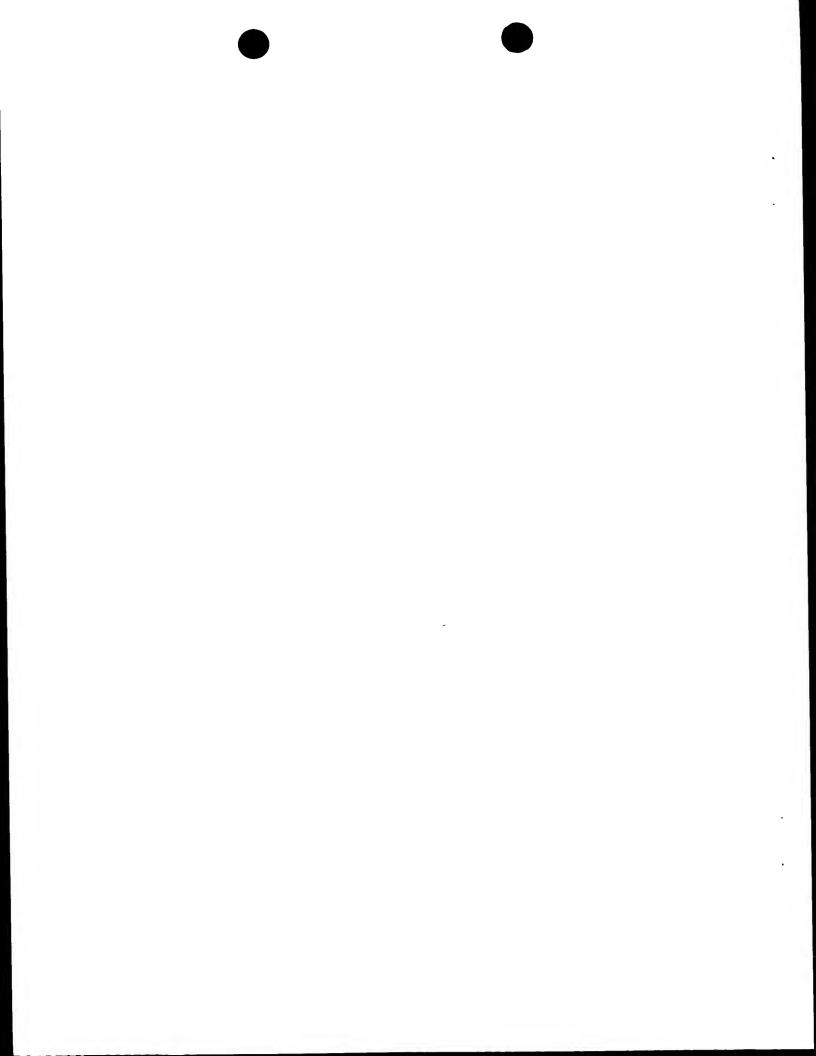
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Focal ischemic infarcts are made in the right lateral cerebral cortex in the territory of the middle cerebral artery (MCA) by the method of Chen, et al., *Stroke*, 17:738-743 (1986). Both common carotid arteries are exposed by midline anterior cervical incision. The animal is placed in a lateral position and a 1 cm skin incision is then made at the midpoint between the right lateral canthus and the anterior pinna. The temporal muscle is retracted, and a small (3 mm diameter) craniectomy is made at the junction of the zygoma and squamosal bone using a dental drill cooled with saline. Using a dissecting microscope, the dura can be opened with fine forceps, and the right MCA can be ligated with two 10-0 monofilament nylon ties just above the rhinal fissure and transected between the ties. Both common carotid arteries then can be occluded by microaneurysm clips for 45 minutes. After removal of the clips, return of flow is visualized in the arteries. Anesthesia is maintained for 15 minutes, and animals are returned to individual cages and fed soft food after surgery.

Twenty four hours after cerebral infarction, animals are again weighed, and then sacrificed by rapid decapitation. Brains are removed, inspected visually for the anatomy of the middle cerebral artery as well as for signs of hemorrhage or infection, immersed in cold saline for 10 minutes, and sectioned into six standard coronal slices (each 2 mm thick) using a rodent brain matrix slicer (Systems, Warren, MI). Brains are also examined visually for the presence of dye (fast green) in the cerebral ventricles. Slices are placed in the vital dye 2,3,5-triphenyl tetrazolium chloride (TTC, 2%; Chemical Dynamics Co., NH) at 37°C in the dark for 30 minutes, followed by 10% formalin at room temperature overnight. The outline of right and left cerebral hemispheres as well as that of infarcted tissue, clearly visualizable by lack of TTC staining (Chen et al., *Stroke*, 17:738-743 (1986)), is outlined on the posterior surface of each slice using an image analyzer (MTI videocamera and Sony video monitor connected to a Bioquant IV Image Analysis System run on an EVEREX computer). Infarct volume is calculated as the sum of infarcted area per slice multiplied by slice



thickness. Both the surgeon and image analyzer operator are blinded to the treatment given each animal.

Volumes of infarcts among vehicle vs. neuregulin-treated animals can be compared by unpaired, two-tailed t-tests for each experiment, and by two-way analysis of variance (ANOVA; Exp. X Treatment) for combined data. A subsequent slice-by-slice analysis of infarct area among pooled neuregulin- vs. vehicle-treated animals is suitably done by repeated measures two-way ANOVA (Treatment X Slice). Other anatomical and physiological measurements are compared among GDF-1- vs. vehicle-treated animals by unpaired, two-tailed t-tests using the Bonferroni correction for multiple pairwise comparisons.

Example 2 -- In vivo behavioral assays

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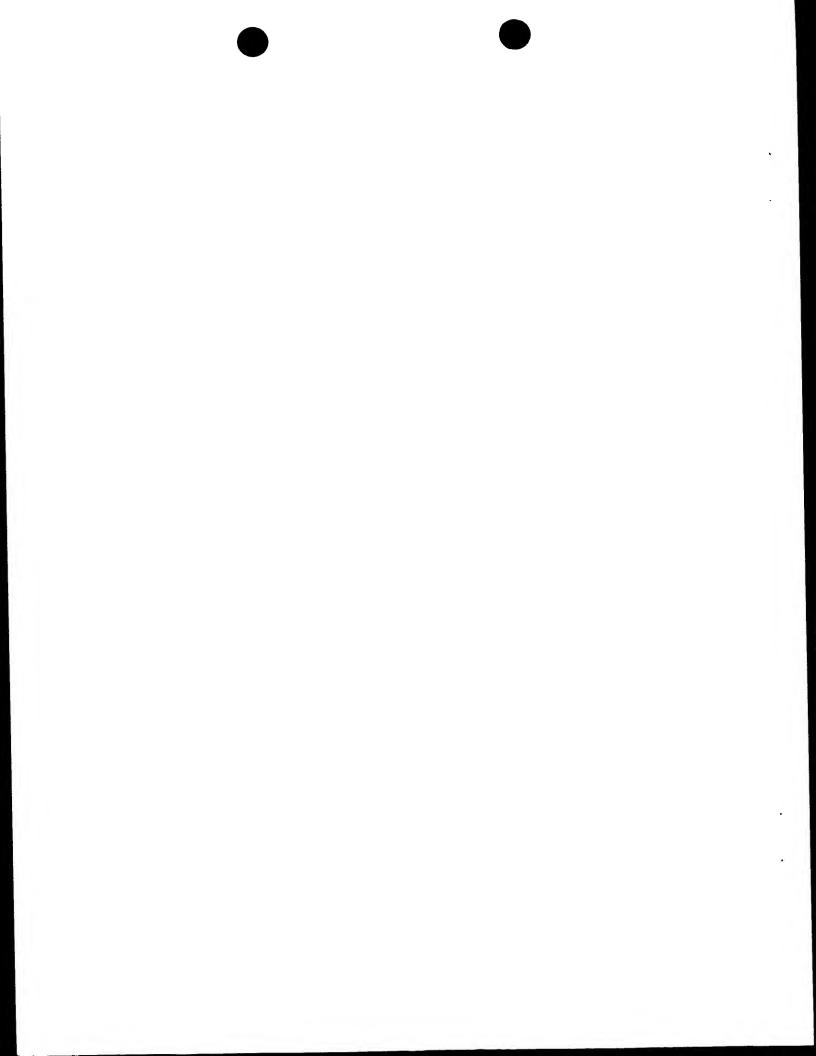
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For behavioral outcome studies, such as to assess recovery, repair and remodeling promoted by administration of a neuregulin or fragment or derivative thereof, or nucleic acid encoding same, a number of assays can be employed such as those described in G. Sinson et al., *J Neurochem*, 65(5):2209-2214 (1995); T.K. McIntosh et al., *Neuroscience*, 28:233-244 (1989); and T.K. McIntosh et al., *J Neurotrauma*, 10:373-384 (1993).

Briefly, one suitable behavioral assay as described in G. Sinson et al., *supra*, entails that test animals (male Sprague-Dawley rats) receive preinjury training in a Morris Water Maze, a circular tank 1 m in diameter that is filled with 18°C water. The water surface is made opaque with a covering of Styrofoam pieces. During training of the animals a submerged platform is present in the maze. Each test animal undergoes 20 training trials over a two day period during which they learn to locate the platform using external visual cues. Immediately following the last training trial, animals are anesthetized and subjected to a lateral (parasagittal) fluid-percussion (FP) brain injury. Briefly, a 5-mm craniectomy is performed over the left parietal cortex, midway between lamda and bregma. A hollow Leur-loc fitting is cemented to the craniectomy site. The injury is delivered after attaching the FP device. The injury should be of moderate severity (2.1-2.3 atm). After injury, the Leur-loc is removed, and the skin is sutured. Normothermia is maintained with warming pads until the animals being to ambulate.



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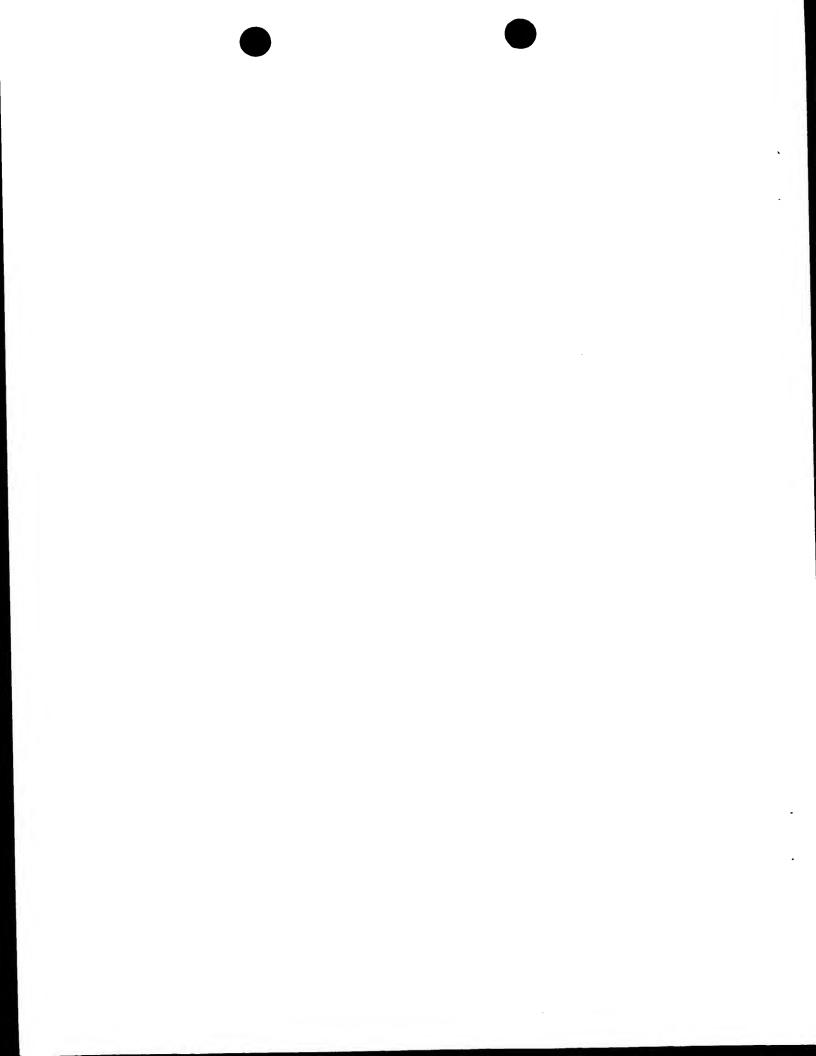
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At 72 hours, 1 week or 2 weeks after injury, animals are assessed for their ability to remember the learned task of locating the platform in the MWM. For this evaluation the platform is removed from the maze, and the animal's swimming pattern is suitably recorded with a computerized video system for 1 minute. The maze is separated in zones that are weighed according to the proximity to the platform's location. A memory score is generated by multiplying the weighted numbers by the time the animal spends in each zone and then adding the products.

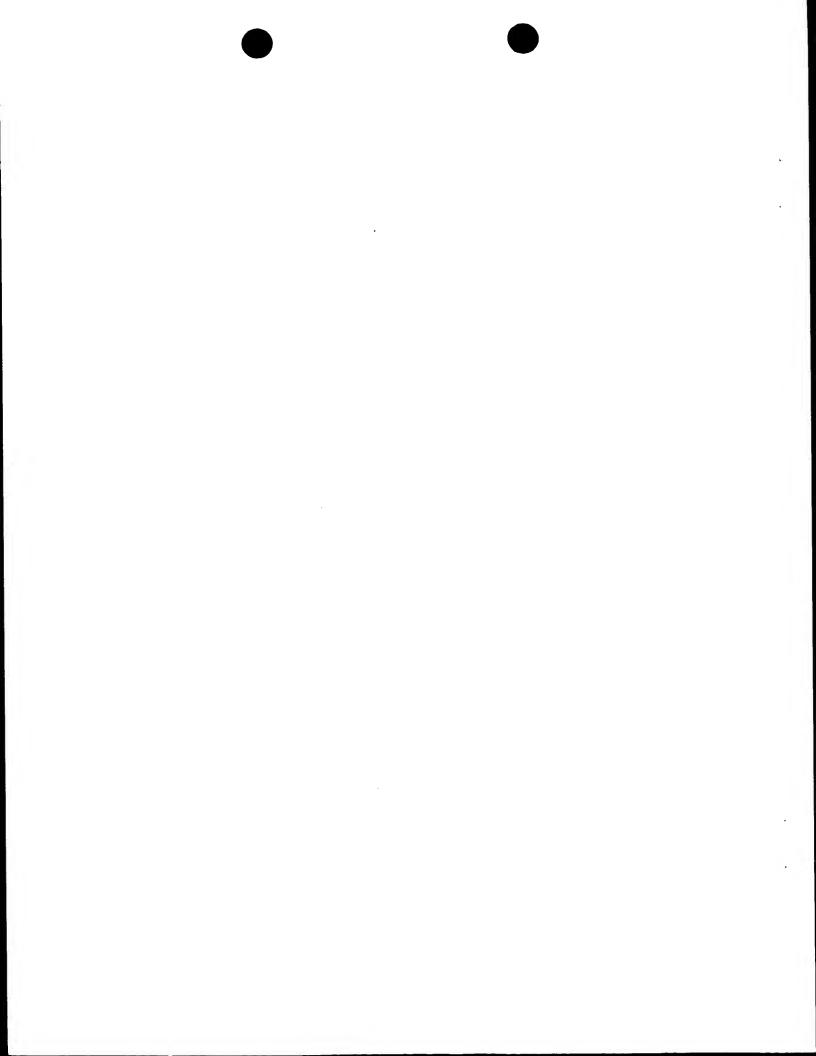
Animals surviving for 1 or 2 weeks also can undergo evaluation of neurologic motor function. Briefly, one suitable assay provides that animals are scored from 0 (severely impaired) to 4 (normal) for each of the following: (1) left and (2) right forelimb during suspension by the tail; (3) left and (4) right hindlimb flexion when the forelimbs remain on a surface and the hindlimbs are lifted up and back by the tail; the ability to resist lateral pulsion to the (5) left and (6) right; and the ability to stand on an inclined plane in the (7) left, (8) right, and (9) vertical positions. Scores are combined for each of the tests (1) through (9). The observer for the tests should be blinded to the animal's previous treatment.

The invention has been described in detail with reference to preferred embodiments thereof. However, it will be appreciated that those skilled in the art, upon consideration of this disclosure, may make modifications and improvements within the spirit and scope of the invention.



What is claimed is:

- 1. A method of treating a mammal suffering from or susceptible to stroke, brain or spinal cord injury or ischemia, or heart attack, comprising administering to the mammal a therapeutically effective amount of a neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a fragment or derivative of a neuregulin.
- 2. A method of treating a mammal suffering from or susceptible to optic nerve injury or retinal injury or ischemia, comprising administering to the mammal a therapeutically effective amount of a neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a fragment or derivative of a neuregulin.
- 3. A method of treating a mammal suffering from or susceptible to effects of post-surgical neurological deficits, hypoxia or hypoglycemia, comprising administering to the mammal a therapeutically effective amount of a neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a fragment or derivative of a neuregulin.
- 4. A method of treating a mammal suffering from or susceptible to epilepsy, Parkinson's disease, Huntington's disease, Amyotrophic Lateral Sclerosis, Alzheimer's disease, Down's Syndrome, Korsakoff's disease, or age-dependent dementia, comprising administering to the mammal a therapeutically effective amount of a neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a fragment or derivative of a neuregulin.
- 5. The method of claim 1 wherein the neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a fragment or derivative of a neuregulin is administered after the subject has suffered a stroke, brain or spinal cord injury or ischemia, or heart attack.
- 6. The method of claim 5 wherein the neuregulin, or fragment or derivative of a neuregulin, or a nucleic acid encoding a neuregulin or a fragment or derivative of a neuregulin is administered to the subject for at least about two weeks after the subject has suffered a stroke, brain or spinal cord injury or ischemia, or heart attack.

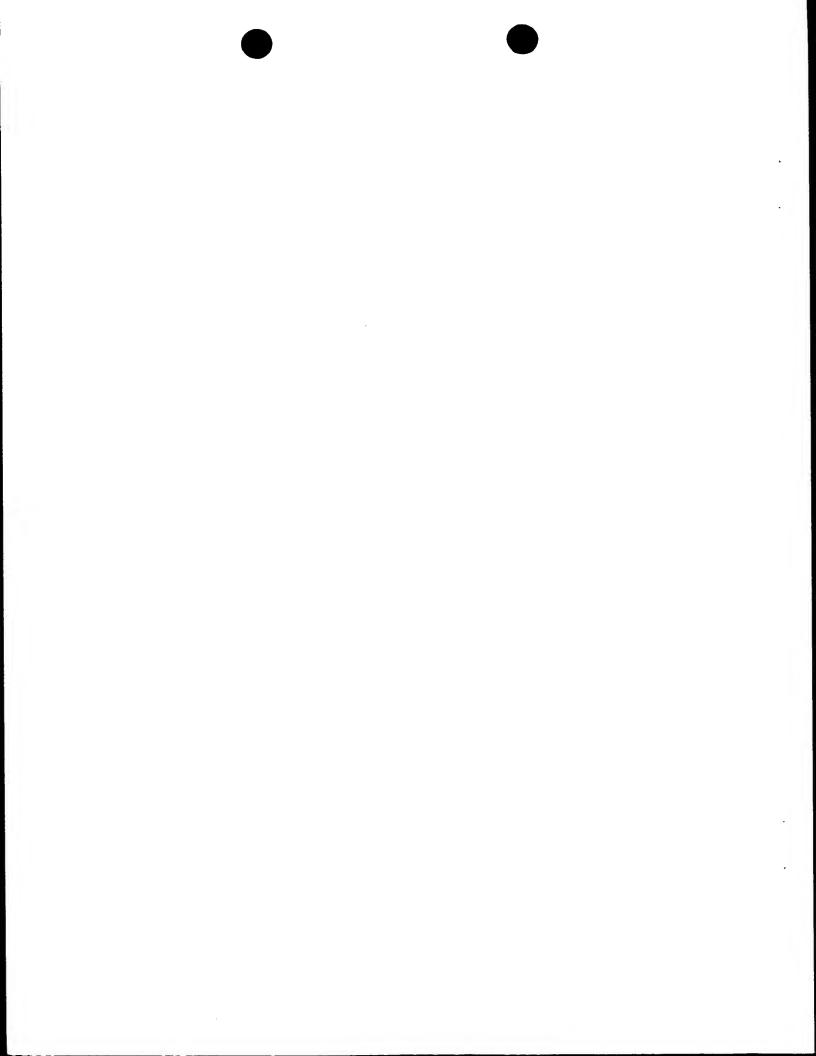


- 7. A method of any one of claims 1-6 wherein a neuregulin or a fragment or derivative thereof is administered to the mammal.
- 8. A method of claim 7 wherein the neuregulin or fragment or derivative thereof comprises an amino acid sequence of the following formula:

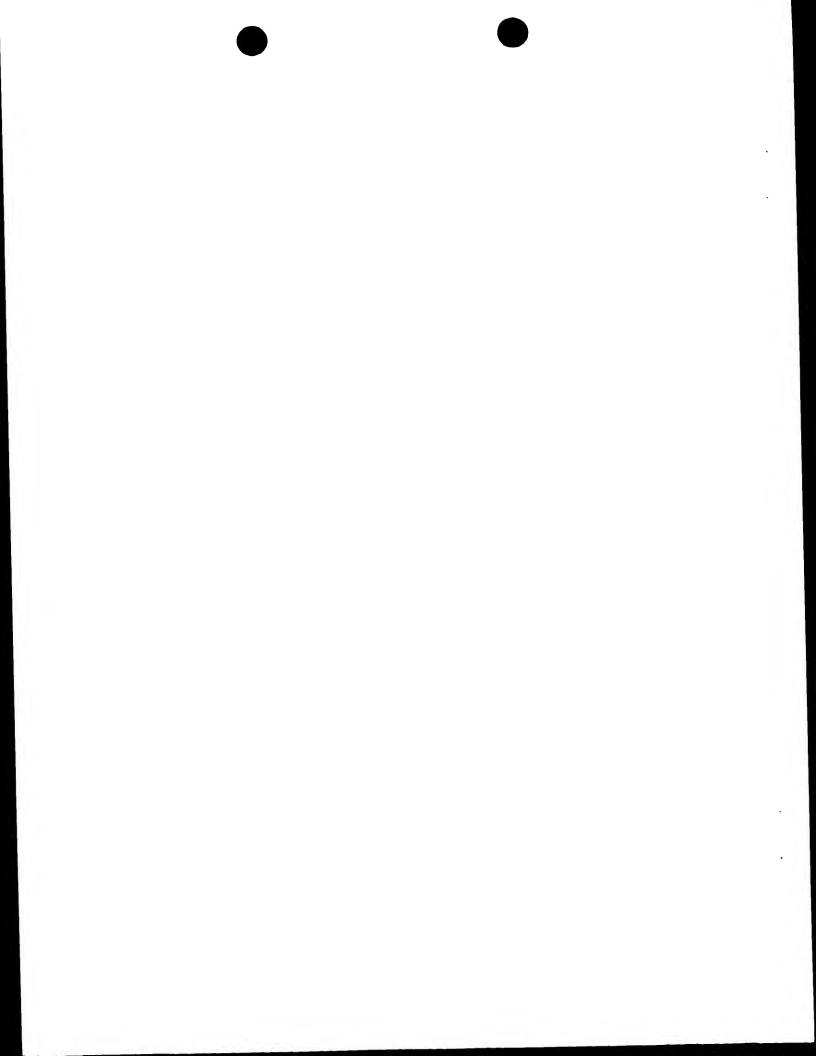
WYBAZCX

wherein WYBAZCX is composed of amino acid sequences that include one or more sequences shown in FIGS. 1 through 15 (which includes SEQ ID NOS:2, 4, 5, 8, 9, 12, 14, 15, 18, 19, 22, 23, 26, 27, 30, 33, 35, 38, 39, 41, 44, 45 and 48), wherein W comprises the polypeptide segment F, or is absent; wherein Y comprises the polypeptide segment E, or is absent; wherein Z comprises the polypeptide segment G or is absent; and wherein X comprise a polypeptide segment selected from the group consisting of C/D HKL, C/D H, C/D HL, C/D D, C/D' HL, C/D' HKL, C/D' H, C/D' D, C/D C/D' HKL, C/D C/D' H, C/D C/D' HL, C/D D' HKL, C/D C/D' D' HKL, C/D C/

- a) at least one of F, Y, B, A, Z, C or X is of bovine origin; or
- b) Y comprises the polypeptide segment E; or
- c) X comprises the polypeptide segments C/D HKL, C/D D, C/D' HKL, C/D C/D' HKL, C/D C/D' D, C/D D' H, C/D D' HL, C/D D' HKL, C/D' D' H, C/D' D' HKL, C/D' D' HKL, C/D' D' HKL, C/D' H, C/D C/D' H or C/D C/D' HL.
- 9. The method of claim 7 wherein the neuregulin or fragment or derivative thereof a) has at least one of F, Y, B, A, Z, C or X is of bovine origin; or b) Y comprises the polypeptide segment E; or c) X comprises the polypeptide segments C/D HKL, C/D D, C/D' HKL, C/D C/D' HKL, C/D C/D' D, C/D D H, C/D D' HL, C/D D' HKL, C/D C/D' D, C/D D' HKL, C/D C/D' D, C/D C/D' HKL, C/D' H
- 10. The method of claim 7 wherein the neuregulin or fragment or derivative thereof comprises FBA polypeptide segments, FEBA polypeptides segments, EBA polypeptide segments or FEBA' polypeptide segments.



- 11. A method of claim 7 wherein the neuregulin is encoded by a nucleic acid that comprises one of SEQ ID NOS:49, 51 and 53.
- 12. A method of claim 7 wherein the neuregulin or fragment or derivative thereof is encoded by a nucleic acid that comprises a sequence that has at least about 70% sequence identity to one of SEQ ID NOS:49, 51 and 53.
- 13. A method of claim 7 wherein the neuregulin or fragment or derivative thereof is encoded by a sequence that hybridizes to one of SEQ ID NOS:49, 51 or 53 under normal stringency conditions.
- 14. A method of claim 7 wherein the neuregulin or fragment or derivative thereof is encoded by a sequence that hybridizes to one of SEQ ID NOS:49, 51 or 53 under high stringency conditions.
- 15. A method of claim 7 wherein the neuregulin or fragment or derivative has at least about 70% sequence identity to SEQ ID NOS:50, 52 or 54.
- 16. A method of claim 7 wherein the neuregulin or fragment or derivative thereof is encoded by a nucleic acid that comprises a sequence that has at least about 70% sequence identity to one of SEQ ID NO:20 (Figure 7); SEQ ID NO:21 (Figure 7); SEQ ID NO:24 (Figure 8); SEQ ID NO:25 (Figure 8); SEQ ID NO:28 (Figure 9); or SEQ ID NO:29 (Figure 9).
- 17. A method of claim 7 wherein the neuregulin or fragment or derivative thereof is encoded by a sequence that hybridizes to one of SEQ ID NO:20 (Figure 7); SEQ ID NO:21 (Figure 7); SEQ ID NO:24 (Figure 8); SEQ ID NO:25 (Figure 8); SEQ ID NO:28 (Figure 9); or SEQ ID NO:29 (Figure 9) under normal stringency conditions.
- 18. A method of claim 7 wherein the neuregulin or fragment or derivative comprises a sequence that has at least about 70% sequence identity to any of the peptide sequences shown in Figures 7, 8 or 9 of the drawings.
- 19. A method of claim 7 where the neuregulin or fragment or derivative comprises a sequence that has at least about 80 percent homology to any of the peptide sequences shown in Figures 7, 8 or 9.
- 20. A method of claim 7 where the neuregulin or fragment or derivative comprises a sequence that has at least about 90 percent homology to any of the peptide sequences shown in Figures 7, 8 or 9.

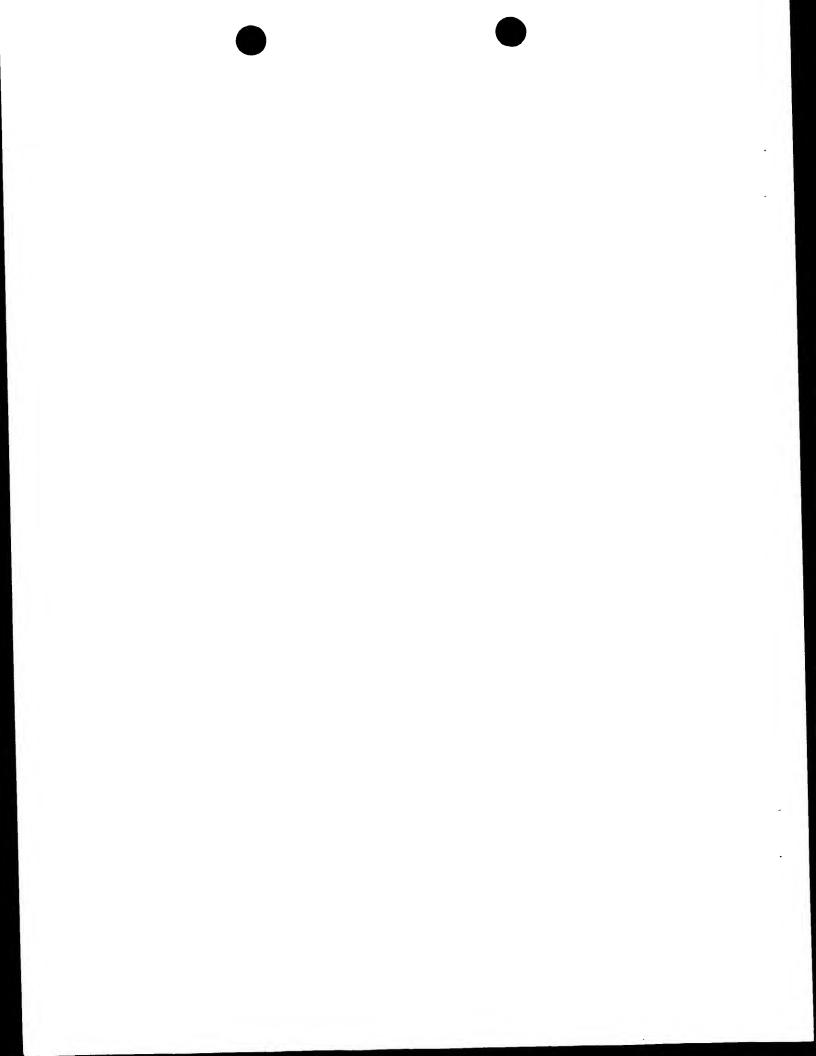


- 21. A method of claim 7 wherein the neuregulin or fragment or derivative comprises a sequence that has at least about 95 percent homology to any of the peptide sequences shown in Figures 7, 8 or 9.
- 22. A method of claim 7 wherein the neuregulin or fragment or derivative comprises a sequence that is shown in Figures 7, 8 or 9.
- 23. A method of any one of claims 1-6 wherein a nucleic acid encoding a neuregulin or a fragment or derivative thereof is administered to the mammal.
- 24. A method of claim 23 wherein the nucleic acid is SEQ ID NO:49, 51 or 53, or the complement thereof.
- 25. A method of claim 23 wherein the nucleic or fragment or derivative thereof encodes a neuregulin or neuregulin fragment or derivative that comprises an amino acid sequence of the following formula:

WYBAZCX

wherein WYBAZCX is composed of amino acid sequences that include one or more sequences shown in FIGS. 1 through 15 (which includes SEQ ID NOS:2, 4, 5, 8, 9, 12, 14, 15, 18, 19, 22, 23, 26, 27, 30, 33, 35, 38, 39, 41, 44, 45 and 48), wherein W comprises the polypeptide segment F, or is absent, wherein Y comprises the polypeptide segment G or is absent; and wherein X comprise a polypeptide segment selected from the group consisting of C/D HKL, C/D H, C/D HL, C/D D, C/D' HL, C/D' HKL, C/D' H, C/D' D, C/D C/D' HKL, C/D C/D' HKL, C/D D' HKL, C/D' D' HKL.

- 27. The method of claim 25 wherein the neuregulin or neuregulin fragment or derivative comprises FBA polypeptide segments, FEBA polypeptides segments,



EBA polypeptide segments, EBA' polypeptide segments or FEBA' polypeptide segments.

- 28. A method of claim 23 wherein the nucleic acid comprises a sequence that hybridizes to SEQ ID NO:20 (Figure 7); SEQ ID NO:21 (Figure 7); SEQ ID NO:24 (Figure 8); SEQ ID NO:25 (Figure 8); SEQ ID NO:28 (Figure 9); or SEQ ID NO:29 (Figure 9) under normal stringency conditions.
- 29. A method of claim 23 wherein the nucleic acid comprises a sequence that hybridizes to SEQ ID NO:SEQ ID NO:20 (Figure 7); SEQ ID NO:21 (Figure 7); SEQ ID NO:24 (Figure 8); SEQ ID NO:25 (Figure 8); SEQ ID NO:28 (Figure 9); or SEQ ID NO:29 (Figure 9) under high stringency conditions.
- 30. A method of claim 23 wherein the nucleic acid comprises a sequence that has at least about 70 percent homology to any of the nucleic acid sequences shown in Figures 7, 8 or 9.
- 31. A method of claim 23 wherein the nucleic acid comprises a sequence that has at least about 80 percent homology to any of the nucleic acid sequences shown in Figures 7, 8 or 9.
- 32. A method of claim 23 wherein the nucleic acid comprises a sequence that has at least about 90 percent homology to any of the nucleic acid sequences shown in Figures 7, 8 or 9.
- 33. A method of claim 23 wherein the nucleic acid comprises a sequence that has at least about 95 percent homology to any of the nucleic acid sequences shown in Figures 7, 8 or 9.
- 34. A method of claim 23 wherein the nucleic acid comprises a sequence shown in Figures 7, 8 or 9.
- 35. A method of any one of claims 1-34 wherein the administered neuregulin fragment or derivative, or the administered nucleic acid encodes a neuregulin fragment or derivative exhibits at least about a 10% reduction in infarct volume in an *in vivo* cerebral ischemia assay.
 - 36. A method of any one of claims 1-35 wherein the mammal is a human.

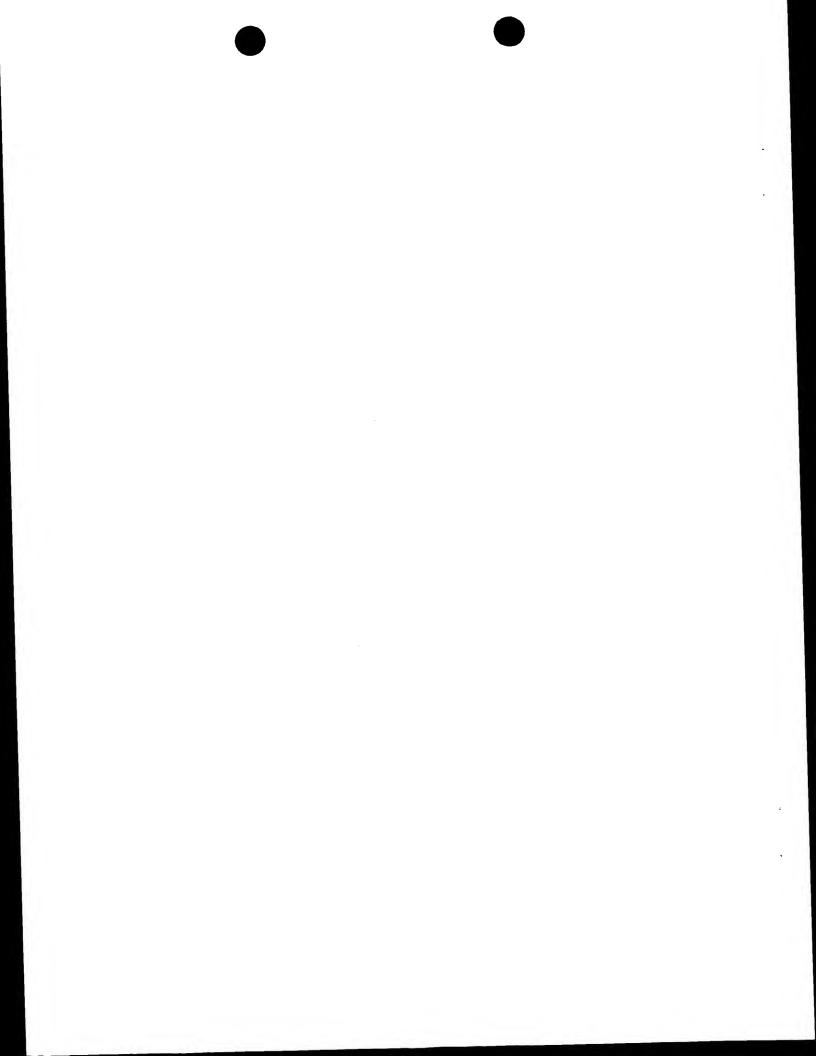


FIG. 1A HUMAN SEGMENT E: (SEQ ID NOS:1-2)

ATG Met 1	AGA Arg	TGG Trp	CGA Arg	CGC Arg 5	GCC Ala	CCG Pro	CGC Arg	CGC Arg	TCC Ser 10	GGG Gly	CGT Arg	CCC Pro	GGC Gly	CCC Pro 15	CGG Arg	48
GCC Ala	CAG Gln	CGC Arg	CCC Pro 20	GGC Gly	TCC Ser	GCC Ala	GCC Ala	CGC Arg 25	TCG Ser	TCG Ser	CCG Pro	CCG Pro	CTG Leu 30	CCG Pro	CTG Leu	96
CTG Leu	CCA Pro	CTA Leu 35	CTG Leu	CTG Leu	CTG Leu	CTG Leu	GGG Gly 40	ACC Thr	GCG Ala	GCC Ala	CTG Leu	GCG Ala 45	CCG Pro	GGG Gly	GCG Ala	144
GCG Ala	GCC Ala 50	GGC Gly	AAC Asn	GAG Glu	GCG Ala	GCT Ala 55	CCC Pro	GCG Ala	GGG Gly	GCC Ala	TCG Ser 60	GTG Val	TGC Cys	TAC Tyr	TCG Ser	192
TCC Ser 65	CCG Pro	CCC Pro	AGC Ser	GTG Val	GGA Gly 70	TCG Ser	GTG Val	CAG Gln	GAG Glu	CTA Leu 75	GCT Ala	CAG Gln	CGC Arg	GCC Ala	GCG Ala 80	240
GTG Val	GTG Val	ATC Ile	GAG Glu	GGA Gly 85	AAG Lys	GTG Val	CAC His	CCG Pro	CAG Gln 90	CGG Arg	CGG Arg	CAG Gln	CAG Gln	GGG Gly 95	GCA Ala	288
CTC Leu	GAC Asp	AGG Arg	AAG Lys 100	GCG Ala	GCG Ala	GCG Ala	Ala	GCG Ala 105	GGC Gly	GAG Glu	GCA Ala	Gly	GCG Ala 110	TGG Trp	GGC Gly	336
GGC Gly	GAT Asp	CGC Arg 115	GAG G1u	CCG Pro	CCA Pro	GCC Ala	GCG Ala 120	GGC Gly	CCA Pro	CGG Arg	GCG Ala	CTG Leu 125	GGG Gly	CCG Pro	CCC Pro	384
GCC Ala	GAG Glu 130	GAG G1u	CCG Pro	CTG Leu	CTC Leu	GCC Ala 135	GCC Ala	AAC Asn	GGG Gly	ACC Thr	GTG Val 140	CCC Pro	TCT Ser	TGG Trp	CCC Pro	432
ACC Thr 145	GCC Ala	CCG Pro	GTG Val	Pro	AGC Ser 150	GCC Ala	GGC Gly	GAG Glu	CCC Pro	GGG Gly 155	GAG Glu	GAG Glu	GCG Ala	CCC Pro	TAT Tyr 160	480
CTG Leu	GTG Val	AAG Lys	GTG Val	CAC His 165	CAG Gln	GTG Val	TGG Trp	GCG Ala	GTG Val 170	AAA Lys	GCC Ala	GGG Gly	GGC Gly	TTG Leu 175	AAG Lys	528
AAG Lys	GAC Asp	TCG Ser	CTG Leu 180	CTC . Leu	ACC Thr	GTG Val	Arg	CTG Leu 185	GGG Gly	ACC Thr	TGG Trp	GGC Gly	CAC His 190	CCC Pro	GCC Ala	576

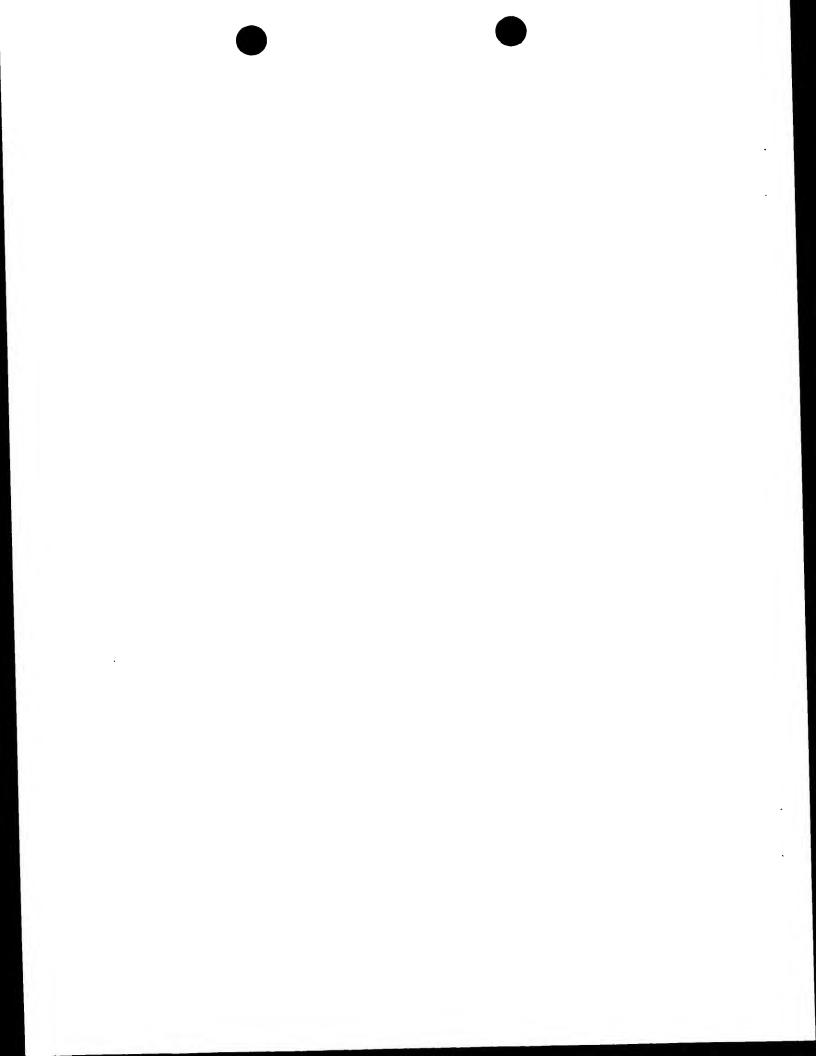


FIG. 1B

TTC Phe	CCC Pro	TCC Ser 195	TGC Cys	GGG Gly	AGG Arg	CTC Leu	AAG Lys 200	GAG Glu	GAC Asp	AGC Ser	AGG Arg	TAC Tyr 205	ATC Ile	TTC Phe	TTC Phe	6	524
ATG Met	GAG Glu 210	CCC Pro	GAC Asp	GCC Ala	AAC Asn	AGC Ser 215	ACC Thr	AGC Ser	CGC Arg	GCG Ala	CCG Pro 220	GCC Ala	GCC Ala	TTC Phe	CGA Arg	6	572
GCC Ala 225	TCT Ser	TTC Phe	CCC Pro	CCT Pro	CTG Leu 230	GAG Glu	ACG Thr	GGC Gly	CGG Arg	AAC Asn 235	CTC Leu	AAG Lys	AAG Lys	GAG Glu	GTC Val 240	7	'20
AGC Ser	CGG Arg	GTG Val	CTG Leu	TGC Cys 245	AAG Lys	CGG Arg	TGC Cys	G								7	'45

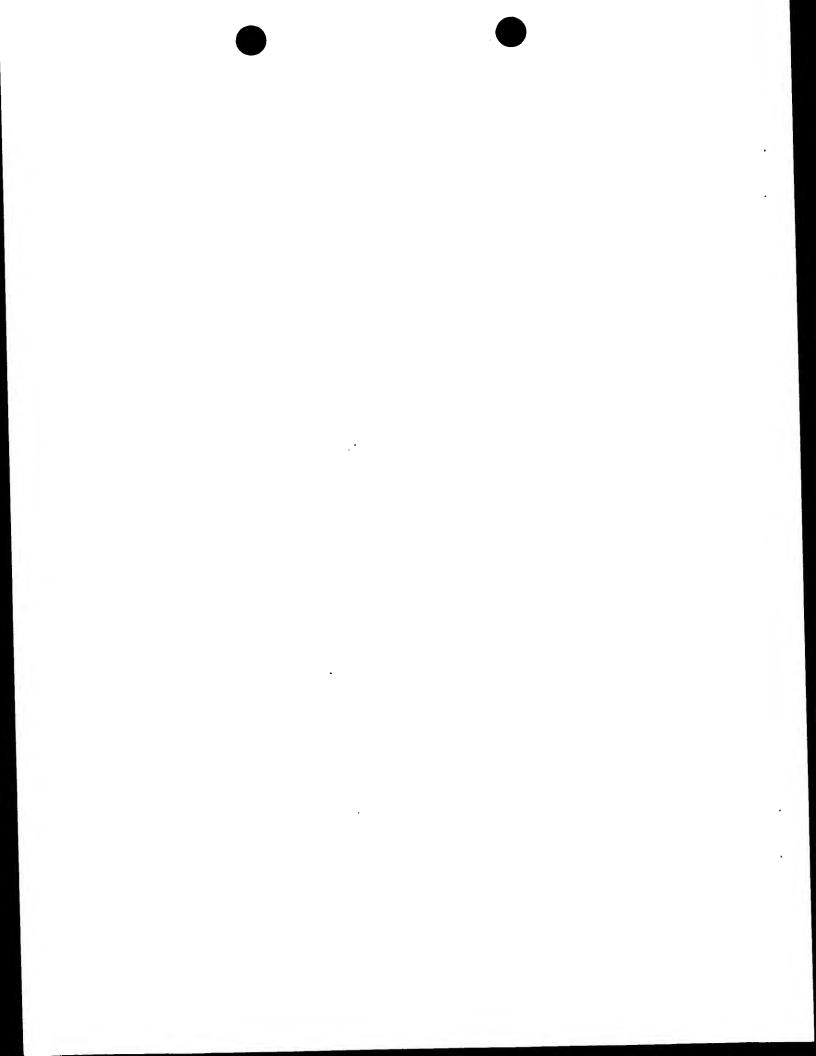


FIG.2 SEGMENT E: (SEQ ID NOS:3-4)

CC	CAT His 1	CAA G1n	GTG Val	TGG Trp	GCG Ala 5	GCG Ala	AAA Lys	GCC Ala	GGG Gly	GGC Gly 10	TTG Leu	AAG Lys	AAG Lys	GAC Asp	TCG Ser 15		47
CTG Leu	CTC Leu	ACC Thr	GTG Val	CGC Arg 20	CTG Leu	GGC Gly	GCC Ala	TGG Trp	GGC Gly 25	CAC His	CCC Pro	GCC Ala	TTC Phe	CCC Pro 30	TCC Ser		95
TGC Cys	GGG Gly	CGC Arg	CTC Leu 35	AAG Lys	GAG Glu	GAC Asp	AGC Ser	AGG Arg 40	lyr	ATC Ile	TTC Phe	TTC Phe	ATG Met 45	Glu	CCC Pro		143
GAG Glu	GCC Ala	AAC Asn 50	AGC Ser	AGC Ser	GGC Gly	GGG Gly	CCC Pro 55	GGC Gly	CGC Arg	CTT Leu	CCG Pro	AGC Ser 60	Leu	CTT Leu	CCC Pro		191
CCC Pro	TCT Ser 65	CGA Arg	GAC Asp	GGG Gly	CCG Pro	GAA G1u 70	CCT Pro	CAA Gln	GAA Glu	GGA Gly	GGT Gly 75	CAG Gln	CCG Pro	GGT Gly	GCT Ala		239
GTG Val 80	CAA Gln	CGG Arg	TGC Cys	G												-	252
					SI	FCME	'NIT		IG.		NOC		21				
					٥.	_UIIL	14.1	υ. ι	SEQ	IU	NOS	:5-8	5)				
001	Tuc		Arg CCC CCC Q	ULI	Lys TGA	G1u AAG	His AGA	Lys TGA	Ser AGA	Gln GTC	Glu AGG	Ser AGT	Val	TGG	CAG		48
Ser GTT	TGC Lys	CTC Leu AAC	CCC Q Val TAG	GAT GAT Leu TGC	Lys TGA TGA Arg TTC	Glu AAG AAG Cys GGT	His AGA AGA	Lys TGA TGA Thr AGA	Ser AGA AAA Ser CCA	Gln GTC GCC	Glu AGG AGG	Ser AGT AAT	Val CTG CGG A	TGG	CAG CAG		48 96
Ser GTT Lys TCA	Lys CCA LII CCA Phe AGT	Leu AAC III AAC Lys TCA	CCC Q Val TAG III TAG Trp AGT	Leu TGC TCC Phe GGT	Lys TGA III TGA Arg TTC III TTC	Glu AAG III AAG Cys GGT III GGT ASn AGA	His AGA AGA Glu GCG GTG GTG ATG ATG	Lys TGA TGA Thr AGA AAA Ser GGA	Ser AGA AAA Ser CCA Glu GTG	Gln GTC GCC Ser GTT GTT Leu AAT	Glu AGG III AGG Glu CTG III CTG Ser TAA	Ser AGT AAT Tyr AAT AAT Arg GCC	Val CTG CGG A Ser ACT	Ser CCT Ser CCT LII CCT Asn AGA	CAG Leu CTC LII CTC Lys ACA		

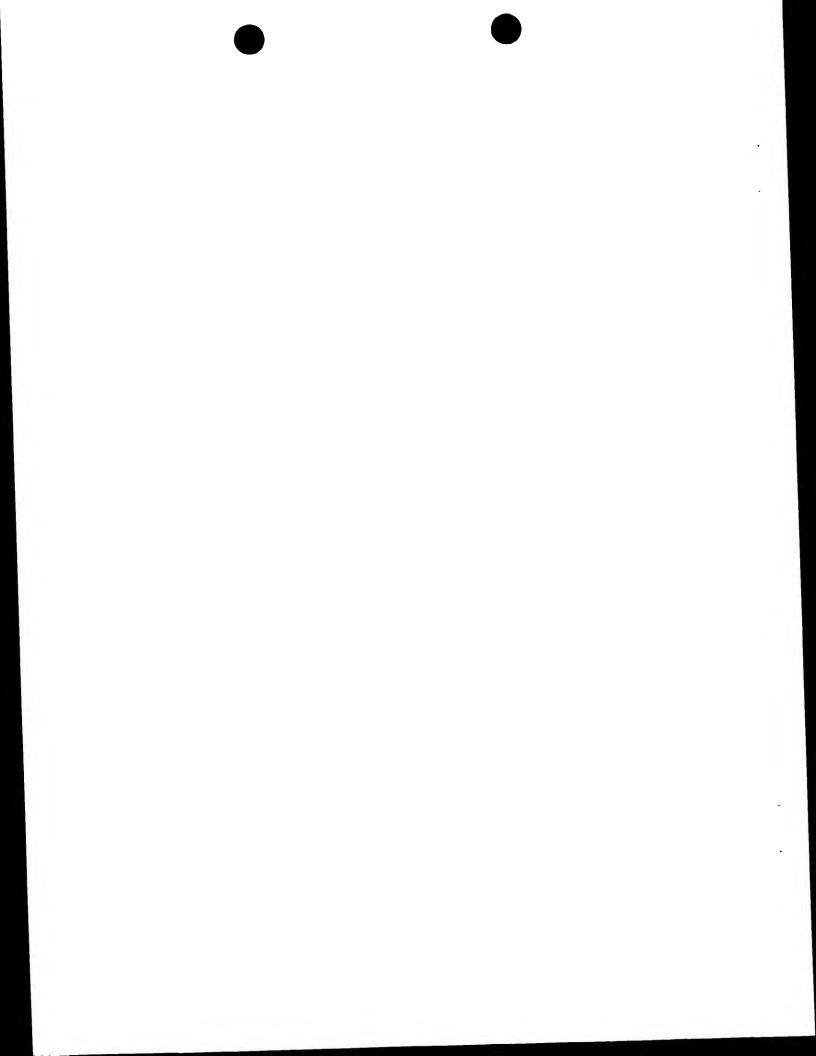


FIG 4 SEGMENT A: (SEQ ID NOS:9-12)

Lys Ser Glu Leu Arg Ile Ser Lys Ala Ser Leu Ala Asp Ser Gly G AAG TCA GAA CTT CGC ATT AGC AAA GCG TCA CTG GCT GAT TCT GGA G AAG TCA GAA CTT CGC ATT AAC AAA GCA TCA CTG GCT GAT TCT GGA N	46
Glu Tyr Met Cys Lys Val Ile Ser Lys Leu Gly Asn Asp Ser Ala Ser GAA TAT ATG TGC AAA GTG ATC AGC AAA CTA GGA AAT GAC AGT GCC TCT 	94
Ala Asn Ile Thr Ile Val Glu Ser Asn Ala GCC AAC ATC ACC ATT GTG GAG TCA AAC G 	122

FIG.5 SEGMENT A': (SEQ ID NOS:13-14)

TCTAAAACTA CAGAGACTGT ATTTTCATGA TCATCATAGT TCTGTGAAAT ATACTTAAAC	60
CGCTTTGGTC CTGATCTTGT AGG AAG TCA GAA CTT CGC ATT AGC AAA GCG Lys Ser Glu Leu Arg Ile Ser Lys Ala 1 5	110
TCA CTG GCT GAT TCT GGA GAA TAT ATG TGC AAA GTG ATC AGC AAA CTA Ser Leu Ala Asp Ser Gly Glu Tyr Met Cys Lys Val Ile Ser Lys Leu 10 25	158
GGA AAT GAC AGT GCC TCT GCC AAC ATC ACC ATT GTG GAG TCA AAC GGT Gly Asn Asp Ser Ala Ser Ala Asn Ile Thr Ile Val Glu Ser Asn Gly 30	206
AAG AGA TGC CTA CTG CGT GCT ATT TCT CAG TCT CTA AGA GGA GTG ATC Lys Arg Cys Leu Leu Arg Ala Ile Ser Gln Ser Leu Arg Gly Val Ile 45	254
AAG GTA TGT GGT CAC ACT TGAATCACGC AGGTGTGTGA AATCTCATTG Lys Val Cys Gly His Thr 60	302
TGAACAAATA AAAATCATGA AAGGAAAACT CTATGTTTGA AATATCTTAT GGGTCCTCCT	362
GTAAAGCTCT TCACTCCATA AGGTGAAATA GACCTGAAAT ATATATAGAT TATTT	417

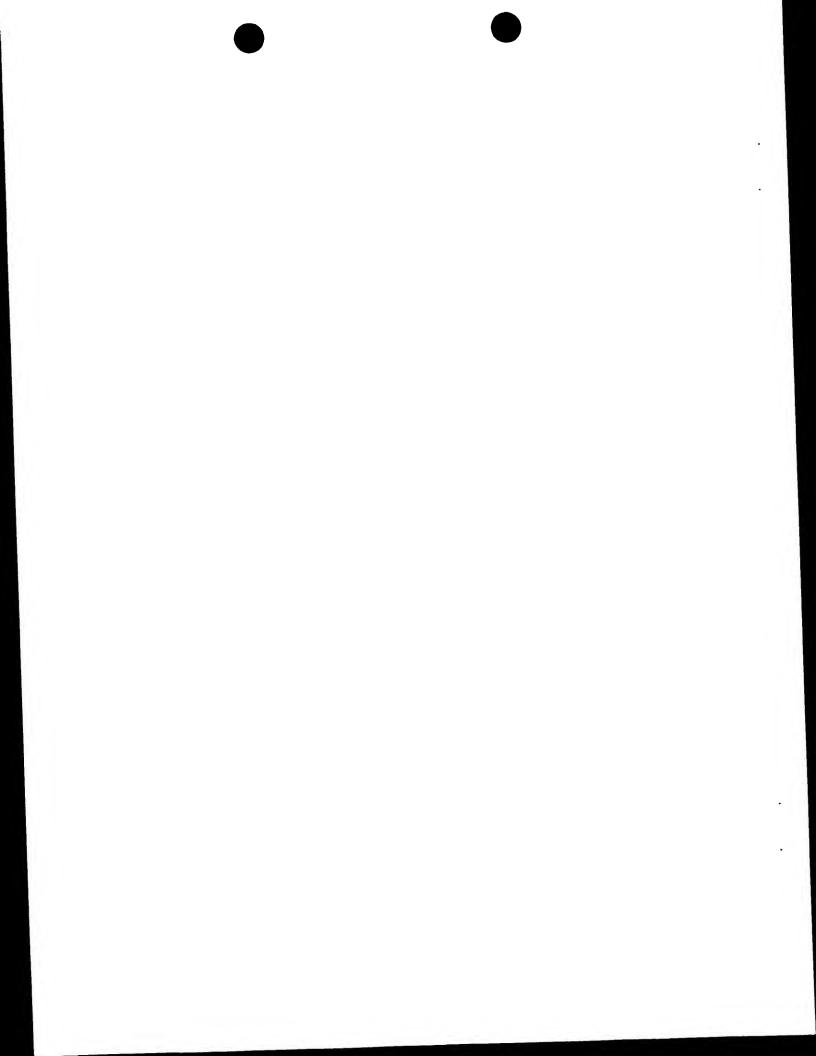


FIG. 6 SEGMENT G: (SEQ ID NOS:15-18)

												Ala GCG GCA			Ser TCT †CT	47
	G, (Q	101	\sim	\sim	MIJM	AIA	HA	1514	1 (A	$M \cap M$	f.: A A	Gly GGA GGA	$\Lambda \cap \Lambda$	A A T	407	95
Ser TCT	Ser TCA []] TCA	Ser T											Α			102

FIG. 7 SEGMENT C: (SEQ ID NOS:19-22)

CC ACA TCC ACA TCT ACA GCT GGG ACA AGC CAT CTT GTC AAG TGT GCA	47
Glu Lys Glu Lys Thr Phe Cys Val Asn Gly Gly Glu Cys Phe Met Val GAG AAG GAG AAA ACT TTC TGT GTG AAT GGA GGC GAG TGC TTC ATG GTG [95
-ys Asp Leu Ser Asn Pro Ser Arg Tyr Leu Cys VAA GAC CTT TCA AAT CCC TCA AGA TAC TTG TGC 	128

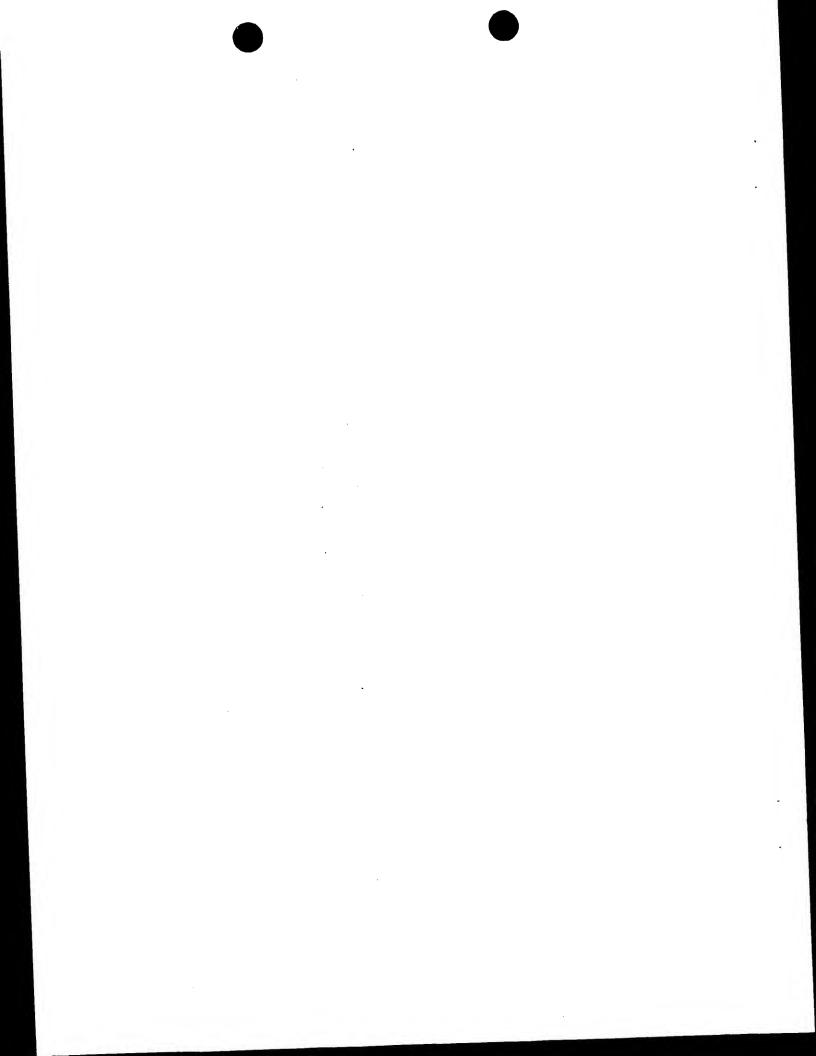


FIG. 8 SEGMENT C/D: (SEQ ID NOS:23-26)

AAG	160	CAA	CCI	GGA	-11C	ACT	GGA	GCG	AGA	Cys TGT []] TGT	ACT	GAG	AAT	GTG	Pro CCC CCC	48
ATG	Lys AAA AAA	GTC	CAA	ACC	CAA	GAA										69

FIG. 9 SEGMENT C/D': (SEQ ID NOS:27-29)

AAG	166	CCA	AA I	GAG	111	ACT	GGT	GAT	CGC	TGC	CAA	AAC	TAC	GTA	Met ATG ATG	~	48	
Ala GCC GCC	AGC	ПC	TAC														60	

FIG. 10 SEGMENT D: (SEQ ID NOS:30-32)

Ser Thr Ser Thr	Pro Phe Leu	Ser Leu Pro Glu *	
AGI ACG FCC ACT	CCC TTT CTG	TCT CTG CCT GAA TAG	36
		tct ctg cct gaa tag	00
AGI ALG ILL ALI	- UUU - 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	HOLDIG ONLIGAD TAG	

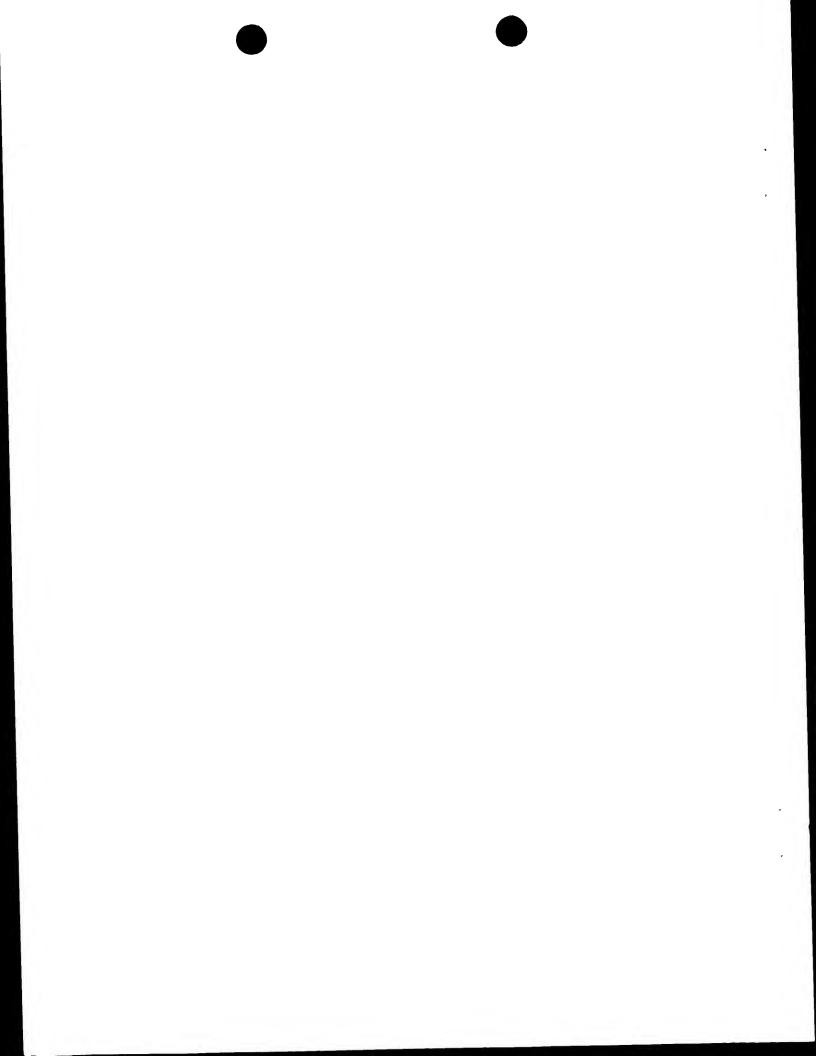


FIG. 11 SEGMENT D': (SEQ ID NOS:33-34)

Lys His Leu Gly Ile Glu Phe Met Glu AAG CAT CTT GGG ATT GAA TTT ATG GAG ${\tt GAG}$

27

FIG. 12A SEGMENT H: (SEQ ID NOS:35-38)

AAA	GCG	GAG	GAG	CIC	TAC	Gln CAG CAG	AAG	AGA	GTG	CTC	ACC	ATT	ACC	GGC	ATT	48
IGC	AIC	GCG	CTG	CTC	GTG	Val GTT GTC	GGC	ATC	ATG	TGT	GTG	GTG	GTC	Tyr TAC 111 TAC	Cys TGC []] TGC	96
AAA	ACC	AAG	AAA	CAA	CGG	Lys AAA AAA	AAG	CTT	CAT	GAC	CGG	CTT	Arg CGG CGG	Gln CAG CAG	AGC	144
CII	CGG	101	GAA	AGA	AAC	Thr ACC AAT N	ATG	ATG	AAC	GTA	GCC	AAC	GGG	CCC	His CAC CAC	192
CAC	CCC	AAT	CCG	CCC	CCC	Glu GAG GAG	AAC	GTG	CAG	CTG	GTG	AAT	CAA	TAC	Val GTA GTA	240
ICI	AAA	AAT	GIC	AIC	101	Ser AGC AGT	GAG	CAT	ATT	GTT	GAG	AGA	GAG	GCG	GAG	288
AGC	101		TCC 111	ACC	AGT	His CAC CAC	TAC	ACT	TCG	ACA	GCT 11	CAT	CAT	Ser TCC TCC	Thr ACT III ACT	336

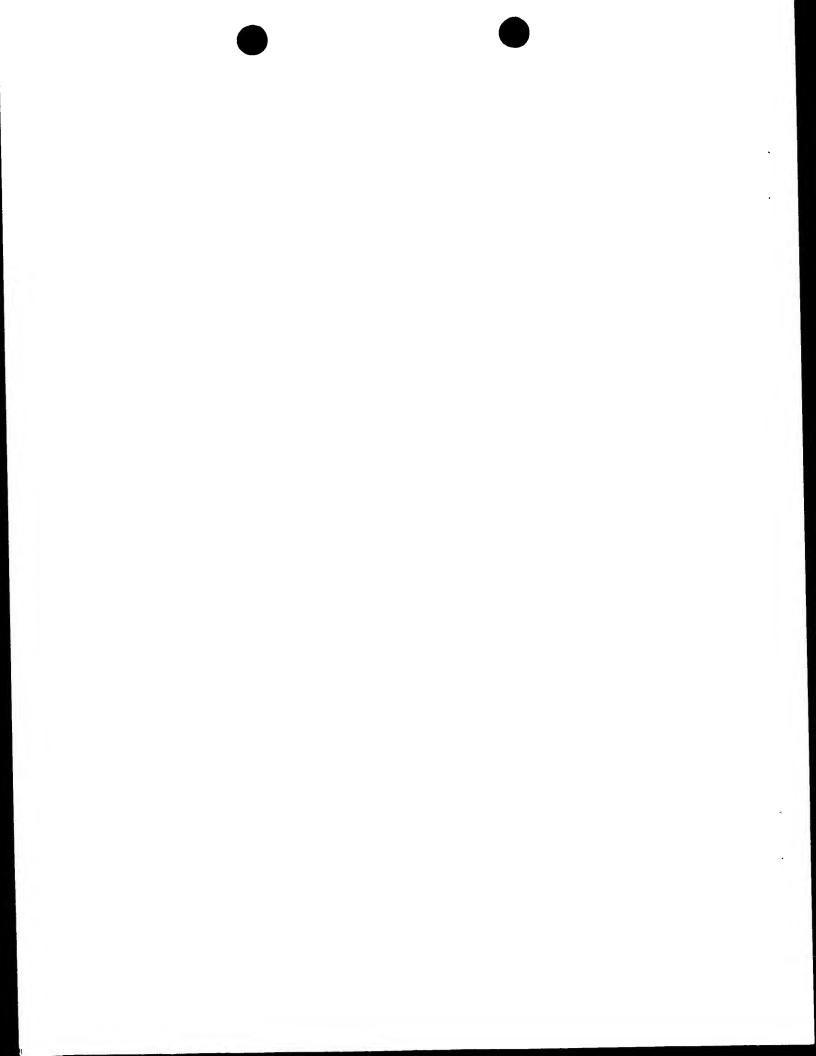


FIG. 12B

AC I	GIC	ACT	CAG	ACT	CCC	AGT	CAC	AGC	TGG	AGC	AAT	GGA	CAC	Thr ACT ACT	GAA	384
AGC	ATC	ATT	TCG	GAA	AGC	CAC	TCT	GTC	ATC	GTG	ATG	TCA	TCC	Val GTA GTA	GAA	432
AAC	AGT	AGG	CAC	AGC	AGC	CCG	ACT	GGG	GGC	CCG	AGA	GGA	CGŤ	Leu CTC CTT	AAT	480
GGC	HG	GGA	GGC	CCT	CGT	GAA	TGT	AAC	AGC	TTC	CTC	AGG	CAT	Ala GCC GCC	ΔGĂ	528
GAA	ACC	CCT	Asp GAC GAT	TCC	TAC	CGA	GAC	TCT	CCT	CAT	AGT	GAA	AG I			569

FIG. 13 SEGMENT K: (SEQ ID NOS:39-40)

A CAT AAC CTT ATA GCT GAG CTA AGG AGA AAC AAG GCC	CAC AGA TCC 46
His Asn Leu Ile Ala Glu Leu Arg Arg Asn Lys Ala	His Arg Ser
1 5 10	15
AAA TGC ATG CAG ATC CAG CTT TCC GCA ACT CAT CTT AG	A GCT TCT TCC 94
Lys Cys Met Gln Ile Gln Leu Ser Ala Thr His Leu Ar	g Ala Ser Ser
20 25	30
ATT CCC CAT TGG GCT TCA TTC TCT AAG ACC CCT TGG CC Ile Pro His Trp Ala Ser Phe Ser Lys Thr Pro Trp Pr 35	T TTA GGA AG 141 o Leu Gly Arg 45

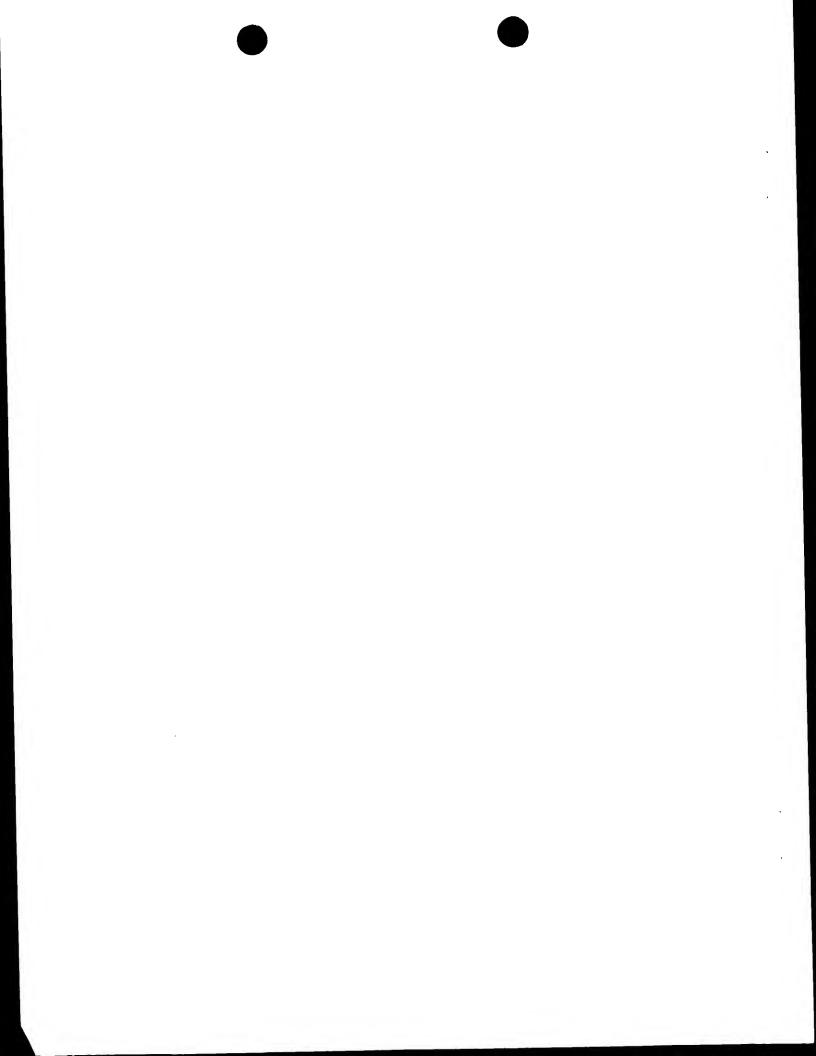


FIG. 14A SEGMENT L: (SEQ ID NOS:41-44)

Tyr Val Ser Ala Met Thr Thr Pro Ala Arg Met Ser Pro Val Asp G TAT GTA TCA GCA ATG ACC ACC CCG GCT CGT ATG TCA CCT GTA GAT 	46
Phe His Thr Pro Ser Ser Pro Lys Ser Pro Pro Ser Glu Met Ser Pro TTC CAC ACG CCA AGC TCC CCC AAG TCA CCC CCT TCG GAA ATG TCC CCG LII LII LII LII LII LII LII LII LII LI	94
Pro Val Ser Ser Thr Thr Val Ser Met Pro Ser Met Ala Val Ser Pro CCC GTG TCC AGC ACG ACG GTC TCC ATG CCC TCC ATG GCG GTC AGT CCC III III III III III III III III II	142
Phe Val Glu Glu Arg Pro Leu Leu Val Thr Pro Pro Arg Leu TTC GTG GAA GAG GAG AGA CCC CTG CTC CTT GTG ACG CCA CCA CGG CTG III III III III III III III III III I	190
Arg Glu Lys - Tyr Asp His His Ala Gln Gln Phe Asn Ser Phe His CGG GAG AAG TAT GAC CAC CAC GCC CAG CAA TTC AAC TCG TTC CAC LII III III III III III III III III CGG GAG AAG AAG AAG TTT GAC CAT CAC CCT CAG CAG TTC AGC TCC TTC CAC K F	238
Cys Asn Pro Ala His Glu Ser Asn Ser Leu Pro Pro Ser Pro Leu Arg TGC AAC CCC GCG CAT GAG AGC AAC AGC CTG CCC CCC AGC CCC TTG AGG CAC AAC CCC GCG CAT GAG AGC AGT AAC AGC CTC CCT GCT AGC CCC TTG AGG N D D AAC AGC CTC CCT GCT AGC CCC TTG AGG	286
Ile Val Glu Asp Glu Glu Tyr Glu Thr Thr Gln Glu Tyr Glu Pro Ala ATA GTG GAG GAT GAG GAA TAT GAA ACG ACC CAG GAG TAC GAA CCA GCT 	334
Gln Glu Pro Val Lys Lys Leu Thr Asn Ser Ser Arg Arg Ala Lys Arg CAA GAG CCG GTT AAG AAA CTC ACC AAC AGC AGC CGG CGG GCC AAA AGA III III III III III III III III	382

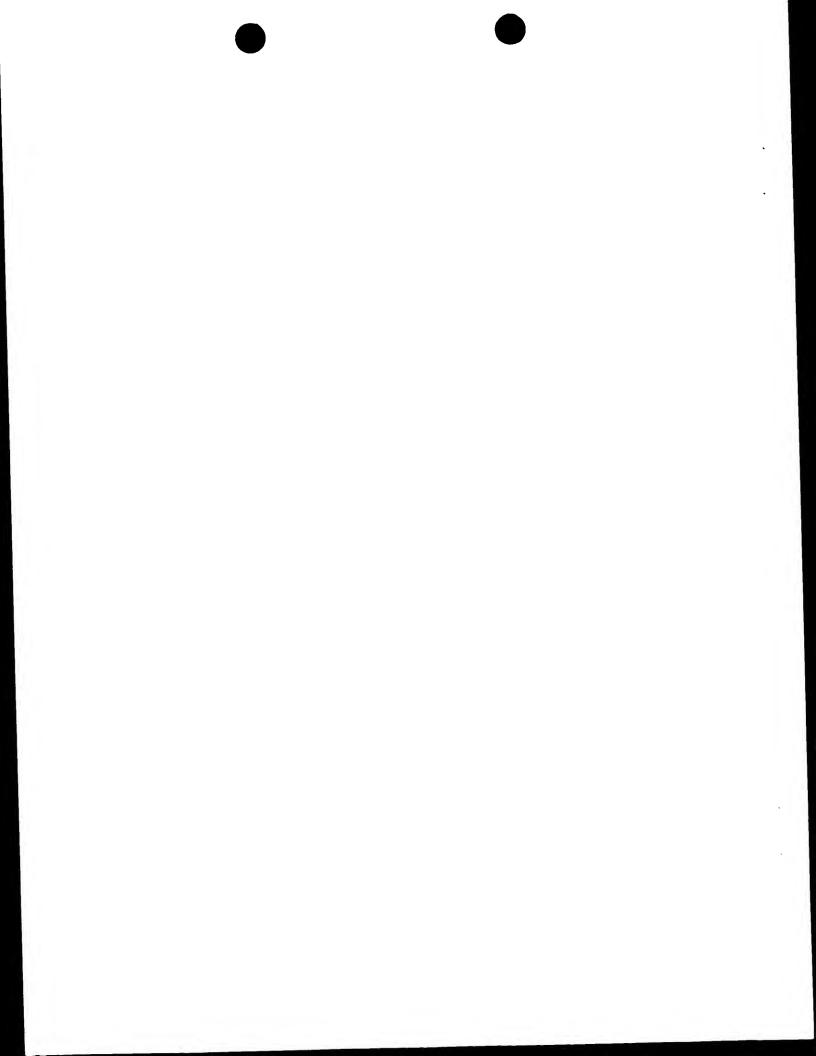


FIG. 14B

Thr ACC ACC	Lys AAG AAG	Pro CCC CCC	Asn AAT III AAT	Gly GGT [] GGC	His CAC []] CAC	Ile ATT ATT	Ala GCC GCT	His CAC II AAC N	Arg AGG AGA	Leu TTG TTG	Glu GAA GAA	Met ATG GTG V	Asp GAC []] GAC	Asn AAC AGC S	Asn AAC AAC	430
ALA	Gly GGC II AGC S	661	GAL	AGC	AGI	AAC	1CA	GAG	AGC	GAA	ACA	GAG	Asp GAT GAT	GAA	Arg AGA AGA	478
GIA 	Gly GGA GGT	GAA	GAI	ACG	111	HC	Leu CTG CTG	GCC	ATA	CAG	AAC	CCC	Leu CTG []] CTG	Ala GCA GCA	GCC	526
AGI		GAG	G (G	GCC	CCT	GCC	110	CGC	CTG	GTC	GAC	AGC	Arg AGG AGG	ACT	AAC	574
CCA	ACA	GUL	GGC	HC	ICI	CCG	CAG	GAA	GAA	TTG	CAG	GCC	Arg AGĞ AGĞ	CTC	Ser TCC TCT	622
GGI	Val GTA GTA	AIC	GCT	AAC	CAA	Asp GAC []] GAC	CCT	ATC	GCT	GTC	111	111	CGA CTA	AAT AAT	ACA AAA	670
CCC CAC	ATA ATA	GAT GAT	TCA TCA	111			CTT []] CTT		TTT 			TAA AA	AGT AGT	ATT 	CCA CCA	718
111	TAA TAA	111		CAA CAA												733

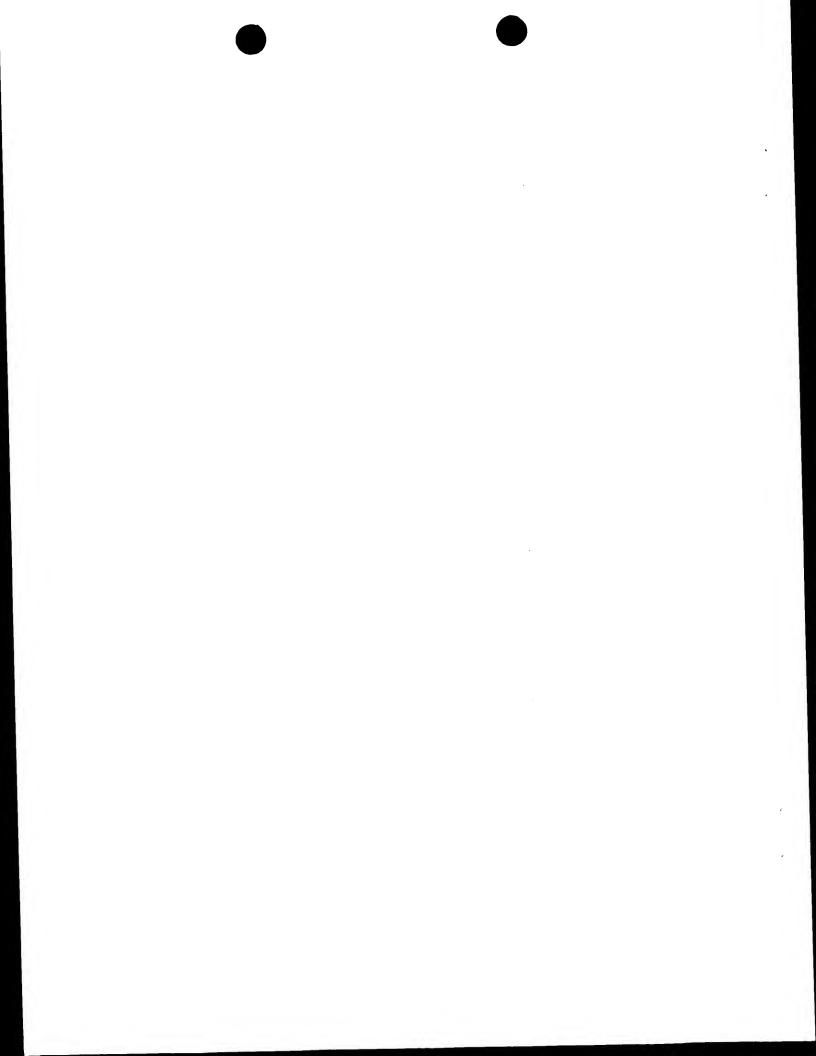


FIG. 15 SEGMENT F: (SEQ ID NOS:45-48)

AGTTTCCCCC CCCAACTTGT CGGAACTCTG GGCTCG	CGCG CAGGGCAGGA GCGGAGCGGC 60
GGCGGCTGCC CAGGCGATGC GAGCGCGGGC CGGACG	GTAA TCGCCTCTCC CTCCTCGGGC 120
TGCGAGCGCG CCGGACCGAG GCAGCGACAG GAGCGGA	ACCG CGGCGGGAAC CGAGGACTCC 180
CCAGCGGCGC GCCAGCAGGA GCCACCCCGC GAGNCG	TGCG ACCGGGACGG AGCGCCCGCC 240
AGTCCCAGGT GGCCCGGACC GCACGTTGCG TCCCCGG	CGCT CCCCGCCGGC GACAGGAGAC 300
GCTCCCCCC ACGCCGCGC CGCCTCGGCC CGGTCGC	
AAACTTTTCC CGAAGCCGAT CCCAGCCCTC GGACCCAAAACTTTTCC CAAACCCGAT CCGAGCCCTT GGACCAA	11 111 111
CGGGAGCCGT CCGCGCAGAG CGTGCACTTC TCGGGCC	
Glu Gly Lys Gly Lys Gly Lys Gly Gly Lys GAA GGC AAA GGC AAG GGG AAG GGC GGC AAG LII LII LII LII LII LII GAA GGC AGA GGC AAA GGG AAG GGC AAG AAG	
Lys Lys Pro Val Pro Ala Ala Gly Gly Pro AAG AAG CCC GTG CCC GCG GCT GGC GGC CCG III III II II II II II AAG AAG CCG GAG TCC GCG GCG GGC AGC CAG	

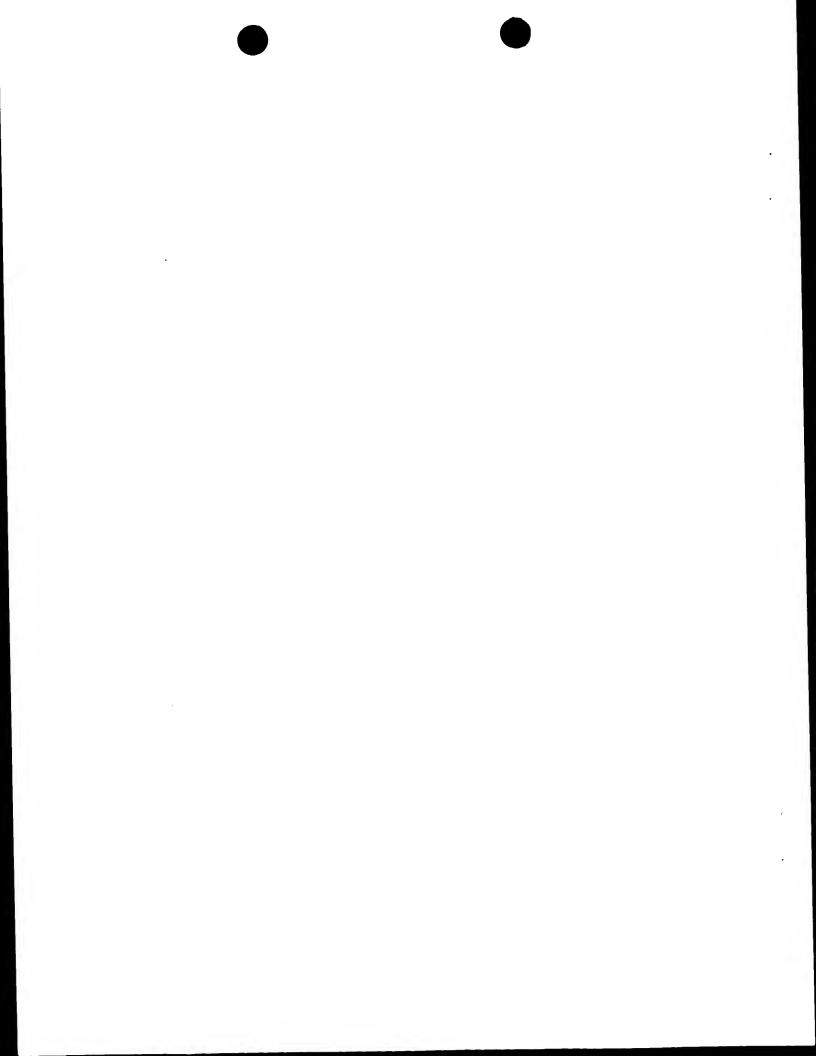


FIG. 16A (SEQ ID NOS:49-50)

G AV	AG 10 ys Se l	LA GA er G	lu Le	II C(eu Ai	aC A ng I	II A(le Se	ac A/ er Ly	AA G(ys A	la Se	CA C er Le 10	TG G(eu A	CT GA	AT TO Sp So	er G	GA GAA ly Glu 15	49
TAT Tyr	ATG Met	TGC Cys	AAA Lys 20	GTG Val	ATC Ile	AGC Ser	AAA Lys	CTA Leu 25	GGA Gly	AAT Asn	GAC Asp	AGT Ser	GCC Ala 30	TCT Ser	GCC Ala	97
AAC Asn	ATC Ile	ACC Thr 35	ATT Ile	GTG Val	GAG G1u	TCA Ser	AAC Asn 40	GCC Ala	ACA Thr	TCC Ser	ACA Thr	TCT Ser 45	ACA Thr	GCT Ala	GGG Gly	145
ACA Thr	AGC Ser 50	CAT His	CTT Leu	GTC Val	AAG Lys	TGT Cys 55	GCA Ala	GAG G1u	AAG Lys	GAG Glu	AAA Lys 60	ACT Thr	TTC Phe	TGT Cys	GTG Val	193
AAT Asn 65	GGA Gly	GGC Gly	GAC Asp	TGC Cys	TTC Phe 70	ATG Met	GTG Val	AAA Lys	GAC Asp	CTT Leu 75	TCA Ser	AAT Asn	CCC Pro	TCA Ser	AGA Arg 80	241
TAC Tyr	TTG Leu	TGC Cys	AAG Lys	TGC Cys 85	CAA Gln	CCT Pro	GGA Gly	TTC Phe	ACT Thr 90	GGA Gly	GCG Ala	AGA Arg	TGT Cys	ACT Thr 95	GAG Glu	289
AAT Asn	GTG Val	CCC Pro	ATG Met 100	AAA Lys	GTC Val	CAA Gln	ACC Thr	CAA Gln LO5	GAA Glu	AAA Lys	GCG Ala	Glu	GAG Glu 110	CTC Leu	TAC Tyr	337
CAG Gln	AAG Lys	AGA Arg 115	GTG Val	CTC Leu	ACC Thr	ATT Ile	ACC Thr 120	GGC Gly	ATT Ile	TGC Cys	ATC Ile	GCG Ala 125	CTG Leu	CTC Leu	GTG Val	385
GTT Val	GGC Gly 130	ATC Ile	ATG Met	TGT Cys	GTG Val	GTG Val 135	GTC Val	TAC Tyr	TGC Cys	AAA Lys	ACC Thr 140	AAG Lys	AAA Lys	CAA Gln	CGG Arg	433
AAA Lys 145	AAG Lys	CTT Leu	CAT His	GAC Asp	CGG Arg 150	CTT Leu	CGG Arg	CAG Gln	AGC Ser	CTT Leu 155	CGG Arg	TCT Ser	GAA Glu	AGA Arg	AAC Asn 160	481
ACC Thr	ATG Met	ATG Met	AAC Asn	GTA Val 165	GCC Ala	AAC Asn	GGG Gly	CCC Pro	CAC His 170	CAC His	CCC Pro	AAT Asn	CCG Pro	CCC Pro 175	CCC Pro	529
GAG Glu	AAC Asn	GTG Val	CAG Gln 180	CTG Leu	GTG Val	AAT Asn	CAA Gln	TAC Tyr 185	GTA Val	TCT Ser	AAA Lys	AAT Asn	GTC Val 190	ATC Ile	TCT Ser	577

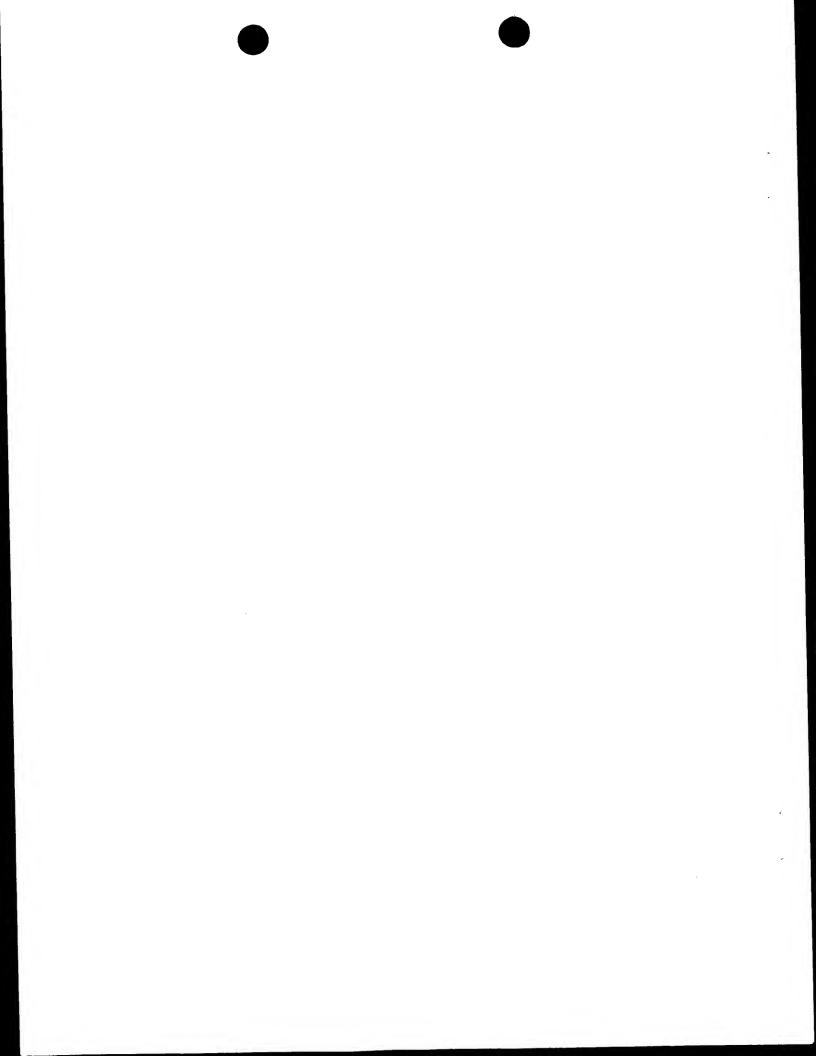


FIG. 16B

						AGA Arg										625	
						CAT His 215										673	
						GGA Gly										721	
						TCA Ser										769	
CCG Pro	ACT Thr	GGG Gly	GGC Gly 260	CCG Pro	AGA Arg	GGA Gly	CGT Arg	CTC Leu 265	AAT Asn	GGC Gly	TTG Leu	GGA Gly	GGC Gly 270	CCT Pro	CGT Arg	817	
GAA Glu	TGT Cys	AAC Asn 275	AGC Ser	TTC Phe	CTC Leu	AGG Arg	CAT His 280	GCC Ala	AGA Arg	GAA Glu	ACC Thr	CCT Pro 285	GAC Asp	TCC Ser	TAC Tyr	865	ı
						GAA Glu 295										913	'
						TCC Ser										961	
						TCC Ser										1009	Į
ACC Thr	CCT Pro	TGG Trp	CCT Pro 340	TTA Leu	GGA Gly	AGG Arg	TAT Tyr	GTA Val 345	TCA Ser	GCA Ala	ATG Met	ACC Thr	ACC Thr 350	CCG Pro	GCT Ala	1057	
CGT Arg	ATG Met	TCA Ser 355	CCT Pro	GTA Val	GAT Asp	TTC Phe	CAC His 360	ACG Thr	CCA Pro	AGC Ser	TCC Ser	CCC Pro 365	AAG Lys	TCA Ser	CCC Pro	1105	,
CCT Pro	TCG Ser 370	GAA Glu	ATG Met	TCC Ser	CCG Pro	CCC Pro 375	GTG Val	TCC Ser	AGC Ser	ACG Thr	ACG Thr 380	GTC Val	TCC Ser	ATG Met	CCC Pro	1153	;

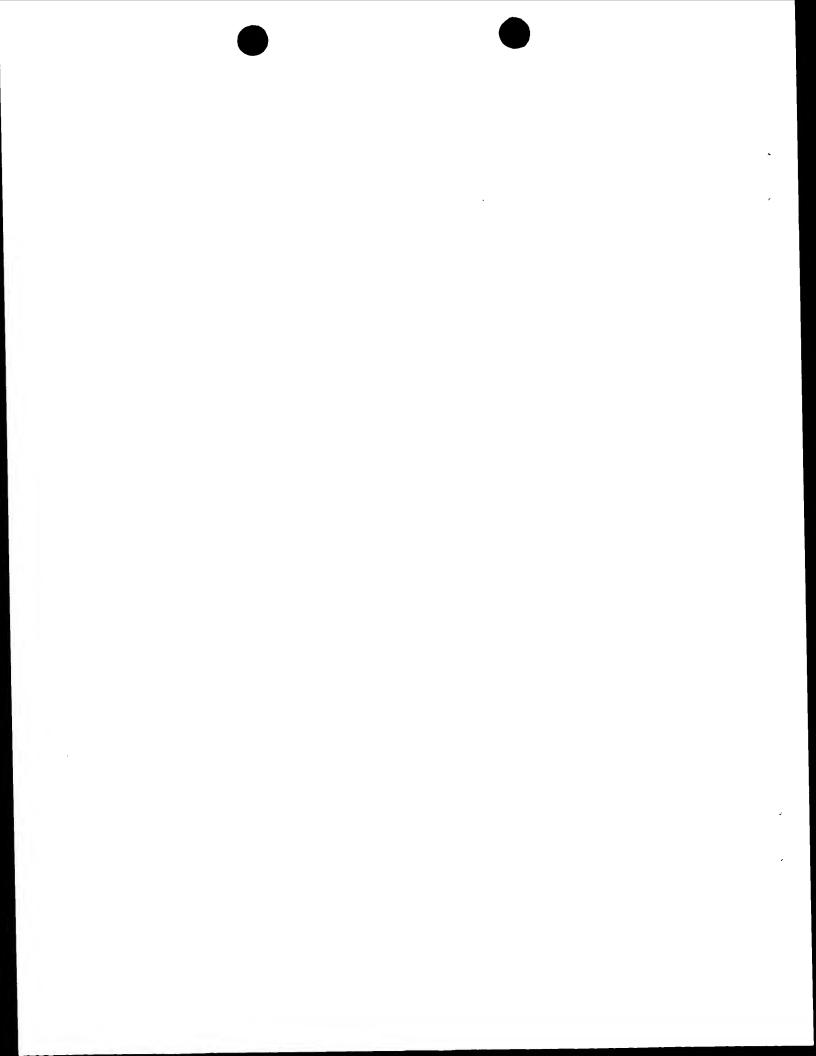


FIG. 16C

	TCC Ser 385	ATG Met	GCG Ala	GTC Val	AGT Ser	CCC Pro 390	TTC Phe	GTG Val	GAA Glu	GAG G1u	GAG G1u 395	AGA Arg	CCC Pro	CTG Leu	CTC Leu	CTT Leu 400	1201
	GTG Val	ACG Thr	CCA Pro	CCA Pro	CGG Arg 405	CTG Leu	CGG Arg	GAG Glu	AAG Lys	TAT Tyr 410	GAC Asp	CAC His	CAC His	GCC Ala	CAG Gln 415	CAA G1n	1249
	TTC Phe	AAC Asn	-TCG Ser	TTC Phe 420	CAC His	TGC Cys	AAC Asn	CCC Pro	GCG Ala 425	CAT His	GAG Glu	AGC Ser	AAC Asn	AGC Ser 430	CTG Leu	CCC Pro	1297
	CCC Pro	AGC Ser	CCC Pro 435	TTG Leu	AGG Arg	ATA Ile	GTG Val	GAG Glu 440	GAT Asp	GAG Glu	GAA Glu	TAT Tyr	GAA G1u 445	ACG Thr	ACC Thr	CAG Gln	1345
	GAG Glu	TAC Tyr 450	GAA Glu	CCA Pro	GCT Ala	CAA Gln	GAG Glu 455	CCG Pro	GTT Val	AAG Lys	AAA Lys	CTC Leu 460	ACC Thr	AAC Asn	AGC Ser	AGC Ser	1393
	CGG Arg 465	CGG Arg	GCC Ala	AAA Lys	AGA Arg	ACC Thr 470	AAG Lys	CCC Pro	AAT Asn	GGT Gly	CAC His 475	ATT Ile	GCC Ala	CAC His	AGG Arg	TTG Leu 480	1441
	GAA Glu	ATG Met	GAC Asp	AAC Asn	AAC Asn 485	ACA Thr	GGC Gly	GCT Ala	GAC Asp	AGC Ser 490	AGT Ser	AAC Asn	TCA Ser	GAG Glu	AGC Ser 495	GAA G1u	1489
	ACA Thr	GAG G1u	GAT Asp	GAA G1u 500	AGA Arg	GTA Val	GGA Gly	GAA Glu	GAT Asp 505	ACG Thr	CCT Pro	TTC Phe	CTG Leu	GCC Ala 510	ATA Ile	CAG G1n	1537
	AAC Asn	CCC Pro	CTG Leu 515	GCA Ala	GCC Ala	AGT Ser	CTC Leu	GAG G1u 520	GCG Ala	GCC Ala	CCT Pro	GCC Ala	TTC Phe 525	CGC Arg	CTG Leu	GTC Val	1585
•	GAC Asp	AGC Ser 530	AGG Arg	ACT Thr	AAC Asn	CCA Pro	ACA Thr 535	GGC Gly	GGC Gly	TTC Phe	Ser	CCG Pro 540	CAG Gln	GAA Glu	GAA Glu	TTG Leu	1633
١	CAG G1n 545	GCC Ala	AGG Arg	CTC Leu	TCC Ser	GGT Gly 550	GTA Val	ATC Ile	GCT Ala	Asn	CAA G1n 555	GAC Asp	CCT Pro	ATC Ile	GCT Ala	GTC Val 560	1681
•	TAAA	ACCG	iAA A	TACA	CCCA	T AG	ATTC	ACCT	GTA	AAAC	П	ATTT	TATA	TA A	TAAA	GTATT	1741
CCACCTTAAA TTAAACAAAA AAA									1764								

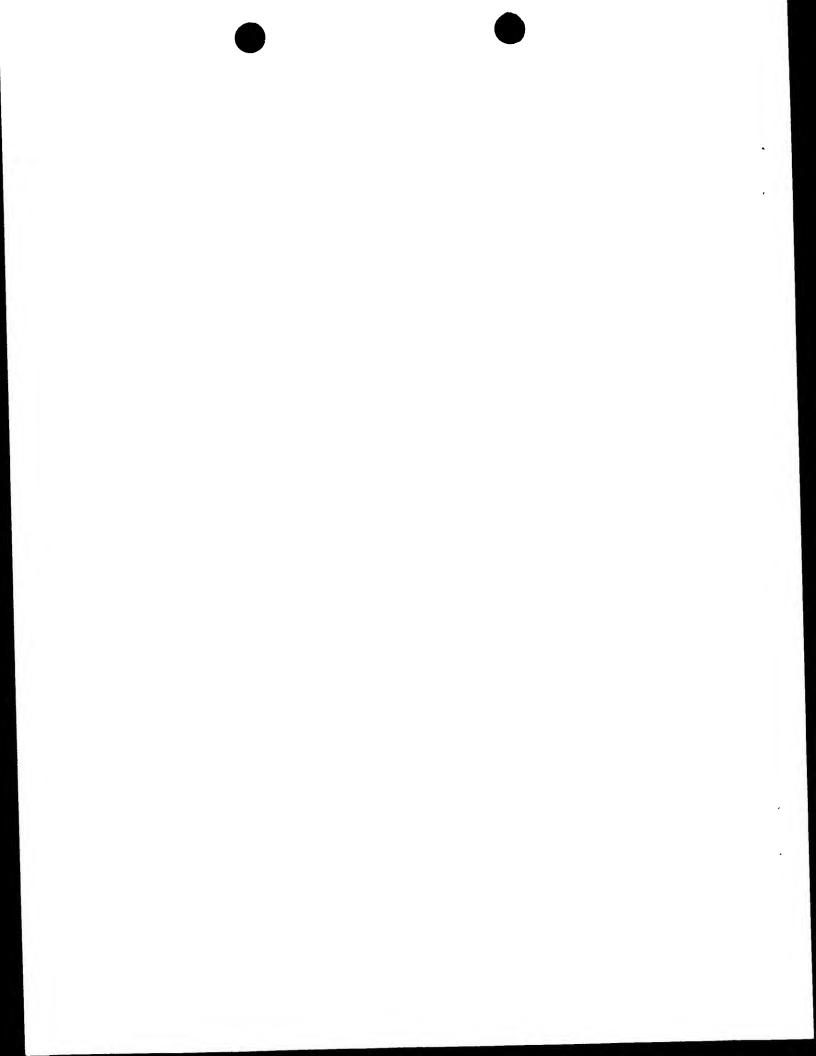


FIG. 17A (SEQ ID NOS:51-52)

CAT His 1	CAA Gln	GTG Val	TGG Trp	GCG Ala 5	GCG Ala	AAA Lys	GCC Ala	GGG Gly	GGC Gly 10	TTG Leu	AAG Lys	AAG Lys	GAC Asp	TCG Ser 15	CTG Leu	4	18
CTC Leu	ACC Thr	GTG Val	CGC Arg 20	CTG Leu	GGC Gly	GCC Ala	TGG Trp	GGC Gly 25.	CAC His	CCC Pro	GCC Ala	TTC Phe	CCC Pro 30	TCC Ser	TGC Cys	9	96
GGG Gly	CGC Arg	CTC Leu 35	AAG Lys	GAG Glu	GAC Asp	AGC Ser	AGG Arg 40	TAC Tyr	ATC Ile	TTC Phe	TTC Phe	ATG Met 45	GAG Glu	CCC Pro	GAG G1u	14	14
GCC Ala	AAC Asn 50	AGC Ser	AGC Ser	GGC Gly	GGG Gly	CCC Pro 55	GGC Gly	CGC Arg	CTT Leu	CCG Pro	AGC Ser 60	CTC Leu	CTT Leu	CCC Pro	CCC Pro	19	12
TCT Ser 65	CGA Arg	GAC Asp	GGG Gly	CCG Pro	GAA Glu 70	CCT Pro	CAA Gln	GAA Glu	GGA Gly	GGT Gly 75	CAG Gln	CCG Pro	GGT Gly	GCT Ala	GTG Val 80	24	0
CAA Gln	CGG Arg	TGC Cys	GCC Ala	TTG Leu 85	CCT Pro	CCC Pro	CGC Arg	TTG Leu	AAA Lys 90	GAG G1u	ATG Met	AAG Lys	AGT Ser	CAG Gln 95	GAG Glu	28	8
TCT Ser	GTG Val	GCA Ala	GGT Gly 100	TCC Ser	AAA Lys	CTA Leu	GTG Val	CTT Leu 105	CGG Arg	TGC Cys	GAG Glu	ACC Thr	AGT Ser 110	TCT Ser	GAA Glu	33	6
TAC Tyr	TCC Ser	TCT Ser 115	CTC Leu	AAG Lys	TTC Phe	AAG Lys	TGG Trp 120	TTC Phe	AAG Lys	AAT Asn	GGG Gly	AGT Ser 125	GAA Glu	TTA Leu	AGC Ser	38	4
CGA Arg	AAG Lys 130	AAC Asn	AAA Lys	CCA Pro	GAA Glu	AAC Asn 135	ATC Ile	AAG Lys	ATA Ile	CAG G1n	AAA Lys 140	AGG Arg	CCG Pro	GGG Gly	AAG Lys	43.	2
TCA Ser 145	GAA Glu	CTT Leu	CGC Arg	ATT Ile	AGC Ser 150	AAA Lys	GCG Ala	TCA Ser	CTG Leu	GCT Ala 155	GAT Asp	TCT Ser	GGA Gly	GAA Glu	TAT Tyr 160	48	0
ATG Met	TGC Cys	AAA Lys	GTG Val	ATC Ile 165	AGC Ser	AAA Lys	CTA Leu	GGA Gly	AAT Asn 170	GAC Asp	AGT Ser	GCC Ala	TCT Ser	GCC Ala 175	AAC Asn	528	8

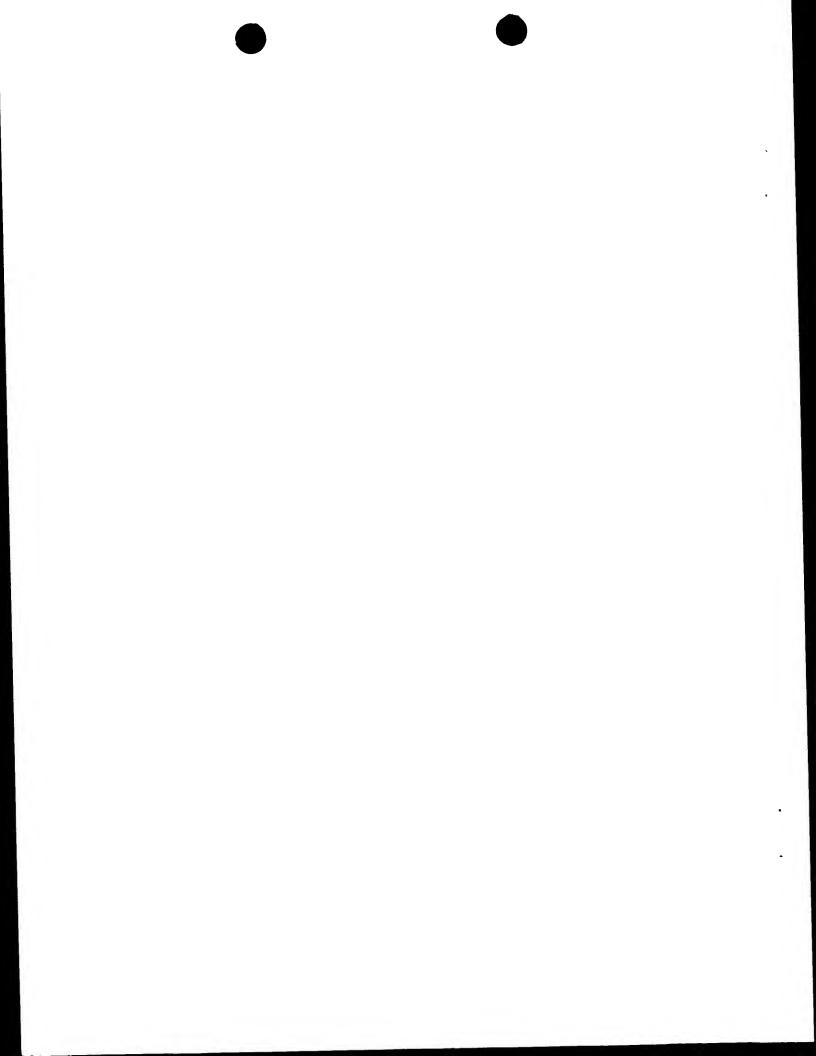


FIG. 17B

ATC ACC ATT GTG GAG TCA AAC GCC ACA TCC ACA TCT ACA GCT GGG ACA Ile Thr Ile Val Glu Ser Asn Ala Thr Ser Thr Ser Thr Ala Gly Thr 180 185 190	576
AGC CAT CTT GTC AAG TGT GCA GAG AAG GAG AAA ACT TTC TGT GTG AAT Ser His Leu Val Lys Cys Ala Glu Lys Glu Lys Thr Phe Cys Val Asn 195	624
GGA GGC GAG TGC TTC ATG GTG AAA GAC CTT TCA AAT CCC TCA AGA TAC Gly Gly Glu Cys Phe Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr 210	672
TTG TGC AAG TGC CAA CCT GGA TTC ACT GGA GCG AGA TGT ACT GAG AAT Leu Cys Lys Cys Gln Pro Gly Phe Thr Gly Ala Arg Cys Thr Glu Asn 235	720
GTG CCC ATG AAA GTC CAA ACC CAA GAA AAG TGC CCA AAT GAG TTT ACT Val Pro Met Lys Val Gln Thr Gln Glu Lys Cys Pro Asn Glu Phe Thr 245	768
GGT GAT CGC TGC CAA AAC TAC GTA ATG GCC AGC TTC TAC AGT ACG TCC Gly Asp Arg Cys Gln Asn Tyr Val Met Ala Ser Phe Tyr Ser Thr Ser 260 265	816
ACT CCC TTT CTG TCT CTG CCT GAA TAGCGCATCT CAGTCGGTGC CGCTTTCTTG Thr Pro Phe Leu Ser Leu Pro Glu 275 280	870
TTGCCGCATC TCCCCTCAGA TTCCNCCTAG AGCTAGATGC GTTTTACCAG GTCTAACATT	930
GACTGCCTCT GCCTGTCGCA TGAGAACATT AACACAAGCG ATTGTATGAC TTCCTCTGTC	990
CGTGACTAGT GGGCTCTGAG CTACTCGTAG GTGCGTAAGG CTCCAGTGTT TCTGAAATTG	1050
ATCTTGAATT ACTGTGATAC GACATGATAG TCCCTCTCAC CCAGTGCAAT GACAATAAAG	1110
GCCTTGAAAA GTCAAAAAAA AAAAAAAAAA	1140

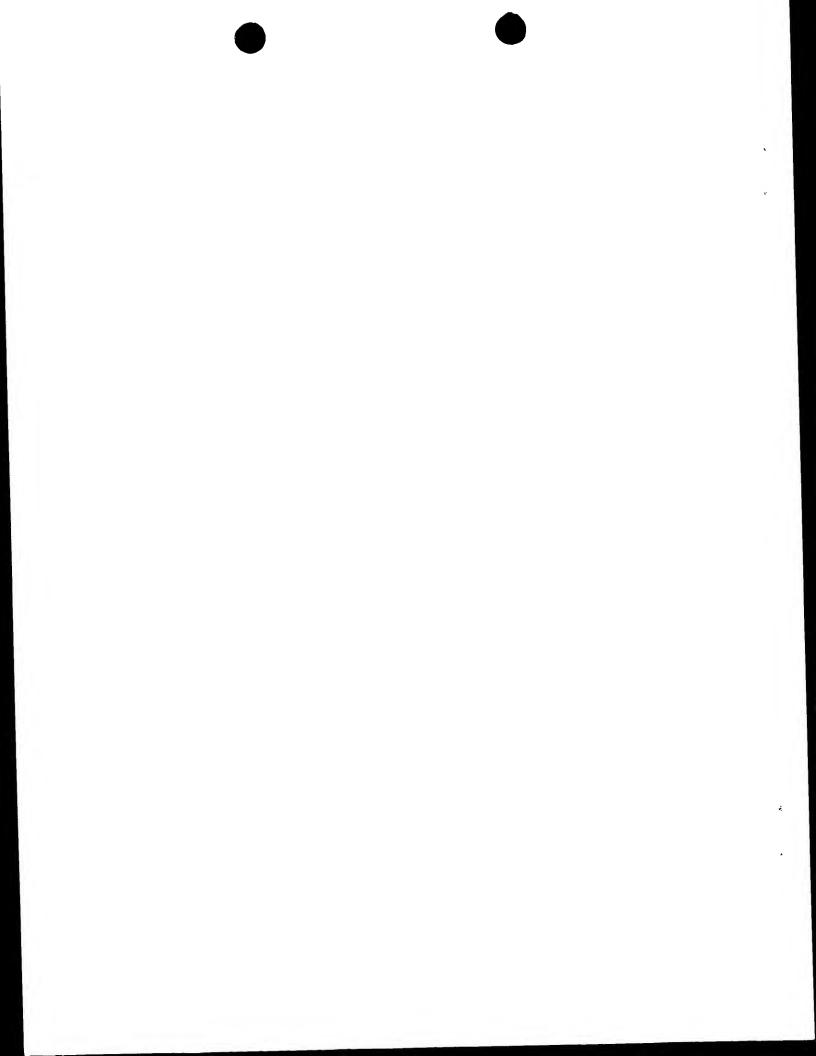


FIG. 18A (SEQ ID NOS:53-54)

Adilicul	L LLLA	ACTIGI	CGGAA	CICIO	וטט ג	J166	JGCG	CAG	iGCA(GGA (GCGG/	AGCGG	C	60
GGCGGCTGC	CC CAGG	CGATGC	GAGCG	CGGG	C CG(GACG	GTAA	TCG	CCTC	TCC	CTCC	TCGGG	2	120
TGCGAGCGC	CG CCGG/	ACCGAG	GCAGC	GACA	G GA	GCGG/	ACCG	CGG	CGGG	AAC (CGAG	GACTC	2	180
CCAGCGGCG	GC GCCA	GCAGGA	GCCAC	CCCG	C GAO	GCGT(GCGA	CCG	GGAC	GGA (GCGC	CCGCC	4	240
GTCCCAGGT	G GCCC	GGACCG	CACGT	TGCG	r cc	CCGC	GCTC	CCCC	GCCG	GCG /	4CAG(GAGAC	ã	300
CTCCCCCCC	CA CGCCC	GCGCGC	GCCTC	GGCC	C GG	rcgc ⁻	rggc	CCG	CCTC	CAC 1	TCCG	GGGAC/	4	360
AACTTTTCC	CC GAAG	CCGATC	CCAGC	CCTC	G GA	CCCAA	V ACT	TGT	CGCG	CGT (CGCC	TTCGC	2	420
GGGAGCCGT	C CGCG	CAGAGC	GTGCA	СТТСТ	r cg(GCG/					GC A(ng Ai			473
GAA GGC A Glu Gly L	AA GGC ys Gly	AAG GG Lys Gl 10	G AAG y Lys	GGC Gly	GGC Gly	AAG Lys 15	AAG Lys	GAC Asp	CGA Arg	GGC Gly	TCC Ser 20	GGG Gly		521
AAG AAG C Lys Lys P	CCC GTG Pro Val 25	CCC GC Pro Al	G GCT a Ala	Gly	GGC Gly 30	CCG Pro	AGC Ser	CCA Pro	Ala	TTG Leu 35	CCT Pro	CCC Pro	-	569
CGC TTG A Arg Leu L	AA GAG ys Glu 40	ATG AA Met Ly	G ATG s Ser	CAG Gln 45	GAG Glu	TCT Ser	GTG Val	GCA Ala	GGT Gly 50	TCC Ser	AAA Lys	CTA Leu		617
GTG CTT C Val Leu A 55	GG TGC arg Cys	GAG AC Glu Th	C AGT r Ser 60	TCT Ser	GAA Glu	TAC Tyr	TCC Ser	TCT Ser 65	CTC Leu	AAG Lys	TTC Phe	AAG Lys		665
TGG TTC A Trp Phe L 70	AG AAT ys Asn	GGG AG Gly Se 7	r Glu	TTA Leu	AGC Ser	CGA Arg	AAG Lys 80	AAC Asn	AAA Lys	CCA Pro	CAA G1n	AAC Asn 85		713
ATC AAG A Ile Lys I	TA CAG le Gln	AAA AG Lys Ar 90	G CCG g Pro	GGG Gly	AAG Lys	TCA Ser 95	GAA Glu	CTT Leu	CGC Arg	ATT Ile	AGC Ser 100	AAA Lys		761
GCG TCA C Ala Ser L	TG GCT eu Ala 105	GAT TC Asp Se	T GGA r Gly	GAA Glu	TAT Tyr 110	ATG Met	TGC Cys	AAA Lys	GTG Val	ATC Ile 115	AGC Ser	AAA Lys		809

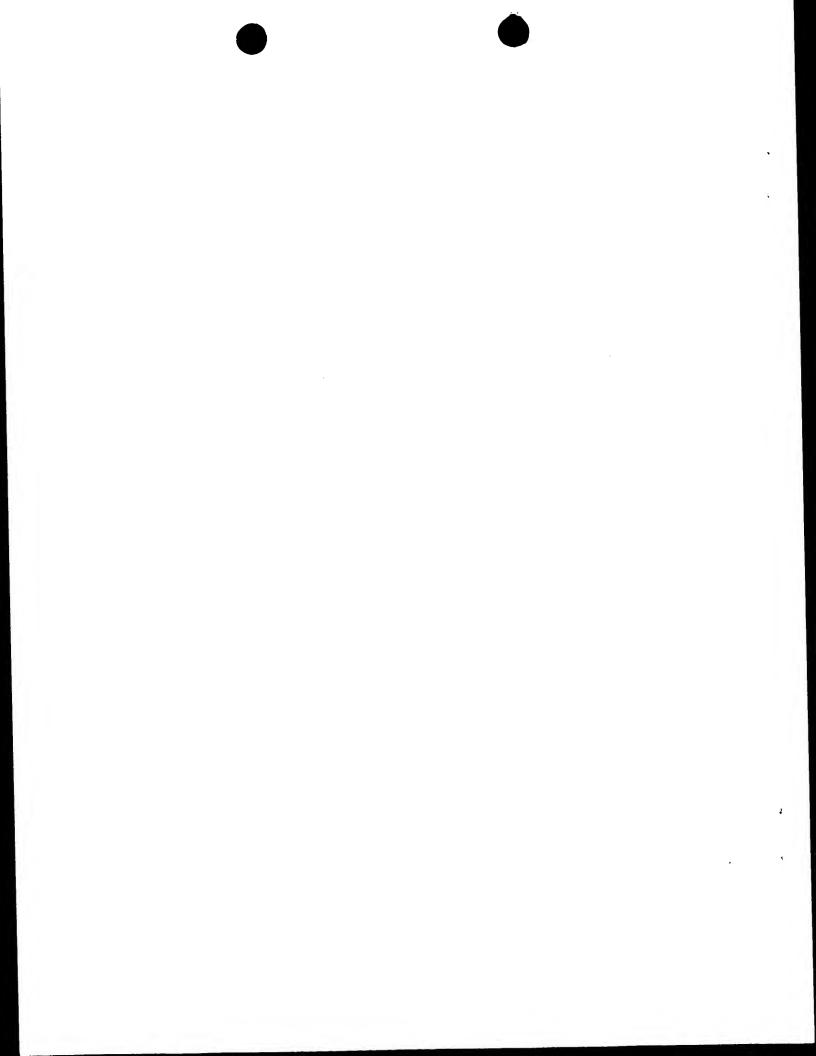
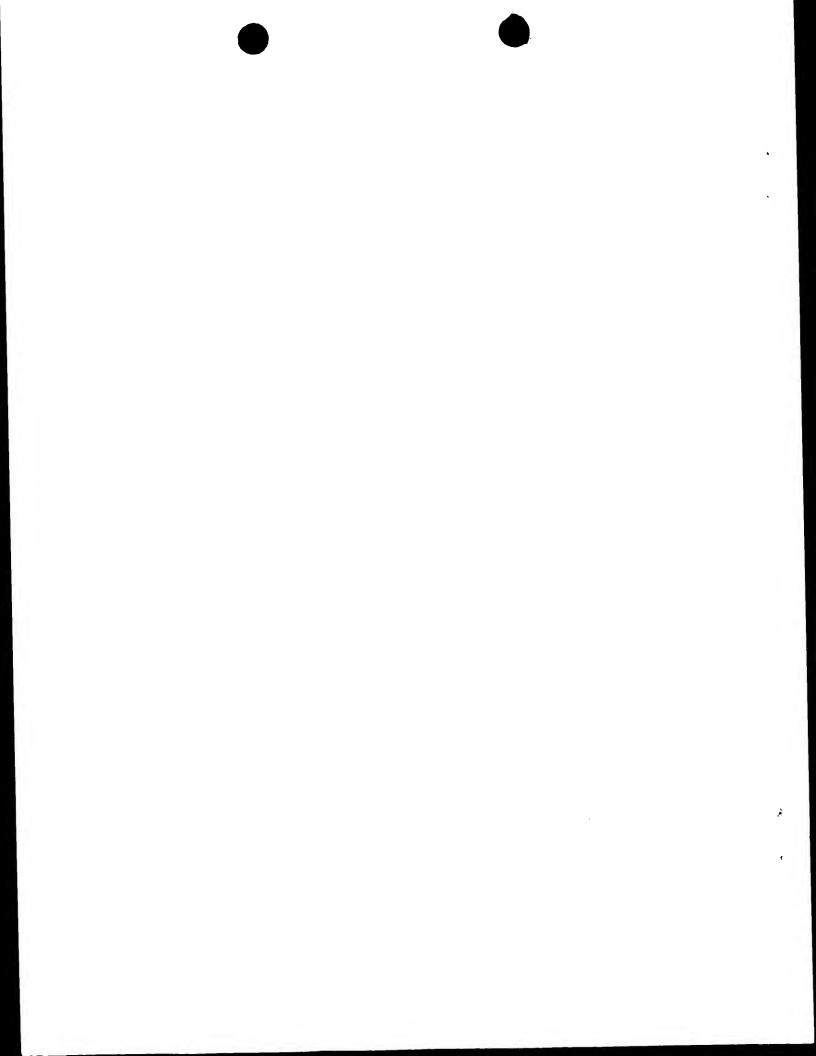


FIG. 18B

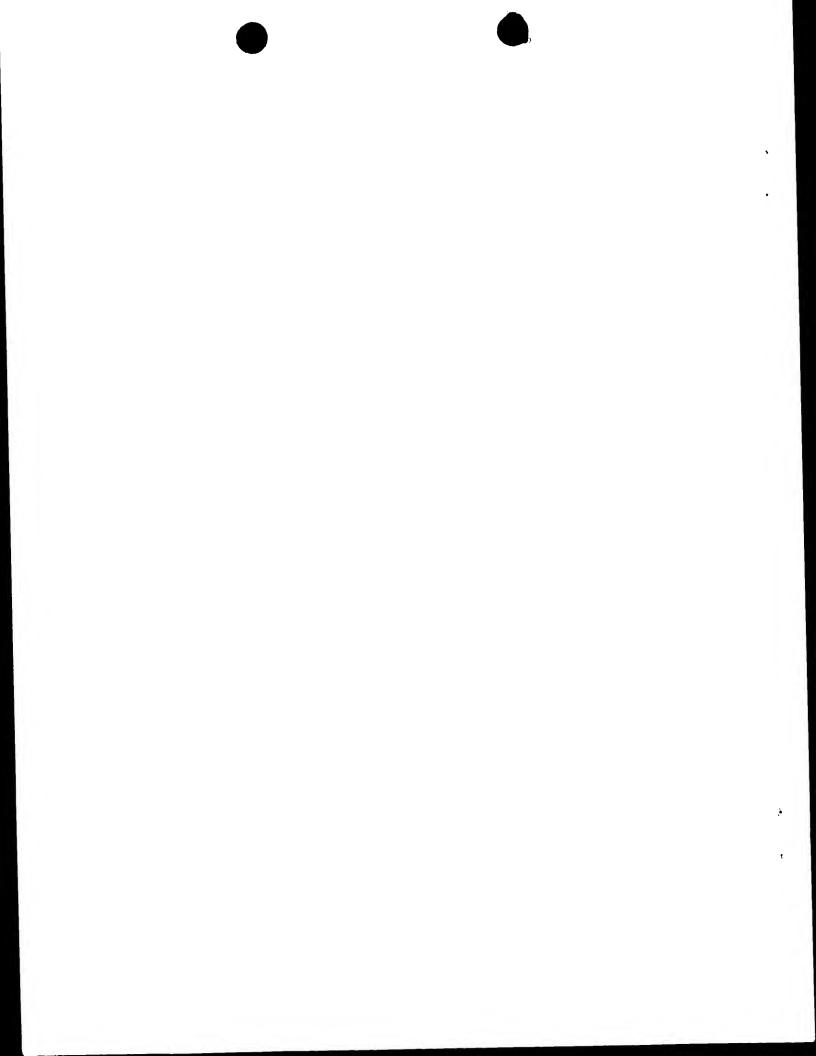
CTA GGA AAT GAC AGT GCC TCT GCC AAC ATC ACC ATT GTG GAG TCA AAC Leu Gly Asn Asp Ser Ala Ser Ala Asn Ile Thr Ile Val Glu Ser Asn 120 125 130	857
GAG ATC ACC ACT GGC ATG CCA GCC TCA ACT GAG ACA GCG TAT GTG TCT Glu Ile Thr Thr Gly Met Pro Ala Ser Thr Glu Thr Ala Tyr Val Ser 135	905
TCA GAG TCT CCC ATT AGA ATA TCA GTA TCA ACA GAA GGA ACA AAT ACT Ser Glu Ser Pro Ile Arg Ile Ser Val Ser Thr Glu Gly Thr Asn Thr 150 165	953
TCT TCA TCC ACA TCC ACA TCT ACA GCT GGG ACA AGC CAT CTT GTC AAG Ser Ser Ser Thr Ser Thr Ala Gly Thr Ser His Leu Val Lys 170 175 180	1001
TGT GCA GAG AAG GAG AAA ACT TTC TGT GTG AAT GGA GGC GAG TGC TTC Cys Ala Glu Lys Glu Lys Thr Phe Cys Val Asn Gly Gly Glu Cys Phe 185	1049
ATG GTG AAA GAC CTT TCA AAT CCC TCA AGA TAC TTG TGC AAG TGC CCA Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr Leu Cys Lys Cys Pro 200 210	1097
AAT GAG TTT ACT GGT GAT CGC TGC CAA AAC TAC GTA ATG GCC AGC TTC Asn Glu Phe Thr Gly Asp Arg Cys Gln Asn Tyr Val Met Ala Ser Phe 215	1145
TAC AGT ACG TCC ACT CCC TTT CTG TCT CTG CCT GAA TAGGCGCATG Tyr Ser Thr Ser Thr Pro Phe Leu Ser Leu Pro Glu 230 235 240	1191
CTCAGTCGGT GCCGCTTTCT TGTTGCCGCA TCTCCCCTCA GATTCAACCT AGAGCTAGAT	1251
GCGTTTTACC AGGTCTAACA TTGACTGCCT CTGCCTGTCG CATGAGAACA TTAACACAAG	1311
CGATTGTATG ACTTCCTCTG TCCGTGACTA GTGGGCTCTG AGCTACTCGT AGGTGCGTAA	1371
GGCTCCAGTG TTTCTGAAAT TGATCTTGAA TTACTGTGAT ACGACATGAT AGTCCCTCTC	1431
ACCCAGTGCA ATGACAATAA AGGCCTTGAA AAGTCTCACT TTTATTGAGA AAATAAAAAT	1491
CGTTCCACGG GACAGTCCCT CTTCTTTATA AAATGACCCT ATCCTTGAAA AGGAGGTGTG	1551
TTAAGTTGTA ACCAGTACAC ACTTGAAATG ATGGTAAGTT CGCTTCGGTT CAGAATGTGT	1611
TCTTTCTGAC AAATAAACAG AATAAAAAAA AAAAAAAAAA	1652

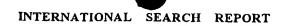


INTERNATIONAL SEARCH REPORT

International application No. PCT/US98/21349

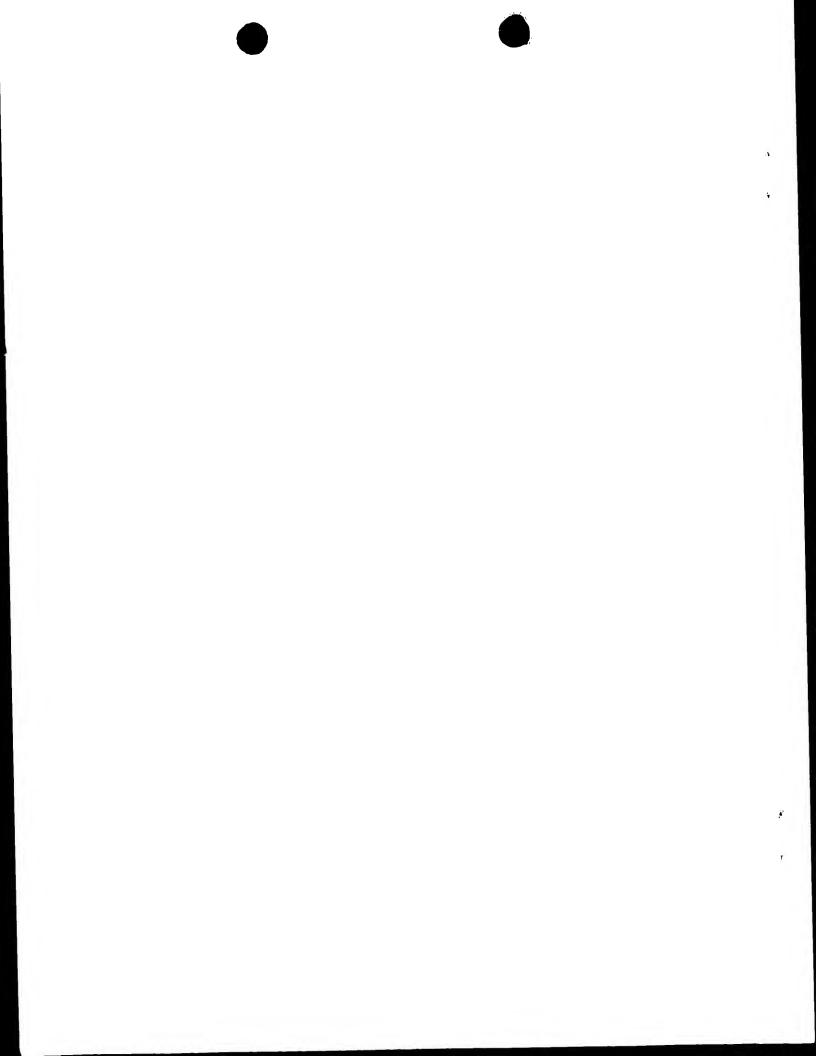
A. CLASSIFICATION OF SUBJECT MATTER IPC(6):A61K 31/70, 38/00, 38/02, 38/18 US CL:514/2, 12, 44, 903, 907 According to International Patent Classification (IPC) or to bo	th national classification and IPC									
B. FIELDS SEARCHED										
Minimum documentation searched (classification system followed by classification symbols)										
U.S. : 514/2, 12, 44, 903, 907										
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched										
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) Please See Extra Sheet.										
C. DOCUMENTS CONSIDERED TO BE RELEVANT										
Category* Citation of document, with indication, where	appropriate, of the relevant passages	Relevant to claim No.								
X US 5,530,109 A (GOODEARL et al. 3 to column 6, line 53 and column 11	US 5,530,109 A (GOODEARL et al.) 25 June 1996, column 3, line 3 to column 6, line 53 and column 11, line 1 to column 12, line 59.									
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International application No. PCT/US98/21349

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. X Claims Nos.: 35-36 because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
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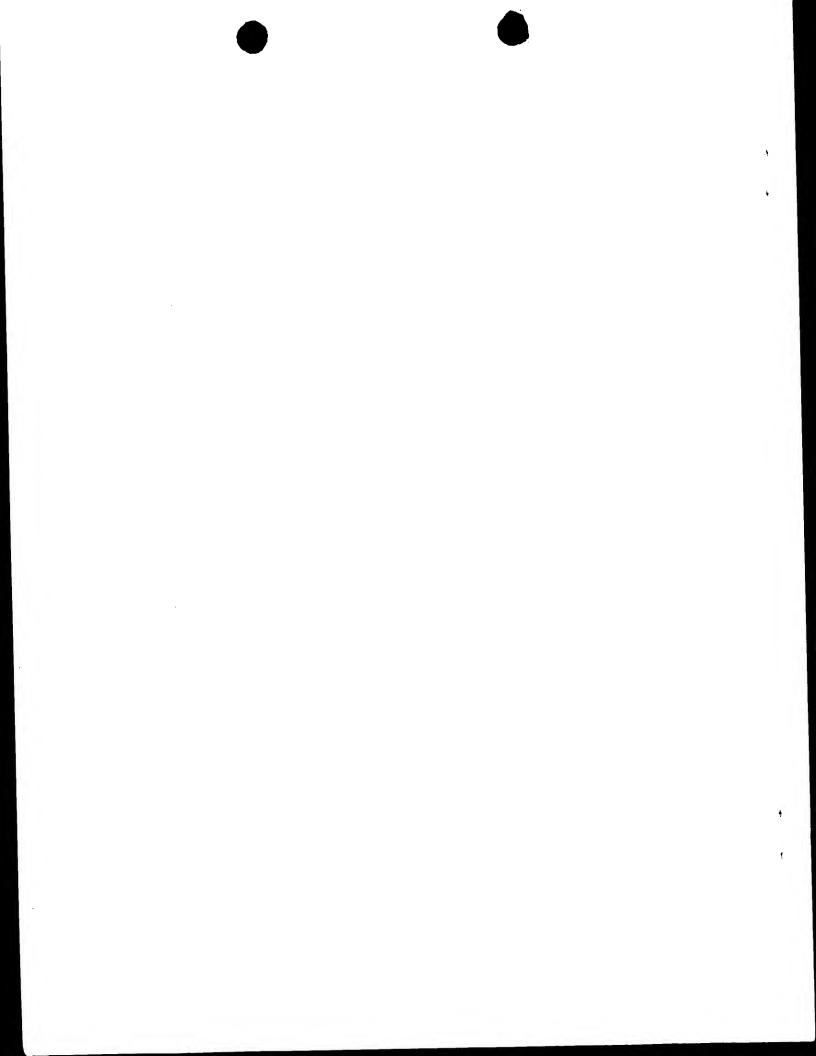


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(74) Agent: BUTLER, Gregory, B.; Cambridge NeuroScience, Inc., Building 700, One Kendall Square, Cambridge, MA 02139 (US). (81) Designated States: AM, AT, AU, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, JP, KE, KG, KP, LR, MG, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SI, SK, TJ, TT, UA, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published

With international search report.

(54) Title: METHODS OF TREATING DISORDERS OF THE EYE

(57) Abstract

The present invention relates to methods for the prophylaxis or treatment of retinal cells by the administration of a therapeutically effective amount of a neuregulin polypeptide.

FOR THE PURPOSES OF INFORMATION ONLY

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METHODS OF TREATING DISORDERS OF THE EYE

GOVERNMENT SUPPORT

5

10

This invention was made with the support of a federal grant from the U.S. Government (Grant No. 5ROINS28308-06). The Government has certain rights in the invention.

FIELD OF THE INVENTION

This invention relates to methods of affecting retinal cell function.

BACKGROUND OF THE INVENTION

15

20

The invention relates to prophylactic or affirmative treatment of diseases and disorders of retina and associated tissues of the eye by administering polypeptides found in vertebrate species, which polypeptides are growth, differentiation and survival factors for several cell types. Normal function of retinal cells including survival, proliferation, differentiation, and maintenance is dependent upon the controlled expression of a variety of peptide growth factors. Some of these factors can be produced by neuronal cells and by other cells of the retina, which provide a signal to regulate retinal cell function.

Anatomy and Function of the Retina

25

The retina is that component of the visual system which senses light and transmits impulses via the optic nerve to the visual cortex where the signals are deciphered and interpreted as images. The retina is comprised of a series of layers and cell types as illustrated in Figure 1.

30

35

The basic function of the retina is to transduce the visual image into a pattern of electrical potential changes that can be processed by the visual centers in the brain. The changes in electrical potentials in the retinal cells are then relayed to the brain. The structure of the retina reflects these functions (Figure 1). The cells of the retina are arrayed in three layers: (1) the outer nuclear layer, which contains the photoreceptor cells; (2) the inner nuclear layer, which contains the cell nuclei of most of the retinal interneurons and glia; and (3) the ganglion cell layer, which contains the cell bodies of the cells that relay the visual information to the brain via the optic nerve. In addition to these nuclear layers, there are three other distinct layers in the retina. The outermost layer is composed of the outer

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segments of the photoreceptor cells: this is where the actual process of light-to-electrical signal transduction take place. The outer plexiform layer lies between the outer and inner nuclear layers. It is made up of synapses between the terminals of the photoreceptors and the dendrites of the retinal interneurons of the inner nuclear layer. The inner plexiform layer lies between the inner nuclear layer and the ganglion cell layer. This layer is where the interneurons of the inner nuclear layer synapse with the retinal ganglion cell dendrites.

The retina is composed of five classes of neurons, and two classes of supporting cells (Principles of Neural Science, 3rd ed., Ed. by E.R. Kandel, J. H. Schwartz, and T. M. Jessell. Elsevier, New York, NY 1991). Of the neuronal types, the receptor cells are the cells that transduce light into electrical signals. Receptor cells are of two subtypes: cones which mediate form and color perception in daylight, and rods - which mediate form perception in dim light. Ganglion cells of the retina project axons into the brain via the optic nerve and are the output cells of the retina. The remaining neuronal types are interneurons that modulate retinal output: bipolar cells connect receptor cells to ganglion cells; horizontal cells mediate lateral interactions between receptors and bipolar cells; and amacrine cells mediate lateral interactions between bipolar cells and ganglion cells. The supporting cell types are the glial cells of the retina, Müller cells, and the pigment epithelium cells. The latter cell type plays an important role in the maintenance of receptor cells.

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The basic flow of information through the retina is as follows (Refer to Figure 1): (1) light passes through the cells of the retina and is absorbed by the outer segments of the photoreceptor cells; (2) the photons are transduced into potential changes in the photoreceptor cells; (3) this change in potential is relayed to one type of retinal interneuron in the inner nuclear layer, the bipolar cell, via synapses in the outer plexiform layer; (4) the bipolar cells relay the electrical potential changes to the ganglion cells through their synapses in the inner plexiform layer; and (5) the ganglion cells convert the potential changes into action potentials that are sent along the optic nerve to the brain. This process results in a pattern of action potentials in the optic nerves that reflects the pattern of light and dark in the visual world. Some initial processing of the visual information takes place in the retina before it is relayed to the other visual areas in the brain.

Proper development and maintenance of the retina is necessary for sustaining normal vision. Degeneration of components of the retina can lead to partial or total blindness.

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Peptide Growth Factors

The development and physiology of multicellular organisms requires multiple modes of intercellular communication. Such communication may be systemic, as in the case of hormones delivered via the bloodstream, or can be highly localized. In the latter case two modes are commonly recognized: synaptic signaling from neurons, and paracrine signaling from adjacent or nearby cells (*Molecular Biology of the Cell*. Alberts et al., 2nd ed. Garland Publishing, New York, NY 1989). A function of such signaling is to coordinate cell survival, proliferation, differentiation, and/or metabolic activity. The molecules that serve as transmitted signals vary in their chemical composition; one group of molecules are proteins, the peptide growth factors. Peptide growth factors act upon cells by binding to cell surface receptors. These receptors are coupled to intracellular signal transduction pathways that give rise to the above described activities when activated by growth factor binding. The genesis and differentiation of the varied retinal cell types and the generation of distinct layers in the retina from progenitor cells of the optic cup are the result of developmental events that are mediated by intercellular communication involving peptide growth factors.

Peptide Growth Factors in the retina

The roles of growth factors in the development and maintenance of the retina have been studied in cell culture, by molecular analysis of the expressed growth factors and their receptors, and in animal models of disease or injury.

As an example of *in vitro* studies, explants and partially-dissociated chick retinal pigmented epithelium (RPE) can trans-differentiate into neural retina in the presence of bFGF (Coulombre and Coulombre, *Dev. Biol.* 12:79, 1965). Proliferation of dissociated RPE cells is stimulated by α FGF, α FGF, EGF, PDGF, IGF, and insulin; and it is inhibited by TGFB (Sternfeld et al., *Curr. Eye Res.* 8: 1029, 1989; Leschey et al., *Invest. Ophthalmol. Vis. Sci.* 31: 839, 1990; Song and Lui, *J. Cell Physiol.* 143:196, 1990). Cultured RPE cells are induced by cytokines to release nitric oxide, which is cytotoxic--and the induction can be blocked by FGF (Goureau et al., *Biochem. Biophys Res. Comm.* 186:854, 1992; op. cit., 198: 120, 1994). Further, retinal explants from the *rd* mouse are rescued from cell death by combined treatment with NGF and bFGF (Caffe et al., *Curr. Eye Res.* 12:719, 1993).

The presence of growth factor receptors in retinal cells has been demonstrated by a variety of molecular analytical techniques, including immunostaining, in situ hybridization and tissue binding using radio-labeled ligands. Cells in the RPE express FGF receptors

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(Malecaze et al., J. Cell Physiol. 154: 1105, 1993). Ganglion cells and Müller cells express receptors for BDNF, CNTF, FGF, trkA and trkB (Jelsma et al., J. Neurobiol. 24:1207, 1993; Takahashi et al., Neurosci. Lett. 151:174, 1993; Carmignoto et al., Exp. Neurol. 111:302 191; reviewed in Steinberg, Curr. Opin. Neurobiol. 4:515, 1994). Müller cells also express PDGF receptors (Mudhar et al., Development 118: 539, 1993). Receptors for IGF are detected on photoreceptor cells (Waldbillig et al., Exp. Eye Res. 47:587 1988; Ocrant et al., Exp. Eye Res. 52:581, 1991), and depending on the species and developmental stage that are analyzed receptors for bFDF have been localized on several cell types, including retinal ganglion cells (Sternfeld et al., Curr. Eye Res. 8:1029, 1992; Schweigerer et al., Biochem Biophys. Res. Comm. 143:934, 1987).

Studies on retinal ganglion cell survival in vivo in animal models of optic nerve axotomy and retinal ischemia have demonstrated effects due to FGF (Sievers et al., Neurosci. Lett. 76:157, 1987), NGF (Carmignoto et al., J. Neurosci. 2:1263, 1989), CNTF (Mey and Thanos, Brain Res. 602:304, 1993), BDNF (Mansour-Robaey et al., PNAS USA 91:1632, 1994; Mey and Thanos, Brain Res. 602:304, 1993), NT4/5 (Cohen et al., J. Neurobiol. 25:953, 1994) and bFGF (Ferguson et al., J. Neurosci. 10:2176, 1990). Some undesirable retinal complications, including macrophage proliferation, inflammation, disorganization of retinal structure and angiogenesis are associated with treatment of the retina with several of the above factors.

Neuregulins

A recently described family of growth factors, the neuregulins (reviewed by Mudge, Curr. Biol. 3:361, 1993; Peles and Yarden, Bioessays 15:815, 1993), are synthesized by neurons (Marchionni et al. Nature 362:313, 1993) and by mesenchymal cells from several parenchymal organs (Meyer and Birchmeier, PNAS 91:1064, 1994). The neuregulins and related factors that bind p185erbB2 have been purified, cloned and expressed (Benveniste et al. PNAS, 82:3930, 1985; Kimura et al., Nature 348:257, 1990; Davis and Stroobant, J. Cell Biol. 110:1353, 1990; Wen et al., Cell 69:559, 1992; Yarden and Ullrich, Ann. Rev. Biochem. 57:443, 1988; Dobashi et al., Proc. Natl. Acad. Sci. 88:8582, 1991; Lupu et al., Proc. Natl. Acad. Sci. 89:2287, 1992; Wen et al., Mol. Cell. Biol. 14:1909, 1994). Recombinant neuregulins have been shown to be mitogenic for peripheral glia (Marchionni et al., Nature 362:313, 1993) and have been shown to influence the formation of the neuromuscular junction (Falls et al., Cell 72:801, 1993; Jo et al., Nature 373: 158, 1995; 35 Chu et al., Cell 14: 329, 1995).

The neuregulin gene consists of at least thirteen exons. The neuregulin transcripts are alternatively spliced and these encode many distinct peptide growth factors, which are referred to as the neuregulins (Marchionni et al., Nature 362:313, 1993). DNA sequence comparisons revealed that neu differentiation factor (NDF) (Wen et al., Cell 69:559, 1992) and heregulins (Holmes et al., Science 256:1205, 1992), which were purified as ligands of the p185erbB2 (also known as neu or HER2) receptor tyrosine kinase, also are splicing variants of the neuregulin gene. The acetylcholine receptor inducing activity (ARIA) also is a product of the neuregulin gene (Falls et al., Cell 72:801, 1993). Common structural features of the neuregulins are the presence of a single immunoglobulin-like (Ig) fold and a single epidermal growth factor-like (EGF) domain.

The sites of neuregulin gene expression have been characterized by use of nucleic acid probes to analyze RNA samples by a variety of methods, such as Northern blotting, RNase protection, or *in situ* hybridization. Transcripts have been detected in the nervous system and in a variety of other tissues (Holmes et al., *Science* 256:1205, 1992 Wen et al., *Cell* 69:559, 1992; Meyer and Birchmeier, *PNAS* 91:1064, 1994). Sites of gene expression have been localized in the brain and spinal chord and in other tissues. (Orr-Urteger et al., *PNAS* 90:1867, 1993; Falls et al., *Cell* 72:801, 1993; Marchionni et al., *Nature* 362:313, 1993; Meyer and Birchmeier, *PNAS* 91:1064, 1994; Chen et al., *J. Comp. Neurol.* 349; 389, 1994; Corfas et al., *Neuron* 14:103, 1995). Specifically in the retinal neurepithelium, expression of neuregulin transcripts has been detected at embryonic day 18 in rat (Meyer and Birchmeier, *PNAS* 91:1064, 1994).

Although a large number of neuregulins may be produced by alternative splicing, they can be broadly sorted into the putative membrane-bound and the soluble isoforms. The former contains a putative trans-membrane domain and may be presented at the cell surface. Membrane-anchored peptide growth factors may mediate cell-cell interactions through cell-adhesion or juxtacrine mechanisms (reviewed by Massagué and Pandiella, Ann. Rev. Biochem. 62:515, 1993). Alternatively, the putative membrane-bound isoforms may be cleaved from the cell surface and function as soluble proteins (Wen et al., Cell 69:559, 1992; Falls et al., Cell 72:801, 1993). The soluble neuregulin isoforms contain sequence corresponding to the extracellular domains of the putative membrane-bound isoforms, but terminate before the transmembrane domain. These neuregulin isoforms may be secreted, and hence could affect cells at a distance; or they may be present in the cytoplasm, but could be released upon cellular injury. In the latter case, neuregulins may function as injury factors, as has been postulated for the ciliary neurotrophic factor (Stöckli et al., Nature

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342:920, 1989). Any one of these modes of action of the neuregulins may occur in the retina.

Cellular targets of peptide growth factors are those which bear receptors for the factor(s) and those that are shown to respond in a bioassay either in vitro or in vivo. Based on studies demonstrating phosphorylation on tyrosine residues or cross-linking experiments, neuregulins are candidate ligands for the receptor tyrosine kinases p185erbB2 (or HER-2 in human), p185erbB3 (HER-3 in human), p185erbB4 (or HER-4 in human) or related members of the EGFR gene family. Collectively, these receptors can be referred to as erbB receptors. Though the precise ligand-receptor relationship of each neuregulin protein with each member of the EGFR family is yet to be clarified, several lines of evidence suggest that binding of ligands is mediated by either erbB3 and erbB4, but signaling occurs through either erbB2, erbB4 and heterodimers of the various subunits (e.g., Carraway and Cantley, Cell 78:5, 1994). These receptors are known to be present on Schwann cells and muscle cells (Jo et al., Nature 373: 158, 1995), and other neuregulin targets, such as cell lines derived from various tumor tissues, such as breast and gastric epithelia. Sites of expression of the HER-4 gene have been localized by in situ hybridization to several regions of the brain, including: hippocampus, dentate gyrus, neo cortex, medial habenula, reticular nucleus of the thalamus, and the amygdala (Lai and Lemke, Neuron 6:691, 1991). The distribution of the HER-4 receptor has not been studied by methods that allow detection of the protein or the activated receptor tyrosine kinase in vivo or in cultures of primary cells. The expression pattern of erbB2, erbB3 and erbB4 in the retina has not been described.

Neuregulins have been shown to have a variety of biological activities depending on the cell type being studied. Several neuregulins, including native bovine GGFI, II and III and recombinant human GGF2 (rhGGF2) are mitogenic for Schwann cells (Marchionni et al., Nature 362:313, 1993), as is heregulin B1 (Levi et al., I Neurosci. 15:1329, 1995). On human muscle culture, rhGGF2 has a potent trophic effect on myotubes (Sklar et al., U.S. Pat. Applic. # 08/059, 022). The differentiation response to rhGGF2 also includes induction of acetylcholine receptors in cultured myotubes (Jo et al., Nature 373: 158, 1995). This activity is associated with other forms of neuregulin, including ARIA (Falls et al., Cell 72:801, 1993) and heregulin B1 (Chu et al., Neuron 14:329, 1995), as well as with rhGGF2. Further, ARIA has been shown to induce synthesis of voltage-gated sodium channels in chick skeletal muscle (Corfas and Fischbach, J. Neurosci. 13:2118, 1993). Glial growth factor (GGF), and more specifically rhGGF2, can restrict neural crest stem cells to differentiate into glial cells in vitro (Shah et al., Cell 77:349, 1994). Activities of neuregulin

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on retinal cells have not been described. In summary, there are examples of neuregulin proteins affecting proliferation, survival and differentiation of target cells.

Pharmaceutical need for treating disorders of the eye

A variety of retinal diseases and related disorders are known that produce impaired vision and in some cases progress to total blindness. These disorders of the eye include, but are not necessarily limited to: various retinopathies, such as hypertensive retinopathy, diabetic retinopathy and occlusive retinopathy; also injuries and disorders resulting in retinal degeneration, such as retinal tearing and detachment and inherited diseases, such as retinitis pigmentosa; also age-related macular degeneration; diseases of the optic nerve; glaucoma and retinal ischemia.

Diabetic Retinopathy is the leading cause of blindness in patients 25-74 years. It is responsible for 12,000-24,000 new cases of blindness per year in the United States. Of the 6 million diabetics in the US 50% show detectable retinopathy after 7 years of diabetes. Agerelated macular degeneration (ARMD) is estimated to be present in over 9% of the population 52 years and older and in 33% of the population 75 years and older. Glaucoma is associated with chronically high intraocular pressure and approximately 2 million people in the US are currently being treated. In the US approximately 100,000 people are blinded each year by glaucoma.

There is precedent for the use of growth factors that have been shown to be active on retinal cultures in the treatment of retinal degenerative diseases. FGF supports the survival of photoreceptor cells in culture and has been injected into the extracellular space surrounding the rods and cones or into the vitreous body to rescue the photoreceptors in rats which have degeneration as a result of light damage or because of an inherited disease (LaVail et al, PNAS 89: 11249, 1992, Faktorovich et al J. NeuroSci 12: 3554, 1992). Similarly TGFB2 has been used for the treatment of Macular holes in humans. The TGFB used was derived from bovine sources and was administered by directly infusing the factor into the area of the macular hole (Glaser et al., Opthalmol. 92: 1162, 1992).

Currently, there are limited options for therapy for the promotion of retinal cell function, including survival, proliferation, differentiation, growth and changes in gene activity and metabolic activity. Such a therapy would be useful for treatment of a variety of eye disorders resulting in loss of sight.

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SUMMARY OF THE INVENTION

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In general, the present invention provides methods for promoting the function of retinal cells using neuregulins. A novel aspect of the invention involves the use of neuregulins as growth factors to promote survival of retinal cells. Treating of the retinal cells to provide these effects may be achieved by contacting retinal cells with a polypeptide described herein. The treatments may be provided to slow or halt net cell loss or to increase the amount or quality of retinal tissue present in the vertebrate.

Neuregulins are a family of protein factors heretofore described as glial growth factors, acetylcholine receptor inducing activity (ARIA), heregulins, neu differentiation factor, which are encoded by one gene. A variety of messenger RNA splicing variants (and their resultant proteins) are derived from this gene and many of these products show binding to and activation of erbB2 (neu) and closely related receptors erbB3 and erbB4. The invention provides methods for using all of the known products of the neuregulin gene, as well as, other not yet discovered splicing variants of the neuregulin gene. Thus, the above factors, regulatory compounds that induce synthesis of these factors, and small molecules which mimic the effect of these factors by binding to the receptors on retinal tissues or by stimulating through other means the second messenger systems activated by the ligand-receptor complex are all extremely useful as prophylactic and affirmative therapies for retinal tissue diseases and related disorders of the eye.

The survival of retinal cells as used herein refers to the prevention of loss of retinal cells by necrosis or apoptosis or the prevention of other mechanisms of retinal loss. Survival as used herein indicates a decrease in the rate of cell death of at least 10%, more preferably by at least 50%, and most preferably by at least 100% relative to an untreated control. The rate of survival may be measured by counting cells stainable with a dye specific for dead cells (such as propidium iodide) in culture.

Methods for treatment of diseases or disorders using the polypeptides or other compounds described herein are also part of the invention. Examples of retinal tissue disorders that may be treated include eye diseases and disorders resulting from sensorineural pathologies, such as loss of sight, which may also be treated using the methods of the invention. These disorders of the eye include, but are not necessarily limited to: various retinopathies, such as hypertensive retinopathy, diabetic retinopathy and occlusive retinopathy; also injuries and disorders resulting in retinal degeneration, such as retinal

tearing and detachment and inherited diseases, such as retinitis pigmentosa; also age-related macular degeneration; diseases of the optic nerve; glaucoma and retinal ischemia.

The methods of the invention make use of the fact that the various neuregulin proteins are encoded by the same gene. A variety of messenger RNA splicing variants (and their resultant proteins) are derived from this gene and many of these products show binding to p185erbB2 (or related receptors erbB3 and erbB4) and activation of the same. Products of this gene are used to show retinal cell survival activity (see Example 2, below). This invention provides a use for all of the known products of the neuregulin gene (described herein and in the references listed above), which have the stated activities as promoting retinal cell function. Most preferably, recombinant human GGF2 (rhGGF2) is used in these methods.

The invention also relates to the use of other, not yet naturally isolated, splicing variants of the neuregulin gene. Figure 12 shows the known patterns of splicing. These patterns are derived from polymerase chain reaction experiments (on reverse transcribed RNA), analysis of cDNA clones (as presented within) and from analysis of published sequences encoding neuregulins (Peles et al., Cell 69:205, 1992; Wen et al., Cell 69:559, 1992; Wen et al., Mol. Cell Biol. 14:1909, 1994) These patterns, as well as additional patterns disclosed herein, represent probable splicing variants which exist. The splicing variants are fully described in Goodearl et al., USSN 08/036,555, filed March 24, 1993, incorporated herein by reference.

Advantages of the present invention include the development of new therapeutic approaches to injury or diseases of the eye, more specifically degenerative diseases of the retina, based on the promotion of retinal cell function through the use of neuregulins. Loss of retinal cells is a common feature of degenerative eye diseases, and there are no available treatments, including growth factors, that prevent the death of retinal ganglion cells. The factor can be formulated for intraocular injection and administered to patients that suffer from degenerative disorders, which lead to loss of sight. Thus, this approach to therapy can halt or slow the progressive loss of sight, which ensues in various eye diseases.

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BRIEF DESCRIPTION OF THE FIGURES

Figure 1 shows the series of layers and cell types which form the retina: (1) corresponds to the ganglion cell layer; (2) corresponds to the inner plexiform layer; (3) corresponds to the inner nuclear layer; (4) corresponds to the outer plexiform layer; (5) corresponds to the outer nuclear layer.

Figure 2 is immunostaining showing that neuregulin protein is expressed in the retinal ganglion cell layer during embryonic retinal development. Arrows point to labeling in the developing ganglion cell layer.

Figure 3 is in situ hybridization showing that neuregulin mRNA is expressed in cells of the retinal ganglion cell layer during embryonic development. Arrows point to the labeling in the ganglion cell layer, showing that the distribution is similar to the neuregulin immunoreactivity shown in Figure 2.

Figure 4 is immunostaining showing that neuregulin protein is present in the inner and outer plexiform layers of the adult retina.

Figure 5 is immunostaining showing that TUJ1 immunoreactivity is expressed in the newborn rat retina and shows that retinal ganglion cells are the primary cell class that expresses this antigen at this stage of development. The retinal ganglion cell layer is marked with large arrows, while the labeled amacrine cells are marked with the small arrows and the labeled horizontal cells are marked with the arrowheads.

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Figure 6 is an immunostained culture of rat retinal neurons, which were grown for two days in the presence of rhGGF2 (neuregulin) on collagen gels showing extended long processes labeled with the TUJ1 antibody.

Figure 7 is an immunostained culture of rat retinal cells showing that neuregulin (rhGGF2) causes a significant increase in TUJ1 immunoreactive in embryonic day 18 rat retinal cells after two days of culture.

Figure 8 is an immunostained culture of rat retinal cells showing that the neuregulin (rhGGF2)- induced cell survival is age dependent: neuregulin (rhGGF2) does not cause a significant increase in TUJ1 immunoreactive embryonic day 15 rat retinal cells after two days of culture.

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Figure 9 is a bar graph of the results of three separate experiments with embryonic day 18 rat retinal cells.

Figure 10 is a bar graph of the experimental results showing the effects of GGF on retinal cell survival.

Figure 11A is a listing of the coding strand DNA sequence and deduced amino aid sequence of the cDNA obtained from the splicing pattern of GGFBPP1 shown in Figure 12. Potential glycosylation sites are underlined (along with polyadenylation signal AATAAA);

Figure 11B is a listing of the coding strand DNA sequence and deduced amino acid sequence of the cDNA obtained from splicing pattern of GGF2BPP2. Potential glycosylation sites are underlined (along with polyadenylation signal AATAAA);

Figure 11C is a listing of the coding strand DNA sequence and deduced amino acid sequence of the cDNA obtained from splicing pattern of GGF2BPP3. Potential glycosylation sites are underlined (along with polyadenylation signal AATAAA).

Figure 12 shows products of the neuregulin gene.

Figure 13 is a listing of the DNA sequences and predicted peptide sequences of the coding segments of GGF. Line 1 is a listing of the predicted amino acid sequences of bovine GGF, line 2 is a listing of the nucleotide sequences of bovine GGF, line 3 is a listing of the nucleotide sequences of human GGF (heregulin) (nucleotide base matches are indicated with a vertical line) and line 4 is a listing of the predicted amino acid sequences of human GGF/heregulin where it differs from the predicted bovine sequence. Coding segments E, A' and K represent only the bovine sequences. Coding segment D' represents only the human (heregulin) sequence.

Figure 14 is the predicted GGF2 amino acid sequence and nucleotide sequence of BPP5. The upper line is the nucleotide sequence and the lower line is the predicted amino acid sequence.

Figure 15 is the predicted amino acid sequence and nucleotide sequence of GGF2BPP2.

The upper line is the nucleotide sequence and the lower line is the predicted amino acid sequence.

Figure 16 is the predicted amino acid sequence and nucleotide sequence of GGF2BPP4. The upper line is the nucleotide sequence and the lower line is the predicted amino acid sequence.

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Figure 17 is a list of splicing variants derived from the sequences shown in Figure 13.

Figure 18 is the predicted amino acid sequence, bottom, and nucleic sequence, top, of EGFL1.

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Figure 19 is the predicted amino acid sequence, bottom, and nucleic sequence, top, of EGFL2.

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Figure 20 is the predicted amino acid sequence, bottom, and nucleic sequence, top, of EGFL3.

Figure 21 is the predicted amino acid sequence, bottom, and nucleic sequence, top, of EGFL4.

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Figure 22 is the predicted amino acid sequence, bottom, and nucleic sequence, top, of EGFL5.

Figure 23 is the predicted amino acid sequence, bottom, and nucleic sequence, top, of EGFL6.

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Figure 24 is the predicted amino acid sequence (middle) and nucleic sequence (top) of GGF2HBS5. The bottom (intermittent) sequence represents peptide sequences derived from GGF-II preparations.

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Figure 25 is the sequences of GGFHBS5, GGFHB1 and GGFBPP5 polypeptides.

Figure 26 is the amino acid sequence of cDNA encoding mature hGGF2.

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Figure 27 depicts a stretch of the putative bovine GGF-II gene sequence from the recombinant bovine genomic phage GGF2BG1. The figure is the coding strand of the DNA sequence and the deduced amino acid sequence in the third reading frame.

DETAILED DESCRIPTION OF THE INVENTION

It is intended that all references cited shall be incorporated herein by reference.

5 General

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The invention pertains to methods of promoting function of retinal cells. The function is affected by the administration of a neuregulin to a vertebrate where the neuregulin interacts with a retinal cell to promote one or more aspects of retinal cell function, including proliferation, differentiation, growth, survival, changes in the pattern of gene expression and secretion, and metabolic change of the retinal cell.

Definition of key terms

The term <u>administration</u> as used herein refers to the act of delivering a substance, including but not limited to the following routes: parenteral, intravenous, subcutaneous, intramuscular, intraorbital, ophthalmic, intravitreal, subretinal, intraperitoneal, topical, intranasal, aerosol or oral.

The term <u>affecting</u> as used herein refers to the induction of a quantitative change in the response of a target cell, as a result of an interaction with a neuregulin.

The term <u>amacrine cell</u> as used herein refers to local interneurons in the inner plexiform layer of the retina that mediate interactions between bipolar and ganglion cells.

The term <u>bipolar cell</u> as used herein refers to the interneurons of the retina that connect the photoreceptor cells with the retinal ganglion cells.

The term <u>differentiation</u> as used herein refers to a morphological and/or chemical change that results in the generation of a different cell type or state of specialization. The differentiation of cells as used herein refers to the induction of a cellular developmental program which specifies one or more components of a cell type. The therapeutic usefulness of differentiation can be seen, in increases in quantity of any component of a cell type in diseased tissue by at least 10% or more, more preferably by 50% or more, and most preferably by more than 100% relative to the equivalent tissue in a similarly treated control animal.

The term <u>disorder</u> as used herein refers to a disturbance of function and/or structure of a living organism, resulting from an external source, a genetic predisposition, a physical or chemical trauma, or a combination of the above, including but not limited to any mammalian disease.

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The term <u>erbB receptor</u> as used herein refers to erbB2. erbB3 and erbB4 (also HER-2. HER-3 and HER-4 of human) existing as monomeric. homodimeric and heterodimeric (e.g., erbB2/erbB3) cell surface receptor tyrosine kinases that bind and/or are activated by one or more neuregulins.

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The term <u>function</u> as used herein refers to any activity or response of a cell. These include but are not limited to proliferation, differentiation, growth, survival, changes in the pattern of gene expression and secretion, and metabolic changes.

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The term <u>horizontal cell</u> used herein refers to local interneurons in the outer plexiform layer of the retina that mediate interactions between bipolar and receptors cells.

The term <u>mammal</u> as used herein describes a member of the Class Mammalia (Subphylum Vertebrata).

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The term mitosis as used herein refers to the division of a cell where each daughter nucleus receives identical complements of the numbers of chromosomes characteristic of the somatic cells of the species. Mitosis as used herein refers to any cell division which results in the production of new cells in the patient. More specifically, a useful therapeutic is defined in vitro as an increase in mitotic index relative to untreated cells of 50%, more preferably 100%, and most preferably 300%, when the cells are exposed to labeling agent for a time equivalent to two doubling times. The mitotic index is the fraction of cells in the culture which have labeled nuclei when grown in the presence of a tracer which only incorporates during S phase (i.e., BrdU) and the doubling time is defined as the average time required for the number of cells in the culture to increase by a factor of two.

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The term <u>neuregulin</u> as used herein refers to the glial growth factors, the heregulins. neu differentiation factor, acetylcholine receptor inducing activity, and erbB2, 3 and 4 binding proteins. A more complete definition of neuregulins can be found in the specification herein and in the following materials: U.S. Patent No. 5,237,056; U.S. Patent Application SN 08/249,322; WO 92/20798; EPO 0 505 148 A1; Marchionni, et al., *Nature* 362:313, 1993; Benveniste, et al., *PNAS* 82:3930, 1985; Kimura, et al., *Nature* 348:257,

1990; Davis and Stroobant, J. Cell. Biol. 110:1353, 1990; Wen, et al., Cell 69:559, 1992; Yarden and Ullrich, Ann. Rev. Biochem. 57:443, 1988; Holmes, et al., Science 256:1205, 1992; Dobashi, et al., Proc. Natl. Acad. Sci. 88:8582, 1991; Lupu, et al., Proc. Natl. Acad. Sci. 89:2287, 1992; Peles and Yarden, BioEssays 15:815, 1993, Mudge, Current Biology 3:361, 1993, all hereby incorporated by reference.

The term <u>neurological disorder</u> as described herein refers to a disorder of the nervous system.

The term <u>photoreceptor cell</u> as used herein refers to two retinal cell types, rods and cones, that are the cells that transduce light into an electrical signal.

The term <u>retinal cell</u> as used herein refers to any of the cell types that comprise the retina, such as retinal ganglion cells, amacrine cells, horizontal cells, bipolar cells, and photoreceptor cells including rods and cones, Müller glial cells and retinal pigmented epithelium.

The term <u>retinal ganglion cell</u> as used herein refers to neurons of the retina that project axons via the optic nerve to the lateral geniculate nucleus and the superior colliculus.

The term <u>survival</u> as used herein refers to any process where a cell avoids death. The term survival as used herein also refers to the prevention of cell loss as evidenced by necrosis or apoptosis or the prevention of other mechanisms of cell loss. Survival as used herein indicates a decrease in the rate of cell death by at least 10%, more preferably by at least 50%, and most preferably by at least 100% relative to an untreated control. The rate of survival may be measured by counting cells stainable with a dye specific for dead cells (such as propidium iodide) in culture.

The term therapeutically effective amount as used herein refers to that amount which will produce a desirable result upon administration and which will vary depending upon a number of issues, including the dosage to be administered, and the route of administration.

The term <u>treating</u> as used herein may refer to a procedure (e.g. medical procedure) designed to exert a beneficial effect on a disorder. Treating as used herein means any administration of a substance described herein for the purpose of increasing retinal cell function. Most preferably, the treating is for the purpose of reducing or diminishing the symptoms or progression of a disease or disorder of retinal cells. Treating as used herein

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also means the administration of a substance to increase or alter the cells in healthy individuals. The treating may be brought about by the contacting of the cells which are sensitive or responsive to the neuregulins described herein with an effective amount of the neuregulin.

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The term <u>TUJ1</u> as used herein refers to an antibody that recognizes a neural-specific form of \(\beta\)-tubulin, which is expressed in the longitudinal cells, amacrine cells and ganglion cells of the retina.

The term <u>vertebrate</u> as used herein refers to an animal that is a member of the Subphylum Vertebrata (Phylum Chordata).

Neuregulins

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A novel aspect of the present invention relates to the ability of neuregulins to affect retinal cell function. Neuregulins are the products of a gene which produce a number of variably-sized, differentially-spliced RNA transcripts that give rise to a series of proteins. These proteins are of different lengths and contain some common peptide sequences and some unique peptide sequences. The conclusion that these factors are encoded by a single gene is supported by the differentially-spliced RNA sequences which are recoverable from bovine posterior pituitary, human spinal chord and human breast cancer cells (MDA-MB-231). Further support for this conclusion derives from the size range of proteins which act as ligands for the erbB receptors (see below).

Further evidence to support the fact a single gene encodes the various neuregulins derives from nucleotide sequence comparisons. Holmes et al., (Science 256:1205, 1992) demonstrate the purification of a 45-kilodalton human protein (Heregulin-a) which specifically interacts with the receptor protein p185erbB2. Peles et al., (Cell 69:559, 1992) describe a complementary DNA isolated from rat cells encoding a protein call "neu differentiation factor" (NDF). The translation product of the NDF cDNA has p185erbB2 binding activity. Several other groups have reported the purification of proteins of various molecular weights with erbB receptor binding activity. These groups include the following: Lupu et al., Proc. Natl. Acad. Sci. USA 89:2287, 1992; Yarden and Peles, Biochemistry 30:3543, 1991; Lupu et al., Science 249:1552, 1990; Dobashi et al., Biochem. Biophys. Res. Comm. 179:1536, 1991; and Huang et al., J. Biol. Chem. 257:11508, 1992.

We have found that proteins that bind p185erbB2 and related receptors (i.e., p185erbB3 and p185erbB4) affect retinal cell survival (Example 2). Further, the presence of immunologically-detectable neuregulin protein (Example 1) in retinal ganglion cells in vivo indicates that neuregulin has a role in retinal cell survival in vivo.

These neuregulins may be identified using the protocols described herein and in Holmes et al., Science 256; 1205, 1992; Peles et al., Cell 69:205, 1992; Wen et al., Cell 69:559, 1992; Lupu et al., Proc. Natl. Acad. Sci. USA 89:2287, 1992; Yarden and Peles, Biochemistry 30:3543, 1991; Lupu et al., Science 249:1552, 1990; Dobashi et al., Biochem. Biophys. Res. Comm. 179:1536, 1991; Huang et al., J. Biol. Chem. 257:11508-11512, 1992; Marchionni et al., Nature 362:313, 1993; and in U.S. Patent Application Serial No. 07/931.041, filed August 17, 1992, all of which are incorporated herein by reference.

Specifically, the invention provides for use of polypeptides of a specified formula, and DNA sequences encoding those polypeptides. The polypeptides have the formula WYBAZCX

wherein WYBAZCX is composed of the amino acid sequences shown in Figure 13; wherein W comprises the polypeptide segment F, or is absent: wherein Y comprises the polypeptide segment E. or is absent; wherein Z comprises the polypeptide segment G or is absent: and wherein X comprises the polypeptide segments C/D HKL. C/D H. C/D HL, C/D D, C/D HL, C/D D, C/D HKL, C/D HKL, C/D HKL, C/D C/D D HKL, C/D D HKL,

- a) at least one of F, Y, B, A, Z, C, or X is of bovine origin; or
- b) Y comprises the polypeptide segment E; or
- c) X comprises the polypeptide segments C/D HKL, C/D D, C/D' HKL, C/D C/D' HKL, C/D D' H, C/D D' HL, C/D D' HKL, C/D D' H, C/D D' HKL, C/D C/D' D' HKL, C/D C/D' D' HKL, C/D C/D' HKL, C/D C/D' HL.

In addition, the invention includes the use of the DNA sequence comprising coding segments 5'FBA3' as well as the with corresponding polypeptide segments having the amino acid sequences shown in Figure 13;

the DNA sequence comprising the coding segments ⁵FBA'³ as well as the corresponding polypeptide segments having the amino acid sequences shown in Figure 13;

the DNA sequence comprising the coding segments ⁵'FEBA³' as well as the corresponding polypeptide segments having the amino acid sequences shown in Figure 13;

the DNA sequence comprising the coding segments 5'FEBA'3' as well as the corresponding polypeptide segments having the amino acid sequences shown in Figure 13;

the DNA sequence comprising the polypeptide coding segments of the GGF2HBS5 cDNA clone (ATCC Deposit No. 75298, deposited September 2, 1992), also known as GGF-II.

The invention further includes the use of peptides of the formula FBA, FEBA, FBA' FEBA' and DNA sequences encoding these peptides wherein the polypeptide segments correspond to amino acid sequences shown in Figure 13. The polypeptide purified GGF-II polypeptide is also included as part of the invention.

Also included in this invention is the mature GGF peptide and the DNA encoding said peptide, exclusive of the N-terminal signal sequence, which is also useful for treatment of conditions involving abnormalities in retinal cell function.

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Furthermore, the invention includes a method of retinal cell function by the application to a vertebrate of a

- 30 kD polypeptide factor isolated from the MDA MB 231 human breast cell line;
- 5 or
 - 35 kD polypeptide factor isolated from the rat I-EJ transformed fibroblast cell line to the glial cell: or
 - -75 kD polypeptide factor isolated from the SKBR-3 human breast cell line; or
 - -44 kD polypeptide factor isolated from the rat I-EJ transformed fibroblast cell line,
- 10 or

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- -25 kD polypeptide factor isolated from activated mouse peritoneal macrophages; or
- -45 kD polypeptide factor isolated from the MDA MB 231 human breast cell; or
- -7 to 14 kD polypeptide factor isolated from the ATL-2 human T-cell line to the glial cell; or
- -25 kD polypeptide factor isolated from the bovine kidney cell; or
 - -42 kD polypeptide factor (ARIA) isolated from brains.

The invention further includes a method for the use of the EGFL1, EGFL2, EGFL3, EGFL4, EGFL5, and EGFL6 polypeptides, Figures 18 to 23 and respectively, for the methods of affecting retinal cell function in vivo and in vitro.

Also included in the invention is the administration of the GGF-II polypeptide whose sequence is shown in Figure 24 for affecting retinal cell function.

- Thus, the invention further embraces a polypeptide factor capable of affecting retinal cell function and including an amino acid sequence encoded by:
 - (a) a DNA sequence shown in Figure 11;
 - (b) a DNA sequence shown in Figure 27:
 - (c) the DNA sequence represented by nucleotides 281-557 of the sequences shown in Figure 11; or
 - (d) a DNA sequence hybridizable to any one of the DNA sequences according to (a), (b) or (c).

The invention further includes sequences which have greater than 60%, preferably 80%, sequence identity of homology to the sequences indicated above.

While the present invention is not limited to a particular set of hybridization conditions, the following protocol gives general guidance which may, if desired, be followed:

DNA probes may be labeled to high specific activity (approximately 10^8 to 10^9 32p dpm/µg) by nick-translation or by PCR reactions according to Schowalter and Sommer (Anal. Biochem. 177:90, 1989) and purified by desalting on G-150 Sephadex columns. Probes may be denatured (10 minutes in boiling water followed by immersion into ice water), then added to hybridization solutions of 80% buffer B (2g polyvinylpyrolidine, 2g Ficoll-400, 2g bovine serum albumin. 50ml 1 M Tris HCL (pH 7.5), 58g NaCl. 1g sodium pyrophosphate, 10g sodium dodecyl sulfate, 950 ml H₂O) containing 10% dextran sulfate at 10^6 dpm 32 P per ml and incubated overnight (approximately 16 hours) at 60° C. The filters may then be washed at 60° C first in buffer B for 15 minutes followed by three 20-minute washes in 2X SSC, 0.1% SDS then one for 20 minutes in 1XSSC, 0.1% SDS.

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In other respects, the invention provides:

(a) a basic polypeptide factor which has, if obtained from bovine pituitary material, an observed molecular weight, whether in reducing conditions or not, of from about 30 kD to about 36 kD on SDS-polyacrylamide gel electrophoresis using the following molecular weight standards:

Lysozyme (hen egg white)	14,400
Soybean trypsin inhibitor	21,500
Carbonic anhydrase (bovine)	31,000
Ovalbumin (hen egg white)	45,000
Bovine serum albumin	66,200
Phosphorylase B (rabbit muscle)	97,400;

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which factor has glial cell mitogenic activity including stimulating the division of rat Schwann cells in the presence of fetal calf plasma, and when isolated using reversed-phase HPLC retains at least 50% of said activity after 10 weeks incubation in 0.1 % trifluoroacetic acid at 4° C; and

(b) a basic polypeptide factor which has, if obtained from bovine pituitary material, an observed molecular weight, under non-reducing conditions, or from about 55 kD to about 63 kD on SDS-polyacrylamide gel electrophoresis using the following molecular weight standards:

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Lysozyme (hen egg white)	14,400
Soybean trypsin inhibitor	21,500
Carbonic anhydrase (bovine)	31,000
Ovalbumin (hen egg white)	45,000
Bovine serum albumin	66,200
Phosphorylase B (rabbit muscle)	97,400;

which factor the human equivalent of which is encoded by DNA clone GGF2HBS5 described herein and is capable of affecting retinal cell function.

For convenience of description only, the lower molecular weight and higher molecular weight factors of this invention are referred to hereafter as "GGF-I" and "GGF-II", respectively. The "GGF2" designation is used for all clones isolated with peptide sequence data derived from GGF-II protein (i.e., GGF2HBS5, GGF2BPP3).

It will be appreciated that the molecular weight range limits quoted are not exact, but are subject to slight variations depending upon the source of the particular polypeptide factor. A variation of, say, about 10% would not, for example, be impossible for material from another source.

Another important aspect of the invention is a DNA sequence encoding a polypeptide capable of affecting retinal cell function and comprising:

- (a) a DNA sequence shown Figure 11;
- (b) a DNA sequence shown in Figure 27;
- (c) the DNA sequence represented by nucleotides 281-557 of the sequence shown in Figure 11; or
- (d) a DNA sequence hybridizable to any one of the DNA sequences according to (a), (b) or (c).

Thus other important aspects of the invention are:

- (a) A series of human and bovine polypeptide factors capable of affecting retinal cell function. These peptide sequences are shown in Figures 13, 14, 15 and 16 respectively.
- (b) A series of polypeptide factors capable of affecting retinal cell function and purified and characterized according to the procedures outlined by Lupu et al., Science 249:1552, 1990; Lupu et al., Proc. Natl. Acad. Sci USA 89: 2287, 1992; Holmes et al., Science 256:1205, 1992; Peles et al., Cell 69:205, 1992; Yarden and Peles, Biochemistry 30:3543, 1991; Dobashi et al., Proc. Natl. Acad. Sci. 88: 8582, 1991; Davis et al., Biochem.

Biophys. Res. Commun. 179:1536, 1991; Beaumont et al., Patent Application PCT/US91/03443 (1990); Greene et al., Patent Application PCT/US91/02331 (1990); Usdin and Fischbach, J. Cell. Biol. 103:493, 1986; Falls et al., Cold Spring Harbor Symp. Quant. Biol. 55:397, 1990; Harris et al., Proc. Natl. Acad. Sci. USA 88:7664, 1991; and Falls et al., Cell 72:801, 1993.

(c) A polypeptide factor (GGFBPP5) is capable of affecting retinal cell function. The amino acid sequence is shown in Figure 14. and is encoded by the bovine DNA sequence shown in Figure 14.

The novel human peptide sequences described above and presented Figures 13, 14, 15, and 16, respectively, represent a series of splicing variants which can be isolated as full length complementary DNAs (cDNAs) from natural sources (cDNA libraries prepared from the appropriate tissues) or can be assembled as DNA constructs with individual exons (e.g., derived as separate exons) by someone skilled in the art.

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Other compounds, in particular peptides, which bind specifically to erbB receptors can also be used according to the invention as effectors of retinal cell function. A candidate compound can be routinely screened for erbB receptor binding, and, if it binds, can then be screened for affecting retinal cell function, more specifically, retinal cell survival, using the methods described herein.

The invention includes any modifications or equivalents of the above polypeptide factors which do not exhibit a significantly reduced activity. For example, modifications in which amino acid content or sequence is altered without substantially adversely affecting activity are included. By way of illustration, in EP-A 109748 mutations of native proteins are disclosed in which the possibility of unwanted disulfide bonding is avoided by replacing any cysteine in the native sequence which is not necessary for biological activity with a neutral amino acid. The statements of effect and use contained herein are therefore to be construed accordingly, with such uses and effects employing modified or equivalent factors being part of the invention.

The new sequences of the invention open up the benefits of recombinant technology. The invention thus also includes the following aspects:

(a) DNA constructs comprising DNA sequences as defined above in operable reading frame position within vectors (positioned relative to control sequences so as to permit expression of the sequences) in chosen host cells after transformation thereof by the constructs (preferably the control sequence includes regulatable promoters, e.g. Trp). It will

be appreciated that the selection of a promoter and regulatory sequences (if any) are matters of choice for those of skill in the art:

(b) host cells modified by incorporating constructs as defined in (a) immediately above so that said DNA sequences may be expressed in said host cells - the choice of host is not critical, and chosen cells may be prokaryotic or eukaryotic and may be genetically modified to incorporate said constructs by methods known in the art; and,

(c) a process for the preparation of factors as defined above comprising cultivating the modified host cells under conditions permitting expression of the DNA sequences. These conditions can be readily determined, for any particular embodiment, by those of skill in the art of recombinant DNA technology. Glial cell mitogens prepared by this means are included in the present invention.

None of the factors described in the art has the combination of characteristics possessed by the present new polypeptide factors.

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The invention also includes a neuregulin as defined above, by extracting vertebrate brain material to obtain protein, subjecting the resulting extract to chromatographic purification by hydroxyapatite HPLC and then subjecting these fractions to SDS-polyacrylamide gel electrophoresis. The fraction which as an observed molecular weight of about 30 kD to 36 kD and/or the fraction which has an observed molecular weight of about 55 kD to 63 kD is collected. In either case, the fraction is subjected to SDS-polyacrylamide gel electrophoresis using the following molecular weight standards:

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Lysozyme (hen egg white)	14,400
Soybean trypsin inhibitor	21,500
Carbonic anhydrase (bovine)	31,000
Ovalbumin (hen egg white)	45,000
Bovine serum albumin	66,200
Phosphorylase B (rabbit muscle)	97,400

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In the case of the smaller molecular weight fraction, the SDS-polyacrylamide gel is run in non-reducing conditions in reducing conditions or, and in the case of the larger molecular weight fraction the gel is run under non-reducing conditions. The fractions are then tested for activity stimulating the division of rat Schwann cells against a background of fetal calf plasma.

Preferably, the above process starts by isolating a relevant fraction obtained by carboxymethyl cellulose chromatography, e.g. from bovine pituitary material. It is also preferred that hydroxyapatite HPLC, cation exchange chromatography, gel filtration, and/or reversed-phase HPLC be employed prior to the SDS-Polyacrylamide gel electrophoresis. At each stage in the process, activity may be determined using Schwann cell incorporation of radioactive iododeoxyuridine as a measure in an assay generally as described by Brockes in Meth. Enz. 147:217, 1987, but modified by substituting 10% FCP for 10% FCS.

Compounds can be assayed for their usefulness in vitro using the methods provided in the description and examples below. Following the in vitro demonstration of the effect of neuregulins on retinal cell function, the in vivo therapeutic benefit of the effect can be accomplished by the administration of neuregulins, neuregulin producing cells or DNA encoding neuregulins to a vertebrate requiring therapy.

In Vitro Assays of Neuregulin Effects on Retinal Cells

Several in vitro assays are used to determine which neuregulin protein(s) promote retinal cell function and which retinal cell types are affected by contacting neuregulin protein. Described below are methods for detecting the ability of a neuregulin to promote function of a retinal cell. In vitro assays for determining neuregulin effects on retinal cell function depend on establishing retinal cultures. A general reference on cell and tissue culture is Cell and Tissue Culture: Laboratory Procedures (Ed. by A. Doyle, J. B. Griffiths. and D. G. Newell, John Wiley and Sons, New York, NY, 1994). General references on the culture of neural cells and tissues are Methods in Neurosciences, Vol. 2 (Ed. by P. M. Conn. Academic Press, Sand Diego, CA, 1990) and Culturing Nerve Cells (Ed. by G. Banker and K. Goslin, MIT Press, Cambridge, MA, 1991). General references of immunocytochemistry are Antibodies: A Laboratory Manual (E. Harlow and D. Lane, Cold Spring Harbor Laboratory. Cold Spring Harbor, NY, 1988), and Immunocytochemistry II (Ed. by A. C. Cuello, John Wiley and Sons, New York, NY, 1993).

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The retinal cells from a vertebrate used in this invention may be cultured in a variety of media. Commercially available media such as Ham's F10(Sigma), Minimal Essential Medium ([MEM], Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium ([DMEM], Sigma) are suitable for culturing retinal cells. In addition, any of the media described in Ham and Wallace. *Meth. Enz.* 58:44, 1979; Barnes and Sato. *Anal. Biochem.* 102:255, 1980; U.S. Pat. Nos. 4,767,704; 4,657,866; 4,927,762; or 4.560,655; WO 90/03430; WO 87/00195 and U.S. Pat. Re. 30,985, may be used as culture media for retinal

cells. Any of these media may be supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate), buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as GentamycinTM drug), trace elements (defined as inorganic compounds usually present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The culture conditions, such as temperature, pH. and the like, will be apparent to the ordinarily skilled artisan.

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The use of retinal cell cultures to demonstrate that neuregulin promotes retinal cell function is in accordance with methods described in general terms above and further described in Pittack et al., Devel. 113:577, 1991. The retina is dissected from either embryonic or adult vertebrate animals and placed into Ca⁺²/Mg⁺²-free Hepes-buffered sterile saline (HBSS) for 15 min., followed by treatment with 0.25% trypsin for an additional 15 min. The trypsin is inactivated by the addition of 1% fetal bovine serum. The cells are subsequently resuspended in fresh medium and gently triturated to yield a singlecell suspension. Cells are plated into wells of 24-well plates and cultured at 37°C. The types of retinal cells present in the culture can be identified through the use of immunocytochemical markers. Specific molecular markers can be stained immunocytochemically for the identification of cell types in the retina: for photoreceptors -e.g., rhodopsin, and red and green cone opsins: for amacrine cells -- e.g., cellular retinoic acid binding protein; for bipolar cells -- e.g., a specific form of protein kinase C and its substrate protein PCP2; for retinal ganglion cells--Thy1 and ß3-tubulin and; for horizontal cells--B3-tubulin. After maintaining the cultures for varying periods of time, preferably greater than 1 day and less than 7 days, a variety of assays can be utilized to assess various aspects of cellular phenotype such as, but not limited to, cell survival, proliferation, differentiation, morphology, and production of enzymes and secreted products.

In Vitro Method I

The survival function is assayed by methods that identify and count either viable cells or dead retinal cells following culture at low density (e.g., for retinal ganglion cells 10,000 cells/cm²) over a period from one to six days in the presence of varying amounts of neuregulin added to the culture medium. Included in these methods are specific stains for dead cells, such as propidium iodide, which enters the nucleus of dead cells and is detected

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by fluorescence microscopy. Alternatively, the counting of retinal cells adhering to the culture substratum over a six day period also can be used as an indicator of cell survival.

In Vitro Method II

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An alternative procedure to monitor retinal cell death utilizes labeling of nicked DNA strands, which are characteristic of cells undergoing apoptotic cell death, with digoxygenin-11-dUTP using terminal deoxynucleotidyl transferase (TUNEL) according to the protocol described in Gavrieli et al., J. Cell Biol. 119: 493-501, 1992. The labeled DNA strands are detected using standard kits available from commercial vendors (e.g., Genius kit from Boehringer Mannheim). Further, a cell death detection ELISA system, which is based on the DNA fragmentation that occurs in dying cells (Boehringer Mannheim catalog no. 1585 045) can be utilized to quantify cell death in accordance with the instructions provided by the commercial vendor.

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In Vitro Method III

The release into the culture medium of the cytosolic enzyme lactate dehydrogenase (LDH) also can be used to quantify the extent of retinal cell death in vitro (Kirk et al, J. Pharmacol. Exper. Therapeut. 271:1080, 1994). LDH levels are measured by an automated kinetic colorimetric assay in which oxidation of lactate to pyruvate is coupled to reduction of the tetrazolium dye, INT. Briefly, 80 ul samples of the culture medium are mixed with an equal volume of the substrate solution containing (in mg/l) INT, 334; phenazine methosulfate, 86; nicotinamide adenine dinucleotide, 862; L-(+)-lactate, 4900 (lithium salt); and 0.1% Triton X-100 in 0.2 M Tris buffer, pH 8.2. In the assay, LDH activity is directly proportional to the rate of appearance of the resulting INT formazan (absorbance max. at 492 nm). The product is monitored quantitatively in a microplate reader (UVmax, Molecular Devices, Menlo Park, CA) as the change in absorbance at 490 nm over a 2 min. interval.

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In Vitro Method IV

The proliferative function of neuregulins on retinal cells can be assayed by incorporation of 125I-Urd, 3H-dT or BrdU into replicating DNA strands of dividing cells, or by cell counting. The assays developed to measure the mitogenic activity of neuregulins on Schwann cells by incorporation of DNA synthesis precursors (Brockes et al., Brain Res.

165:105, 1979; Davis and Stroobant, J. Cell Biol. 110: 1353, 1990) can be adapted to retinal cells by one of normal skill in the art of cell culture.

In Vitro Method V

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The differentiation function of neuregulin on retinal cells can be assayed by employing analytical methods, such as immunostaining or in situ hybridization, which can detect and quantify marker proteins associated with the various cell types of the retina. Retinal ganglion cells are recognized by staining with the specific tubulin antibody TUJ1, as shown in Example 2 (Figure 4). The glial cells of the retina, Müller glia, are recognized by staining with antibodies that recognize glial fibrillary acidic protein (GFAP). For example, neurogenesis of retinal cells in culture can be achieved by dissociating embryonic retinal progenitor cells of the rat (from E15 through E18), then contacting the cells with the neuregulin and quantifying the distribution of various cell types identified by immunostaining using the markers described herein. In addition to this assay, which is based on determining activity in retinal neurogenesis, the differentiation function of neuregulin can be assayed in mature cultures (e.g., differentiated in culture for approximately two weeks). As such, changes in the level of specific proteins expressed in particular retinal cell types can be quantified.

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In Vitro Method VI

Several peptide growth factors and their receptors have been identified in the retina, as described in the prior act. Methods utilized to detect those molecules and activities can be employed to demonstrate a differentiation function of neuregulin on retinal cells. Neuregulins can be shown to induce the synthesis of growth factors and/or their receptors expressed in the retina. The analysis can be by in situ hybridization or other methods of quantitative RNA analysis, such as, but not limited to, reverse transcription-PCR, RNAse protection and Northern blotting. Alternatively, induced expression of growth factors or their receptors can be assayed by immunocytochemical staining or cell biological assays designed to measure growth factor activity.

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The *in vitro* assays described above to identify neuregulins that have biological activity on retinal cells can be applied to dissociated cells, semi-dissociated cells, explants of whole retina and parts thereof, such as preparations of retinal pigmented epithelium and other layers of the retina. The cultures can be established and maintained using methods described above. In some cases, minor modifications or substitutions to the procedures

described herein, which do not alter the reduction to practice of the invention, can be provided by one of ordinary skill in the art.

In Vivo Assays of Neuregulin Effects on Retinal Cells

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Neuregulin activity on retinal cells also can be shown through in vivo assays. Some in vivo assays represent animal models of retinal degeneration and other diseases and disorders of the eye. For example, photoreceptor cells are lost in inherited retinal degeneration and in age-related macular degeneration. Retinal ganglion cells die in glaucoma and in optic nerve injuries, such as retinal ischemia or axotomy.

In Vivo Method I

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The rescue of photoreceptor cells can be demonstrated in Royal College of Surgeons (RCS) rats, which have an inherited retinal degeneration (Faktorovich et al., Nature 347:83, 1990). The histological analysis (Method H1) consists of vascular perfusion of anesthetized animals, embedding the eye in epoxy resin, then staining I micron sections with toluidine blue. In untreated RCS rats at 53 days after birth (P53) the outer nuclear layer, which contains the photoreceptor cells, is reduced in thickness to only a few rows of cells (approximately 20% of the thickness found in normal rats at the same age). A therapeutically effective dose of neuregulin administered by intravitreal administration (a single injection of 1 microliter) can restore the thickness of the outer nuclear layer, and hence rescue photoreceptor cells. Alternatively, rescue of photoreceptor cells can be demonstrated in the Sprague-Dawley rat models (2-to-3 month old males) of exposure to constant light (115-200 foot-candles) for 1 week (LaVail et al., PNAS USA 89:11249, 1992). Neuregulin can be injected (1 ul) into the subretinal space or into the vitreous humor 48 hours prior to the onset of continuous illumination. Histological analysis (Method H1) of retinas following a fixed recovery period (usually 10 days) is used to assess the damage to and rescue of photoreceptor cells. Retinal detachment also leads to the death of photoreceptor cells, which provides another animal model (Erickson et al., J. Struct. Biol. 108:148, 1992) to demonstrate the in vivo survival activity of neuregulin on retinal cells.

In Vivo Method II

Several mouse genetic models of photoreceptor degeneration (e.g., rd--mutant of b subunit of cGMP phosphodiesterase; rds--mutant of peripherin) can be used to show neuregulin survival effects in vivo using the modes of administration described above. The rd and rds animals show retinal degeneration within a few weeks after birth and following intravitreal injection of neuregulin tissues can be analyzed by histological methods described above (e.g., Method H1). Further, retinal explants from rd mice cultured in neuregulin-containing medium can be assayed for thickness of the outer nuclear layer using methods described in Caffe et al., Curr. Eye Res. 12:719, 1993. Mouse pups are enucleated 48 hours after birth and treated with proteinase K. After enzyme treatment, the neural retina with the retinal pigmented epithelium (RPE) attached is recovered, placed into a multi-well culture dish and incubated in 1.2 ml culture medium (e.g., R16) for up to 4 weeks at 37 C with 5 % CO2. Immunocytochemical staining for opsin of fixed (e.g., 4% paraformaldehyde) sections is used to assess the degeneration and rescue of photoreceptor cells. In the rd mouse the outer nuclear layer (photoreceptor cells) degenerate after 2-to-4 weeks in culture. The

media can be supplemented with varying doses of neuregulin to achieve an effect on retinal cell function, such as rescue of the outer nuclear layer from degeneration. Survival effects also can be shown using the TUNEL method on sections of retina analyzed in the models described above.

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In Vivo Method III

In response to injury to the retina Müller cells undergo a proliferative gliosis. The mitotic activity of neuregulin on Müller glia can be shown by labeling dividing cells with a DNA synthesis precursor following administration of the factor. Labeled cells can be detected by autoradiography (3H-dT) or by immunostaining (BrdU labeling) and quantified.

In Vivo Method IV

Neuregulins can be shown to promote retinal ganglion cell survival following optic nerve axotomy or nerve crush using methods described in Sievers et al., Neurosci. Lett. 76:157, 1987; Carmignoto et al., J. Neurosci. 2:1263, 1989; Mey and Thanos, Brain Res. 602:304, 1993. Briefly, 4-to-6 week old mice are anesthetized, the optic nerve exposed and crushed intraorbitally 2-4 mm posterior to the optic disk between fine forceps for 30-60 sec. Alternatively, the nerve is transected surgically. Administration of neuregulin by intravitreal or subretinal injection is done after the animals recover from surgery using a therapeutically effective dose. The survival of retinal ganglion cells is assessed at several time points between 3 days and 6 weeks after injection by histological analysis (Method H1) or by immunostaining using antibodies that recognize retinal ganglion cells as described herein.

25 In Vivo Method V

Ischemia can be produced in the retina of the albino Lewis rat by raising intraocular pressure by intraocular injection of saline (Unoki and LaVail. *Invest Ophthalmol Vis. Sci.* 35:907, 1994). The thickness of the inner retinal layer is reduced due to loss of retinal ganglion cells when retinas are analyzed histologically (Method H1) at 7 days postischemia. An intravitreal injection of a therapeutically effective amount of neuregulin given two days prior to ischemia can reduce the ischemic damage.

In Vivo Method VI

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Other compounds, in particular peptides, which specifically bind and/or activate erbB receptors also can be used according to the invention as effectors of retinal cell

function. A candidate compound can be routinely screened for erbB receptor binding, and if it binds, can then be screened for affecting retinal cell function using the methods described herein.

5 In Vivo Method VII

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Inadequate amounts of survival-promoting factors can lead to degenerative eye disorders, such as macular degeneration. The present invention demonstrates the survivalpromoting activity of neuregulin indicating that these factors may be used to promote retinal cell survival in a vertebrate (preferably a mammal, more preferably a human) by administering to the vertebrate an effective amount of a polypeptide or a related compound. Neuregulin effects on retinal cells may occur, for example, by preventing the extent of naturally-occurring programmed cell death that occurs during the embryonic development of the retina. In a rat model, retinal ischemia can be induced by increasing intraocular pressure via injection of saline into the eye (Buchi et al., Ophthalmologic. 203:138, 1991; Hughes, Exp. Eye Res. 53:573, 1991). This model has been used to evaluate the efficacy of bFGF. CNTF and BDNF in decreasing neuronal loss (Unoki and LaVail, Invest. Ophthalmol. Vis. Sci. 35:907, 1994). Neuregulins administered by intraocular injection can be shown to decrease neuronal loss associated with retinal ischemia in this animal model. Neuregulin effects on retinal cell survival can be shown in genetic and transgenic mouse models for Retinitis Pigmentosa (rp). Histological analysis (Method H1) of the retina of rp mice following intravitreal administration of neuregulin can be used to rescue retinal cell degeneration.

The demonstration of biological activity of the neuregulins by promoting retinal cell function in any of the animal models described above indicates efficacy in treating disorders of the eye. A variety of retinal diseases and related disorders are known that produce impaired vision and in some cases progress to total blindness. These disorders of the eye include, but are not necessarily limited to: various retinopathies, such as hypertensive retinopathy, diabetic retinopathy and occlusive retinopathy; also injuries and disorders resulting in retinal degeneration, such as retinal tearing and detachment and inherited diseases, such as retinitis pigmentosa; also age-related macular degeneration (ARMD) and related diseases, such as idiopathic central serous chorioretinopathy, central areolar choroidal dystrophy, macular holes, macular coloboma, Stargardt hereditary dystrophy, trauma, diabetic circinate maculopathy, angioid streaks and choroidal neovascularization. presumed ocular histoplasmosis and choroidal neovascularization, angiomatosis retinae, choroidal rupture and choroidal neovascularization, toxoplasmosis and choroidal

neovascularization: diseases of the optic nerve; and glaucoma and retinal ischemia. Thus, administration of neuregulin in a therapeutically effective amount can provide a treatment for disorders of the eye, which otherwise left untreated would result in the loss of sight.

The invention includes the use of any modifications or equivalents of the above polypeptide factors which do not exhibit a significantly reduced activity related to affecting retinal cell function. For example, modifications in which amino acid content or sequence is altered without substantially adversely affecting activity are included. The statements of effect and use contained herein are therefore to be construed accordingly, with such uses and effects employing modified or equivalent factors being part of the invention.

The invention includes the use of the above named family of proteins (i.e. neuregulins) as extracted from natural sources (tissues or cell lines) or as prepared by recombinant means.

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The human peptide sequences described above represent a series of splicing variants which can be isolated as full length complementary DNAs (cDNAs) from natural sources (cDNA libraries prepared from the appropriate tissues) or can be assembled as DNA constructs with individual exons (e.g., derived as separate exons) by someone skilled in the art.

The invention includes methods for the use of any protein which is substantially homologous to the coding segments in Figure 13, as well as other naturally occurring neuregulin polypeptides for the purpose of promoting retinal cell function. Also included are the use of: allelic variations: natural mutants; induced mutants; proteins encoded by DNA that hybridizes under high or low stringency conditions to a nucleic acid naturally occurring (for definitions of high and low stringency see Current Protocols in Molecular Biology, (1989) John Wiley & Sons. New York, NY, 6.3.1 - 6.3.6, hereby incorporated by reference): and the use of polypeptides or proteins specifically bound by antisera to GGF polypeptides. The term also includes the use of chimeric polypeptides that include the GGF polypeptides comprising sequences from Figure 13.

Use of Neuregulins

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A novel aspect of the invention involves the use of neuregulins as factors to promote retinal cell function. Treatment of the cells to achieve these effects may be achieved by contacting cells with a polypeptide described herein.

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The methods of the invention make use of the fact that the neuregulin proteins are encoded by the same gene. A variety of messenger RNA splicing variants (and their resultant proteins) are derived from this gene and many of these products show binding to erbB receptors and activation of the same. This invention provides a use for all of the known products of the neuregulin gene (described herein and in the references listed above). Most preferably, recombinant human GGF2 (rhGGF2) is used in these methods.

The invention also relates to the use of other, not yet naturally isolated, splicing variants of the neuregulin gene. Figure 12 shows the known patterns of splicing. These patterns are derived from polymerase chain reaction experiments (on reverse transcribed RNA), analysis of cDNA clones (as presented within), and analysis of published sequences encoding neuregulins (Peles et al., Cell 69:205, 1992 and Wen et al., Cell 69:559, 1992). These patterns, as well as additional patterns disclosed herein, represent probable splicing variants which exist. The splicing variants are fully described in Goodearl et al., USSN 08/036,555, filed March 24, 1993, incorporated herein by reference.

More specifically, effects on retinal cell function may be achieved by contacting cells with a polypeptide defined by the formula

WYBAZCX

wherein WYBAZCX is composed of the polypeptide segments shown in Figure 13; wherein W comprises the polypeptide segment F, or is absent: wherein Y comprises the polypeptide segment E, or is absent; wherein Z comprises the polypeptide segment G, or is absent; and wherein X comprises the polypeptide segment C/D HKL, C/D H, C/D HL, C/D D, C/D' HL, C/D' HKL, C/D' D, C/D' HKL, C/D C/D' HKL, C/D C/D' D, C/D D' HKL, C/D C/D' D' D' HKL, C/D C/D' D' HKL, C/D C/D' D' D' D' D' D'

Furthermore, the invention includes a method of treating retinal cells by the application to the retinal cell of a

- -30 kD polypeptide factor isolated from the MDA-MB 231 human breast cell line; or
- -35 kD polypeptide factor isolated from the rat I-EJ transformed fibroblast cell line to the glial cell; or
 - -75 kD polypeptide factor isolated from SKBR-3 human breast cell line; or
- -44 kD polypeptide factor isolated from the rat I-EJ transformed fibroblast cell line:
 - -25 kD polypeptide factor isolated from activated mouse peritoneal macrophages; or

-45 kD polypeptide factor isolated from the MDA-MB 231 human breast cell; or

-7 to 14 kD polypeptide factor isolated from the ATL-2 human T-cell line to the glial cell; or

-25 kD polypeptide factor isolated from the bovine kidney cells; or

-42 kD ARIA polypeptide factor isolated from brain: or

-46-47 kD polypeptide factor which stimulates 0-2A glial progenitor cells: or

-43-45 kD polypeptide factor, GGFIII, U.S. patent application Serial No. 07/931,041. filed August 17, 1992, incorporated herein by reference.

The invention includes use of any modifications or equivalents of the above polypeptide factors which do not exhibit a significantly reduced activity. For example, modifications in which amino acid content or sequence is altered without substantially adversely affecting activity are included. The statements of effect and use contained herein are therefore to be construed accordingly, with such uses and effects employing modified or equivalent factors being part of the invention.

The human peptide sequences described above and presented in Figs. 13, 14, 15, and 16, respectively, represent a series of splicing variants which can be isolated as full-length complementary DNAs (cDNAs) from natural sources (cDNA libraries prepared from the appropriate tissues) or can be assembled as DNA constructs with individual exons (e.g., derived as separate exons) by someone skilled in the art.

Another aspect of the invention is the use of a pharmaceutical or veterinary formulation comprising any factor as defined above formulated for pharmaceutical or veterinary use, respectively, optionally together with an acceptable diluent, carrier or excipient and/or in unit dosage form. In using the factors of the invention, conventional pharmaceutical or veterinary practice may be employed to provide suitable formulations or compositions.

A medicament is made by administering the polypeptide with a pharmaceutically effective carrier. Neuregulins can be administered intravitreally by insertion of a needle through the sclera, choroid and retina and then injection of factor formulated in an appropriate vehicle for administration. The factor may also be delivered subretinally by a transpleural injection. There is also the option of delivering the factor intraocularly using ethylene-vinyl acetate copolymer implants or by delivery to the corneal surface via eye drops.

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Thus, the formulations to be used as a part of the invention can be applied to parenteral administration, for example, intravenous, subcutaneous, intramuscular, intraorbital, ophthalmic, intraperitoneal, topical, intranasal, aerosol, transdermal and by other slow release devices (i.e., osmotic pump-driven devices; see also USSN 08/293,465, hereby incorporated by reference).

The formulations of this invention may also be administered by the transplantation into the patient of host cells expressing the DNA encoding polypeptides which are effective for the methods of the invention or by the use of surgical implants which release the formulations of the invention.

Parenteral formulations may be in the form of liquid solutions or suspensions; for oral administration, formulations maybe in the form of tablets or capsules; and for intranasal formulations, in the form of powders, nasal drops, or aerosols.

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Methods well-known in the art for making formulations are to be found in, for example, "Remington's Pharmaceutical Sciences." Formulations for parenteral administration may, for example, contain as excipients sterile water or saline, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin, or hydrogenated naphthalenes, biocompatible, biodegradable lactide polymer, or polyoxyethylene-polyoxypropylene copolymers may be used to control the release of the present factors. Other potentially useful parenteral delivery systems for the factors include ethylene-vinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes. Formulations for inhalation may contain as excipients, for example, lactose, or may be aqueous solutions containing, for example, polyoxyethylene-9-lauryl ether, glycocholate and deoxycholate, or may be oily solutions for administration in the form of nasal drops, or as a gel to be applied intranasally. Formulations for parenteral administration may also include glycocholate for buccal administration, methoxysalicylate for rectal administration, or citric acid for vaginal administration.

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The present factors can be used as the sole active agents, or can be used in combination with other active ingredients, e.g., other growth factors which could facilitate neuronal survival in neurological diseases, or peptidase or protease inhibitors.

The concentration of the present factors in the formulations of the invention will vary depending upon a number of issues, including the dosage to be administered, and the route of administration.

In general terms, the factors of this invention may be provided in an aqueous physiological buffer solution containing about 0.1 to 10% w/v compound for parenteral administration. General dose ranges are from about 1 µg/kg to about 1g/kg of body weight per day; a preferred dose range is from about 0.01 mg/kg to 100 mg/kg of body weight per day. The preferred dosage to be administered is likely to depend upon the type and extent of progression of the pathophysiological condition being addressed, the overall health of the patient, the make up of the formulation, and the route of administration.

A further general aspect of the invention is the use of a factor of the invention in the manufacture of a medicament, preferably for the treatment of a disease or disorder. The "GGF2" designation is used for all clones which were previously isolated with peptide sequence data derived from GGF-II protein (i.e., GGF2HBS5, GGF2BPP3) and, when present alone (i.e., GGF2 OR rhGGF2), to indicate recombinant human protein encoded by plasmids isolated with peptide sequence data derived from the GGF-II protein (i.e., as produced in insect cells from the plasmid HBS5). Recombinant human GGF from the GGFHBS5 clone is called GGF2, rhGGF2 and GGF2HBS5 polypeptide.

Methods for treatment of diseases or disorders using nucleic acid constructs encoding neuregulins or neuregulin producer cells are also part of the invention.

Delivery of DNA to a cell or tissue that will take up the DNA, express the DNA and produce neuregulin as shown by Wolff et al., (Science 247:1465, 1990) and Ascadi et al., (Nature 352:815, 1991) is an aspect of the invention. Genetic modification of cultured cells (or their precursors) such as fibroblasts (as shown by Wolff et al. Proc. Nat'l Acad. Sci. USA 86:1575, 1988) or such as those derived from the nervous system (as shown by Weiss et al. International Patent Application number PCT/US94/01053; publication number WO 94/16718) to induce the production of neuregulin from the cultured cells is another aspect of this invention. The genetically modified neuregulin producer cells can be transplanted to a position near the retinal cell type and elicit the responses described above.

Other Embodiments

The invention includes methods for the use of any protein which is substantially homologous to the coding segments in Figure 13 as well as other naturally occurring GGF or neuregulin polypeptides for the purpose of promoting retinal cell function. Also included are the use of: allelic variations: natural mutants; induced mutants; proteins encoded by

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DNA that hybridizes under high or low stringency conditions to a nucleic acid naturally occurring (for definitions of high and low stringency see Current Protocols in Molecular Biology, John Wiley & Sons, New York, 1989; 6.3.1-6.3.6, hereby incorporated by reference); and the use of polypeptides or proteins specifically bound by antisera to GGF polypeptides. The term also includes the use of chimeric polypeptides that include the GGF polypeptides comprising sequences from Figure 11 for the promotion of retinal cell function.

As will be seen from Example 2, below, the present factors exhibit survival activity on retinal cells. The general statements of invention above in relation to formulations and/or medicaments and their manufacture should clearly be construed to include appropriate products and uses.

A series of experiments follow which provide additional basis for the claims described herein. The following examples relating to the present invention should not be construed as specifically limiting the invention, or such variations of the invention, now known or later developed.

The examples illustrate our discovery that recombinant human GGF2 (rhGGF2) confers survival effects on retinal cell culture. These activities indicate efficacy of GGF2 and other neuregulins in inducing wound repair and repair of other retinal tissue damage, and promoting regeneration and prophylactic effects on retinal tissue degeneration.

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EXAMPLES

The following examples are designed to illustrate certain aspects of the present invention. The examples are not intended to be comprehensive of all embodiments of the present invention, and should not be construed as limiting the claims presented herein.

Example 1: Neuregulin expression in embryonic and adult retina.

Neuregulin is expressed in the retina of the developing embryo and in adult rat. The pattern of expression has been demonstrated by in situ hybridization (Figure 3) and also by immunostaining (Figures 2 and 4). Expression is detected in the retinal ganglion cell layer. The expression occurs at a point in development when the retinal layers are undergoing differentiation and when the retinal ganglion cells are extending their axons and making connections to target of innervation in the brain (lateral geniculate and superior colliculus).

The timing and distribution of neuregulin gene products in the retina suggests the neuregulins have a role in the development and/or maintenance of the cells in the retina and their associated tissues.

Methods

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In situ hybridization (see Figure 3). Ten micron frozen section was incubated with a single-stranded digoxigenin-labeled riboprobe (antisense strand) encoding the EGF-like domain through the cytoplasmin domain of the rat cDNA clone GGFRP3 (Marchionni et al., Nature 362:312, 1993).

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Immunostaining. Ten micron frozen section of embryonic day 16 rat retina was incubated in CN16 (anti-rhGGF2) antibody at approximately 10 mg/ml for 12 hours, and the antibody binding was revealed using indirect immunohistochemistry with a peroxidase conjugated secondary antibody (see Figure 2).

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Ten micron frozen section from an adult rat retina was incubated in CN16 as described in Figure 1, except that a fluorescein conjugated secondary antibody was used to reveal the binding of the primary antibody (see Fig 4). Neuregulin immunoreactivity is present in the synaptic layers of the retina, where the processes of the retinal ganglion cells connect with the retinal interneurons (inner plexiform layer, large arrows) and in the outer plexiform layer (small arrows, where the processes of the photoreceptors make synapses with the second order retinal neurons, the bipolar cells and the horizontal cells).

Ten micron section from a newborn rat retina incubated with TUJ1 antibody, a mouse monoclonal antibody that recognizes neuron-specific beta -tubulin (from Dr. A. Frankfurter, UVA). The antibody binding was revealed by indirect immunohistochemistry with a fluorescein conjugated secondary antibody (see Figure 5).

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Example 2: Neuregulin (rhGGF2) promotes survival of retinal cells in vitro.

Embryonic and newborn rat retinal cells were cultured for 2 days on collagen gel coated coverslips and then fixed and labeled with an antibody (TUJ1) that identifies primarily retinal ganglion cells at these stages of development (see Figure 6). All the labeled cells with processes on the sample coverslips were counted. Final concentrations of rhGGF2 in the culture wells ranged from 0.01 to 100 ng/ml. No clear dose response was observed, so the data from all rhGGF2 treated wells was combined for the analysis. Three separate experiments with embryonic day 18 cells all showed an increase in the number of TUJ1 immunoreactive cells with processes after two days in vitro (see Figure 7). Unpaired sample student's T-test showed that the increases in cell survival were statistically significant with p<0.004 and p<0.012. Two experiments with embryonic day 15 cells and one experiment with newborn rat retinal cells did not show any significant differences from control (see Figure 8). From these observations we conclude that rhGGF2 promotes rat retinal cell survival in cell culture in an age-dependent manner. This age-dependence could represent either a changing requirement for this factor of a specific retinal cell population or a change in the relative number of the responsive cells in the population during these developmental stages. When assayed either alone or in combination with EGF, rhGGF2 had no mitogenic activity retinal cells in vitro at any of the ages tested.

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Methods

Embryonic rat retinal cell were dissociated and plated at low density on collagen gels and allowed to survive for two days with 10 ng/ml rhGGF2 (neuregulin) added to the medium. After fixation in 4% paraformaldehyde, the cells were stained with TUJ1 antibody to reveal the full extent of their processes and all the cells that survived for two days with intact, non-fragmented processes were counted.

CLAIMS

What is claimed is:

 1. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide defined by the formula

WYBAZCX

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- 2. A method of claim 1, wherein X is C/D HKL.
- 3. A method of claim 1, wherein X is C/D H.
- 20 4. A method of claim 1, wherein X is C/D HL.
 - 5. A method of claim 1, wherein X is C/D D.
 - 6. A method of claim 1, wherein X is C/D' HL.

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- 7. A method of claim 1, wherein X is C/D' HKL.
- 8. A method of claim 1, wherein X is C/D' H.

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- 9. A method of claim 1, wherein X is C/D' D.
- 10. A method of claim 1, wherein X is C/D C/D' HKL.
- 11. A method of claim 1, wherein X is C/D C/D' H.

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12. A method of claim 1, wherein X is C/D C/D' HL.

13. A method of claim 1, wherein X is C/D C/D' D.

- 14. A method of claim 1, wherein X is C/D D' H.
- 5 15. A method of claim 1, wherein X is C/D D' HL.
 - 16. A method of claim 1, wherein X is C/D D' HKL.
 - 17. A method of claim 1, wherein X is C/D' D' H.

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- 18. A method of claim 1, wherein X is C/D' D' HL.
- 19. A method of claim 1, wherein X is C/D' D' HKL.
- 15 20. A method of claim 1, wherein X is C/D C/D' D' H.
 - 21. A method of claim 1, wherein X is C/D C/D' D' HL.
 - 22. A method of claim 1, wherein X is C/D C/D' D' HKL.

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23. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising FBA polypeptide segments having the amino acid sequences shown in Figure 11.

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24. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising FBA' polypeptide segments having the amino acid sequences shown in Figure 11.

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25. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising FEBA polypeptide segments having the amino acid sequences shown in Figure 11.

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26. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising FEBA' polypeptide segments having the amino acid sequences shown in Figure 11 to said retinal cells.

27. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with GGF2 polypeptide.

- 28. The method of claim 27, wherein said GGF2 is recombinant human GGF2.
- 29. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a compound which binds with erbB receptors of said retinal cells.
- 30. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising EGFL1, having the amino acid sequence shown in Figure 18.
 - 31. A method of treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising EGFL2, having the amino acid sequence shown in Figure 19.
 - 32. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising EGFL3, having the amino acid sequence shown in Figure 20.
 - 33. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising EGFL4, having the amino acid sequence shown in Figure 21.
 - 34. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising EGFL5, having the amino acid sequence shown in Figure 22.
 - 35. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a polypeptide comprising EGFL6, having the amino acid sequence shown in Figure 23.
 - 36. A method for treating retinal cells of a mammal, said method comprising contacting a 45 kD polypeptide factor isolated from the rat I-EJ ras-transformed fibroblast cell line to said retinal cells.

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37. A method for treating retinal cells of a mammal, said method comprising contacting a 75 kD polypeptide factor isolated from SKBR-3 human breast cell line to said retinal cells.

- 38. A method for treating retinal cells of a mammal, said method comprising contacting a 45 kD polypeptide factor isolated from the MDA-MB231 human breast cell line to said retinal cells.
 - 39. A method for treating retinal cells of a mammal, said method comprising contacting a 7 to 14 kD polypeptide factor isolated from the ATL-2 human T-cell line to said retinal cells.
 - 40. A method for treating retinal cells of a mammal, said method comprising contacting a 25 kD polypeptide factor isolated from activated mouse peritoneal macrophages to said retinal cells.
 - 41. A method for treating retinal cells of a mammal, said method comprising contacting a 25 kD polypeptide factor isolated from bovine kidney to said retinal cells.
 - 42. A method for treating retinal cells of a mammal, said method comprising contacting an ARIA polypeptide to said retinal cells.
 - 43. A method for treating retinal cells of a mammal, said method comprising contacting a 46-47 kD polypeptide factor known to stimulate 0-2A glial progenitor cells to said retinal cells.
 - 44. A method for treating retinal cells of a mammal, said method comprising contacting GGF-III to said retinal cells.
 - 45. A method for treating retinal cells of a mammal, said method comprising administration to said mammal of a DNA sequence encoding a polypeptide of the formula

WYBAZCX

wherein WYBAZCX is composed of the polypeptide segments shown in Figure 13; wherein W comprises polypeptide segment F, or is absent; wherein Y comprises polypeptide segment E, or is absent; wherein Z comprises polypeptide segment G, or

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is absent; and wherein X comprises polypeptide segments C/D HKL, C/D H, C/D HL, C/D D, C/D' HL, C/D' HKL, C/D' H, C/D' D, C/D C/D' HKL, C/D C/D' H, C/D D' HL, C/D D' HKL, C/D D' H, C/D' D' HL, C/D' D' HKL, C/D' D' HKL, C/D' D' HKL, C/D' D' HKL, Said DNA in an expressible genetic construction.

- 46. A method for treating retinal cells of a mammal, said method comprising contacting said retinal cells with a therapeutically effective amount of a neuregulin polypeptide.
- 47. A method for the prophylaxis or treatment of pathophysiological condition of retinal cells in a mammal in which said condition involves a retinal cell type which is sensitive or responsive to a neuregulin polypeptide, said method comprising administration of a therapeutically effective amount of said neuregulin polypeptide.
- 48. A method for the treatment of a condition which involves retinal cell damage in a mammal, said method comprising contacting said retinal cell with an effective amount of a neuregulin polypeptide.
- 20 49. The method of any one of claims 1 through 28, wherein a result of said treating is decreased atrophy of said retinal cells.
 - 50. The method on any one of the claims 1 through 28, wherein a result of said treating is an increase of said retinal cells present in said mammal.
 - 51. The method on any one of the claims 1 through 28, wherein a result of said treating is an increase in said retinal cells survival in said mammal.
 - 52. A method of any one of the claims 1 through 28, wherein said retinal cells are in a mammal with a retinal cell disease.
 - 53. A method of claim 52, wherein said retinal cell disease is a retinopathy.
 - 54. A method of claim 53, wherein said retinopathy is hypertensive retinopathy.
 - 55. A method of claim 53, wherein said retinopathy is diabetic retinopathy.

- 56. A method of claim 53, wherein said retinopathy is occlusive retinopathy.
- 57. A method of claim 52, wherein said retinal cell disease is retinal degeneration.
- 5 58. A method of claim 57, wherein said retinal degeneration is caused by injury.
 - 59. A method of claim 57, wherein said retinal degeneration is caused by a genetic disorder.
- 10 60. A method of claim 59, wherein said genetic disorder is retinitis pigmentosa.
 - 61. A method of claim 57, wherein said retinal degeneration is age related macular degeneration.
- 62. A method of claim 52, wherein said retinal disease is caused by elevated intraocular pressure.
 - 63. A method of claim 52, wherein said retinal disease is caused by an optic neuropathy.
 - 64. A method for the prophylaxis or treatment of a pathophysiological condition of a retina in a vertebrate in which said condition involves a retinal cell type which is sensitive or responsive to a neuregulin polypeptide, said method comprising administration to said vertebrate of a therapeutically effective amount of said neuregulin polypeptide.
 - 65. A method of claim 53, wherein said condition involves retinal cell damage.
- 66. A method of any one of claims 1 through 28, wherein said retinal cell is a retinal ganglion cell.
 - 67. A method of any one of claims 1 through 28, wherein said retinal cell is an amacrine cell.
- 35 68. A method of any one of claims 1 through 28, wherein said retinal cell is a horizontal cell.

- 69. A method of any one of claims 1 through 28, wherein said retinal cell is a bipolar cell.
- 70. A method of any one of claims 1 through 28, wherein said retinal cell is a photoreceptor cell.
 - 71. A method of any one of claims 1 through 28, wherein said retinal cell is a pigment cell.
- 72. A method of treating retinal cells of a mammal, said method comprising contacting an N-ARIA polypeptide to said retinal cells.

Figure 1

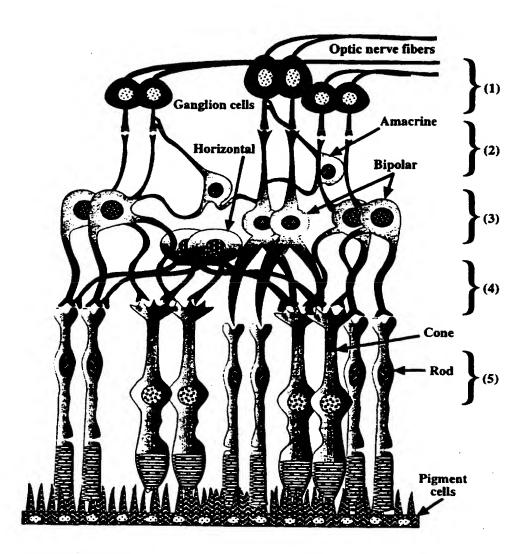


Figure 2

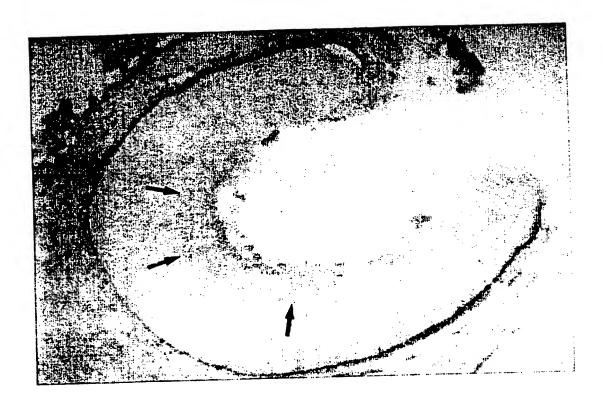


Figure 3



Figure 4

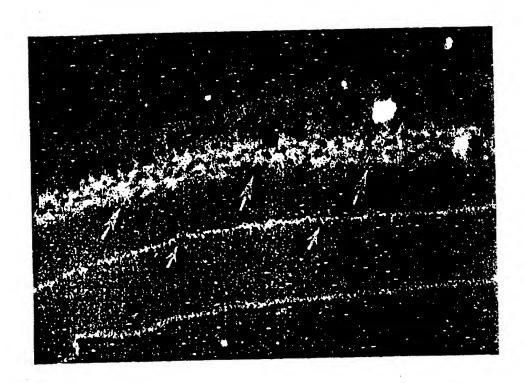


Figure 5

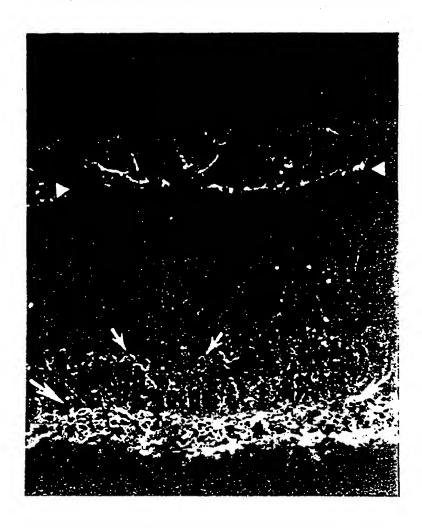


Figure 6



Figure 7

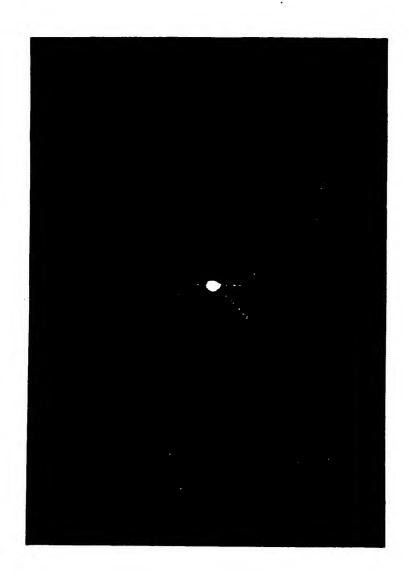


Figure 8

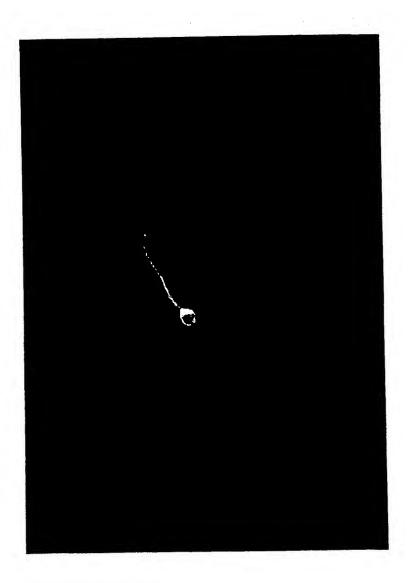


Figure 9

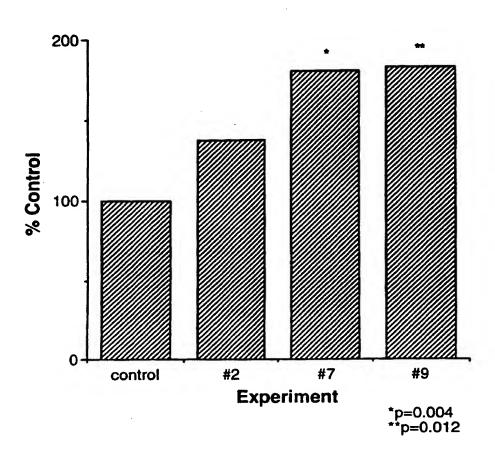


Figure 10

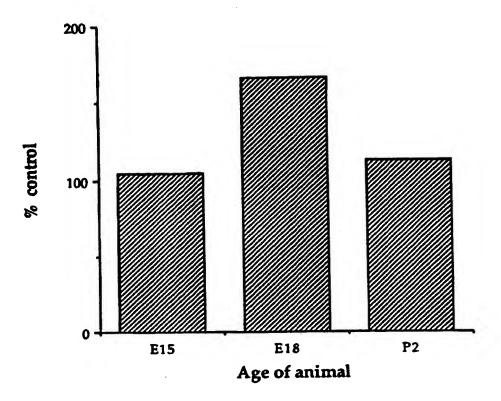


Figure 11 A

CCTG	CAG	CAT His 1	CAA Gln	GTG Val	TGG Trp	GCG Ala 5	GCG Ala	AAA Lys	GCC Ala	GGG Gly	GGC Gly 10	TTG Leu	AAG Lys	AAG Lys	GAC Asp	TCG Ser 15	CTG Leu	55
CTC Leu	ACC Thr	GTG Val	CGC Arg 20	CTG Leu	GGC Gly	GCC Ala	TGG Trp	GGC Gly 25	CAC His	CCC Pro	GCC Ala	TTC Phe	CCC Pro 30	TCC Ser	TGC Cys	t		103
GGG Gly	CGC Arg	CTC Leu 35	AAG Lys	GAG Glu	GAC Asp	AGC Ser	AGG Arg	TAC Tyr	ATC Ile	TTC Phe	TTC Phe	ATG Met 45	GAG Glu	CCC Pro	GAG Glu			151
GCC Ala	AAC Asn 50	NCC	AGC Ser	GGC Gly	GGG Gly	CCC Pro 55	GGC	CGC Arg	CTT Leu	CCG Pro	AGC Ser 60	CTC Leu	CTT Leu	CCC Pro	CCC Pro			199
TCT Ser 65	CGA Arg	GAC Asp	GGG Gly	CCG Pro	GAA Glu 70	CCT Pro	CAA Gln	GAA Glu	GGA Gly	GGT Gly 75	CAG Gln	CCG Pro	GGT Gly	GCT Ala	GTG Val 80			247
CAA Gln	CGG Arg	TGC Cys	GCC Ala	TTG Leu 85	CCT Pro	CCC Pro	CGC Arg	TTG Leu	AAA Lys 90	GAG Glu	ATG Met	AAG Lys	AGT Ser	CAG Gln 95	GAG Glu			295
TCT Ser	GTG Val	GCA Ala	GGT Gly 100	TCC Ser	AAA Lys	CTA Leu	Val	CTT Leu 105	CGG Arg	TGC Cys	GAG Glu	ACC Thr	AGT Ser 110	TCT Ser	GAA Glu			343
TAC Tyr	TCC Ser	TCT Ser 115	Leu	AAG Lys	TTC Phe	AAG Lys	TGG Trp 120	TTC Phe	AAG Lys	AAT Asn	GGG Gly	AGT Ser 125	GAA Glu	TTA Leu	AGC Ser			391
CGA Arg	AAG Lys 130	Asn	AAA Lys	CCA Pro	GAA Glu	AAC Asn 135	ATC Ile	AAG Lys	ATA Ile	CAG Gln	AAA Lys 140	AGG Arg	CCG Pro	GGG Gly	AAG Lys			439
TCA Ser 145	GAA Glu	CTT Leu	CGC Arg	ATT Ile	AGC Ser 150	AAA Lys	GCG Ala	TCA Ser	CTG Leu	GCT Ala 155	GAT Asp	TCT Ser	GGA Gly	GAA Glu	TAT Tyr 160	•		487
ATG Met	TGC Cys	AAA Lys	GTG Val	ATC Ile 165	AGC Ser	AAA Lys	CTA Leu	GGA Gly	AAT Asn 170	Asp	AGT Ser	GCC Ala	TCT Ser	GCC Ala 175	ASI			535
ATC Ile	ACC Thr	Ile	GTG Val 180	Glu	TCA Ser	AAC Asn	GGT Gly	AAG Lys 185	AGA Arg	TGC Cys	CTA Leu	CTG Leu	CGT Arg	Ala	TATI	:		583
TCT Ser	CAG Gln	TCT Ser 195	Leu	AGA Arg	GGA Gly	GTG Val	ATC Ile 200	Lys	GTA Val	TGT Cys	GGT	CAC His 205	Thr	•				625
TGA	ATCA	CGC	AGGT	GTGT	GA A	ATCT	CATT	G TG	AACA	<u>AATA</u>	AAA	LATCA	TGA	AAG	AAA	AAA		685
	-		ma	~ > ~~	ma a	3 CMC	C2 C2	m cm	CCCT	CCAC	: CTY	יכאריו	ፈጥን	GAGG	:ATC	C		744

Figure 11 B

CCTGCAG	CAT His 1	CAA Gln	GTG Val	TGG Trp	GCG Ala 5	GCG Ala	AAA Lys	GCC Ala	GGG	GGC Gly 10	TTG Leu	AAG Lys	AAG Lys	GAC Asp	TCG Ser 15	CTG Leu	55
CTC ACC Leu Thr	GTG Val	CGC Arg 20	CTG Leu	GGC Gly	GCC Ala	TGG Trp	GGC Gly 25	CAC His	CCC Pro	GCC Ala	TTC Phe	CCC Pro 30	TCC Ser	TGC Cys			103
GGG CGC Gly Arg	CTC Leu 35	AAG Lys	GAG Glu	GAC Asp	AGC Ser	AGG Arg 40	TAC Tyr	ATC Ile	TTC Phe	TTC Phe	ATG Met 45	GAG Glu	CCC Pro	GAG Glu			151
GCC AAC Ala <u>Lys</u> 50	AGC Ser	AGC Ser	GGC Gly	GGG Gly	CCC Pro 55	GGC Gly	CGC Arg	CTT Leu	CCG Pro	AGC Ser 60	CTC Leu	CTT Leu	CCC Pro	CCC			199
TCT CGA Ser Arg 65	GAC Asp	GGG Gly	CCG Pro	GAA Glu 70	CCT Pro	CAA Gln	GAA Glu	GGA Gly	GGT Gly 75	CAG Gln	CCG Pro	GGT Gly	GCT Ala	GTG Val 80			247
CAA CGG Gln Arg	TGC Cys	GCC Ala	TTG Leu 85	CCT Pro	CCC Pro	CGC Arg	TTG Leu	AAA Lys 90	GAG Glu	ATG Met	AAG Lys	AGT Ser	CAG Gln 95	GAG Glu			295
TCT GTG Ser Val	GCA Ala	GGT Gly 100	TCC Ser	AAA Lys	CTA Leu	Val	CTT Leu 105	CGG Arg	TGC Cys	GAG Glu	Thr	AGT Ser 110	TCT Ser	GAA Glu			343
TAC TCC Tyr Ser	Ser 115	Leu	AAG Lys	TTC Phe	AAG Lys	TGG Trp 120	TTC Phe	AAG Lys	AAT Asn	GGG Gly	AGT Ser 125	GAA Glu	TTA Leu	AGC Ser			391
CGA AAG Arg Lys 130	Asn	AAA Lys	CCA Gly	GAA Gly	AAC Asn 135	ATC Ile	AAG Lys	ATA Ile	CAG Gln	AAA Lys 140	Arg	CCG Pro	GGG Gly	AAG Lys			439
TCA GAA Ser Glu 145	CTT Leu	CGC Arg	ATT Ile	AGC Ser 150	Lys	GCG Ala	TCA Ser	CTG Leu	GCT Ala 155	Asp	TCT Ser	GGA Gly	GAA Glu	TAT Tyr 160	•		487
ATG TGC Met Cys	AAA Lys	GTG Val	ATC Ile 165	Ser	AAA Lys	CTA Leu	GGA Gly	AAT ASD 170	Asp	AGT Ser	GCC Ala	TCI Ser	GCC Ala 175	ASI	! \		535

Figure 11 B'

Ile	Thr	Ile	Val 180	Glu	Ser	Asn	Ala	Thr 185	Ser	Thr	Ser	Thr	Ala 190	Gly	Thr	58.	3
															AAT Asn	631	1
															TAC Tyr	679	9
															AAT Asn 240	7 27	7
															CTG Leu	775	5
_			GCC Ala 260				TAAT	rggco	CAG (CTTC	racao	GT AC	CGTC	CACTO	2	826	6
CCTI	TCTC	erc r	rctgo	CTG	AA TA	AGCGC	CATCI	CAC	TCG	STGC	CGCT	MTC1	rtg :	rtgco	CGCATC	886	5
TCCC	CTC	GA 7	rrcci	CCT	AG AG	CTAC	ATGO	GT	TTAC	CCAG	GTCT	raac <i>i</i>	ATT (GACTO	SCCTCT	946	5
GCCI	GTCC	CA 1	rgaga	ACAI	T A	CACA	AGC	ATT	GTAT	rgac	TTCC	CTCTC	STC (CGTG	CTAGT	1006	5
GGGC	TCTC	ag c	TACI	CGTA	G GI	GCG1	AAGO	CTC	CAGI	GTT	TCTC	CAAA	TG A	ATCTT	TEAATT	1066	5
ACTG	TGAT	AC C	ACAT	GATA	G TO	CCTC	TCAC	CC	GTGC	CAAT	GAC	ATA	AAG (CCT	AAAD7	1126	5
GTCA	AAAA	AA A	AAAA	AAAA	AA AA	AAAA	TCGA	TGI	CGAC	TCG	AGAT	CTG	CT (CAGO	STCGAC	1186	5
יייא	GAG															1107	2

Figure 11 C

																		55	
CCTG	CAG	CAT His 1	CAA Gln	GTG Val	TGG Trp	GCG Ala 5	GCG Ala	AAA Lys	GCC Ala	GGG Gly	GGC Gly 10	TTG Leu	AAG Lys	AAG Lys	GAC Asp	TCG Ser 15	Leu	55	,
CTC Leu	ACC Thr	GTG Val	CGC Arg 20	CTG Leu	GGC Gly	GCC Ala	TGG Trp	GGC Gly 25	CAC His	CCC Pro	GCC Ala	TTC Phe	CCC Pro 30	TCC Ser	TGC Cys			103	3
GGG Gly	CGC Arg	CTC Leu 35	AAG Lys	GAG Glu	GAC Asp	AGC Ser	AGG Arg 40	TAC Tyr	ATC Ile	TTC Phe	TTC Phe	ATG Met 45	GAG Glu	CCC Pro	GAG Glu			15:	1
GCC Ala	AAC Asn 50	AGC Ser	AGC Ser	GGC Gly	GGG Gly	CCC Pro 55	GGC Gly	CGC	CTT Leu	CCG Pro	AGC Ser 60	CTC Leu	CTT Leu	CCC	CCC			19	9
TCT Ser 65		GAC Asp	GGG Gly	CCG Pro	GAA Glu 70	CCT Pro	CAA Gln	GAA Glu	GGA Gly	GGT Gly 75	CAG Gln	CCG	GGT Gly	GCT Ala	GTG Val 80	;		24	7
	CGG Arg	TGC Cys	GCC Ala	TTG Leu 85	CCT	CCC Pro	CGC	TTG	AAA Lys 90	GAG Glu	ATG Met	AAG Lys	AGT Ser	CAG Gln 95	GAG Glu	; 1		29	5
TCT Ser	GTG Val	GCA Ala	GGT Gly	Ser	AAA Lys	CTA Leu	GTG Val	CTT Leu 105	CGG	TGC Cys	GAG Glu	ACC Thr	AGT Ser 110	TCI Ser	GAA Glu	7		34	13
TAC Tyr	TCC	TCT Ser	Lev	AAG Lys	TTC Phe	AAG Lys	TGG Trr	PILE	AAG Lys	CAA ;	GGC Gly	AG? / Sei 125		A TTA	A AGO	C r		39	91
CGA Arg	AAG Lys 130	: Ası	AAA 1 Lys	A CCA	GAZ Glu	A AAC Asr 135	J TT6	AAG Lys	ATA	A CAC	AAI Ly:	,	g CC	G GG(AA Ly	G S		4:	39
TCA Ser 145	Glu	CT:	r CGC ı Arg	AT Ile	T AGC Set 150	r Lys	A GCC s Ala	G TC/ a Sei	A CTO	G GC' 1 Ala 15	a no	T TC p Se	T GG r Gl	A GA	A TA u Ty 16	_		4	87

Figure 11 C'

			GTG Val														535
			GTG Val 180												ACA Thr		583
			GTC Val														631
			TGC Cys														679
			TGC Cys														727
			AGC Ser														775
GAA Glu	TAGO	GCA1	CT C	AGTC	GGTG	C CG	CTTI	CTTG	TTG	CCGC	CATC	TCCC	CTC	GA T	TCCGCCT	AG	838
AGCI	AGAT	GC G	TTTI	'ACCA	G GI	CTAA	CATI	GAC	TGCC	TCT	GCCI	GTC	CA 1	GAGA	ACATT		898
AACA	CAAG	CG A	TTGT	'ATGA	C TI	CCIC	TGTC	CGI	GACI	AGT	GGGC	TCTG	GAG C	TACI	CGTAG		958
GTGC	GTAA	GG C	TCCA	GTGT	T TC	TGAA	ATTG	ATC	TTGA	ATT	ACTG	TGAT	AC G	ACAT	GATAG	1	018
TCCC	TCTC	AC C	CAGT	GCAA	T GA	CAAT	<u>AAA</u> G	GCC	TTGA	AAA	GTCA	AAAA	AA A	AAAA	AAAAA	1	078
AAAA	ATCG	AT G	TCGA	CTCG	A GA	TGTG	GCTG									1	108

Figure 12

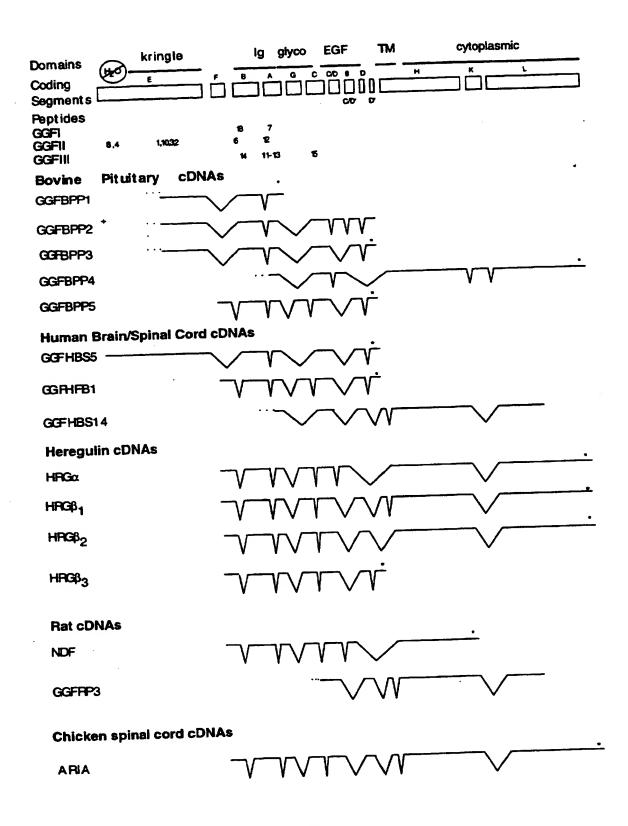


Figure 13 A

CODING SEGMENT F:	
AGTTTCCCCC CCCAACTTGT CGGAACTCTG GGCTCGCGCG CAGGGCAGGA GCGGAGCGGC	60
GGCGGCTGCC CAGGCGATGC GAGCGCGGGC CGGACGGTAA TCGCCTCTCC CTCCTCGGGC	120
TGCGAGCGCG CCGGACCGAG GCAGCGACAG GAGCGGACCG CGGCGGGAAC CGAGGACTCC	180
CCAGCGGCGC GCCAGCAGGA GCCACCCCGC GAGNCGTGCG ACCGGGACGG AGCGCCCGCC	240
AGTCCCAGGT GGCCCGGACC GCACGTTGCG TCCCCGCGGCT CCCCGCCGGC GACAGGAGAC	300
GCTCCCCCC ACGCCGCGC CGCCTCGGCC CGGTCGCTGG CCCGCCTCCA CTCCGGGGAC	360
AAACTTTTCC CGAAGCCGAT CCCAGCCCTC GGACCCAAAC TTGTCGCGCG TCGCCTTCGC	420
Met Ser Glu Arg Arg CGGGAGCCGT CCGCGCAGAG CGTGCACTTC TCGGGCGAG ATG TCG GAG CGC AGA	474
Glu Gly Lys Gly Lys Gly Lys Gly Gly Lys Lys Asp Arg Gly Ser Gly GAA GGC AAA GGC AAG GGG AAG GGC GGC AAG AAG	522
Lys Lys Pro Val Pro Ala Ala Gly Gly Pro Ser Pro Ala AAG AAG CCC GTG CCC GCG GCT GGC GGC CCG AGC CCA G	559

Figure 13 B

CODING SEGMENT E:	47
CC CAT CAN GTG TGG GCG GCG AAA GCC GGG GGC TTG AAG AAG GAC TCG His Gln Val Trp Ala Ala Lys Ala Gly Gly Leu Lys Lys Asp Ser 1 5	
CTG CTC ACC GTG CGC CTG GGC GCC TGG GGC CAC CCC GCC TTC CCC TCC Leu Leu Thr Val Arg Leu Gly Ala Trp Gly His Pro Ala Phe Pro Ser 20 25 30	95
TGC GGG CGC CTC AAG GAG GAC AGC AGG TAC ATC TTC ATG GAG CCC Cys Gly Arg Leu Lys Glu Asp Ser Arg Tyr Ile Phe Phe Met Glu Pro 35 40 45	143
GAG GCC AAC AGC AGC GGG CCC GGC CGC CTT CCG AGC CTC CTT CCC Glu Ala Asn Ser Ser Gly Gly Pro Gly Arg Leu Pro Ser Leu Leu Pro 50 55	191
CCC TCT CGA GAC GGG CCG GAA CCT CAA GAA GGA GGT CAG CCG GGT GCT Pro Ser Arg Asp Gly Pro Glu Pro Glu Gly Gly Gln Pro Gly Ala 65 70	239
GTG CAA CGG TGC G Val Gln Arg Cys 80	252
Figure 13 C	
CODING SEGMENT B:	
Leu Pro Pro Arg Leu Lys Glu His Lys Ser Gln Glu Ser Val Ala Gly CCT TGC CTC CCC GCT TGA AAG AGA TGA AGA GTC AGG AGT CTG TGG CAG	4
Con Luc Leu Val Leu Arg Cys Glu Thr Ser Ser Glu Tyr Ser Ser Leu	!

											_	_		33-	C3	
CCT	Pro TGC TGC	CTC	ĊĊĊ	GCT.	IGA	AAG	III	iii	1 1	1 1	111	1 1	1 1	1	111	48
GTT GTT	Lys CCA CCA	AAC AAC	TAG TAG	TCC	TTC	GGT	GTG	 AAA	CCA	GTT	CTG	 TAA	ACT	CCT	CTC	96
Lys TCA	Phe AGT GAT	Lys TCA	Trp AGT	Phe GGT	Lys TCA	Asn AGA	Gly ATG	Ser	Glu GTG	Leu AAT	Ser TAA	Arg GCC	Lys GAA	Asn AGA	Lys ACA	144
AA	Gly CAC	AAA	ACA	TCA	AGA 111	III			11		١					178

CODING SEGMENT A:

Figure 13 D

G AĀ	s Ser G TCA G TCA	GAA	CTT	CGC	TTA	AGC	AAA 	GCG	TCA	CTG	GCT	GAT	TCT	GGA	46
------	----------------------------------	-----	-----	-----	-----	-----	---------	-----	-----	-----	-----	-----	-----	-----	----

CAA	ጥልጥ	ATC	TGC	AAA	GTG	ATC	AGC	AAA	CTA	GGA	AAT	GAC	AGT	GCC	Ser TCT	94
11	111	111	111	111		111		111	- 11	-111	111		111	111	TCT	

GCC	Asn AAC	ATC	ACC	TTA	GTG	GAG	TCA	AAC	G
	AAT								

Figure 13 E

CODING SEGMENT A':

TCT	AAAA	CTA	CAGA	GACT	GT A	PTTT	CATG	A TC	ATCA?	TAGT	TCT	STGA	LAT A	ATACT	TAAAC	60
CGC'	ITTG	GTC	CTGA!	CTI	GT A									AA GO ys Al		110
TCA Ser 10	CTG Leu	GCT Ala	GAT Asp	TCT Ser	GGA Gly 15	GAA Glu	TAT Tyr	ATG Met	TGC Cys	AAA Lys 20	GTG Val	ATC Ile	AGC Ser	AAA Lys	CTA Leu 25	158
			AGT Ser													206
			CTA Leu 45													254
			GGT Gly			TGA	ATCA(CGC 1	AGGTY	GTGT(GA A	ATCT	TTAC	G		302
TGA	CAA	ATA .	AAAA	CAT	SA AE	AGGA	AAAC'	r cti	ATGT.	rtga	AAT	ATCT	TAT	GGGTY	CCTCCT	362
ርሞአ ፣	י א כיכי	יייטיי	מיר א רי	יייי	מ מיז	2CTYC!	ידעעע	A GAO	тстс	ጥፈፈሬ	מדב	ТАТА	באיר יי	אלא ר עים	r	417

Pigure 13 F

CODING S													_			
Glu Ile AG ATC AG ATC	ACC	ACT	GGC	ATG	LUA	111	111	ÎII	11	1	ĪĪ	111	111	111	47	
Ser Glu TCA GAG TCA GAG	TCT	ccc	ATT	AGA	ATA	ICA	III	II	111	ĬII	111	- 11	111		95	
Ser Ser TCT TCA TCT TCA	T														102	
							Fi	.gur	o 1	3 G	;					
CODING												•	0	212		
Thr CC ACA CT ACA	TCC		TCT	ACA	GCT	111	ACA	AGC	111	ĭii	11	11	III	11	47	,
Glu Ly: GAG AA GAG AA	GA	G AAJ	A ACT	r 1710 	: 11G:	r G10	AA، و ااا	1 662	1 11	ا ا	ĬĬĬ	ĬĬĬ	ĪÏĪ	1111		5
Lys As AAA GA AAA GA	CCT	T TCI	A AA' I II	r cc	C 1C	A AG.	A 1A 	1 11	6 16 1 11	Ī					12	8

Figure 13 H

CODING SEGMENT C/D:

AAG	TGC	CAA	CCT	GGĀ	TTC	ACT	GGA	GCG	AGA	Cys TGT TGT	ACT	GAG	AAT 	GTG	CCC	4.6	}
ATG	AAA	GTC	Gln CAA CAA	ACC	CAA	GAA										69)

Figure 13 I

CODING SEGMENT C/D':

_	TGC	CCA	AAT 	GAG	TTT	ACT	GGT	GAT	CGC	TGC	CAA	Asn AAC AAC	TAC	GTA	ATG	48
Ala GCC GCC	AGC	TTC	TAC													60

Figure 13 J

CODING SEGMENT D:

36

Figure 13 K

CODING SEGMENT D':

Lys His Leu Gly Ile Glu Phe Met Glu AAG CAT CTT GGG ATT GAA TTT ATG GAG

27

Figure 13 L

CODING	SEGMENT	н.

AĀA 	GCG	GAG	GAG	Leu CTC CTG	TAC	CAG	AAG	AGA	GTG 	CTC	ACC	ATT 	ACC	GGC 	ATT 	48
TGC	ATC	GCG	CTG	Leu CTC CTT	GTG	GTT	GGC	ATC	ATG	TGT	GTG	GTG	GTC	TAC	TGC	96
AĀA 	ACC	AĀG	AĀA 	Gln CAA CAG	CGG	аāа 	AAG	CTT	CAT	GAC	CGG	CTT	CGG	CAG	AGC	144
CIT	CGG	TCT	GAA 	Arg AGA CGA	AAC 	ACC	ATG	ATG	AAC	GTA 	GCC	AAC 	GGG	ccc	CAC	192
CAC	ccc	TAA	CCG	Pro CCC CCC	ccc	GAG	AAC 	GTG	CAG	CTG	GTG	TAA	CAA	TAC	GTA	240
TCT	AÃA 	TAA	GTC	Ile ATC ATC	TCT	AGC	GAG	CAT	TTA	GTT	GAG	AGA	GAG	GCG 	GAG	288

Figure 13 L'

AGC	Ser TCT TCC	$\mathbf{T}\mathbf{T}\mathbf{T}$	TCC	ACC	AGT	CAC	IAC	111	ii	111	11	111	11	111	111	336
ACT	Val GTC GTC	ACT	CAG	ACT	CCC	AGT	LAC	III	111	111	1.1	111	111	111	111	384
AGC	Ile ATC ATC	ATT	TCG	GAA	AGC	CAC	101	GIC	îii	III	111	111	111	-111	Glu GAA GAA	432
AAC AAC	AGT AGT	AGG AGG	CAC	AGC AGC	AGC AGC	CCA	ACT	 GGG	GGC	CCA	 AGA	 GGA	CGT	CTI	Asn TAA TAA	480
GG(Leu TTG	Gly GGA	Gly	Pro	Arg	Glu	Cys TGT	Asn	Ser	Phe	Leu CTC	Arg AGG	His CAT	Ala GCC	Arg AGA AGA	528
GA	Thr A ACC	; ÇÇI	GAC	TCC	TAC	; (G <i>p</i>	III	111	iii	Ĭij		ΙĪŪ	H	I		569

Figure 13 M

	ENT K:

AT A is A 1								46
TGC Cys					His			94
CCC								141

Figure 13 N

CODING SEGMENT L:

Tyr Val Ser Ala Met Thr Thr Pro Ala Arg Met Ser Pro Val Asp G TAT GTA TCA GCA ATG ACC ACC CCG GCT CGT ATG TCA CCT GTA GAT	46
Phe His Thr Pro Ser Ser Pro Lys Ser Pro Pro Ser Glu Met Ser Pro TTC CAC ACG CCA AGC TCC CCC AAG TCA CCC CCT TCG GAA ATG TCC CCG	94
Pro Val Ser Ser Thr Thr Val Ser Met Pro Ser Met Ala Val Ser Pro CCC GTG TCC AGC ACG GTC TCC ATG CCC TCC ATG GCG GTC AGT CCC	142
Phe Val Glu Glu Arg Pro Leu Leu Val Thr Pro Pro Arg Leu TTC GTG GAA GAG GAG AGA CCC CTG CTC CTT GTG ACG CCA CCA CGG CTG	190
Arg Glu Lys - Tyr Asp His His Ala Gln Gln Phe Asn Ser Phe His CGG GAG AAG TAT GAC CAC CAC GCC CAG CAA TTC AAC TCG TTC CAC	238
Cys Asn Pro Ala His Glu Ser Asn Ser Leu Pro Pro Ser Pro Leu Arg TGC AAC CCC GCG CAT GAG AGC AAC AGC CTG CCC CCC AGC CCC TTG AGG	286

PCT/US96/04240

Figure 13 N'

ATA 	GTG	GAG	GAT	GAG	GAA	TAT	GAA	ACG	ACC	CAG	GAG	TYT TAC TAC	GAA	CCA	GCT	334	
CAA	GAG	CCG	GTT	AAG	AAA	CIC	ACC	AAC	AGC	AGC	CGG	Arg CGG CGG	GCC 	AĀA 	AGA	382	
ACC	AAG	CCC	TAA 	GGT 	CAC	ATT 	GCC	CAC	AGG	TTG	GAA	Met ATG GTG V	GAC	AAC 	AAC 	430	
ACA	GGC	GCT 	GAC	AGC	AGT	AAC 	TCA	GAG	AGC	GAA	ACA	Glu GAG GAA	GAT	GAA	AGA	478	

Figure 13 N''

STA	GGA	GAA	GAT	ACG	Pro CCT CCT	TIC	CTG	1 1	îiî	III	III	III	III	111	111	526	;
AGT	CTC	GAG	GCG	GÇC	Pro CCT CCT	١١١	110	111	ĬĬĬ	ĭ	111	111	111	111		574	4
CCA	AÇA	GGC	GĞÇ	TIC	TCT	CCG	LAG	III	111	i	111	111	111	11	Ser TCC TCT	62	2
GGT AGT S	GTA GTA	ATC ATT	GCT GCT	AAC AAC	CAA	GAC GAC	CCI	ATT	GCT	 GTA	TAA	AAC	CT	ÄÄ	ACA AAA		
															r cca r cca	•	
111	111		144 1 144 1													7.	33

Figure 13 0

HUMAN CODING SEGMENT E:

	CGA Arg									48
	CCC Pro 20									96
	CTG Leu								•	144
	AAC Asn									192
	AGC Ser									240
	GAG Glu									288
	AAG Lys 100		Ala				Gly			336
	GAG Glu									384
	CCG Pro									432
	GTG Val									480
	GTG Val									528
	CTG Leu 180									576
	TGC Cys									624
	GAC Asp									672
	CCC Pro									720
	CTG Leu			G	20	/52				745

29/52

Figure 14 A

AGTTTCCCCC CCCAACTTGT CGGAACTCTG GGCTCGCGCG CAGGGCAGGA GCGGAGCGGC	60
GGCGGCTGCC CAGGCGATGC GAGCGCGGGC CGGACGGTAA TCGCCTCTCC CTCCTCGGGC	120
TGCGAGCGCG CCGGACCGAG GCAGCGACAG GAGCGGACCG CGGCGGGAAC CGAGGACTCC	180
CCAGCGGCGC GCCAGCAGGA GCCACCCCGC GAGCGTGCGA CCGGGACGGA GCGCCCGCCA	240
GTCCCAGGTG GCCCGGACCG CACGTTGCGT CCCCGCGCGCTC CCCGCCGGCG ACAGGAGACG	300
CTCCCCCCA CGCCGCGCG GCCTCGGCCC GGTCGCTGGC CCGCCTCCAC TCCGGGGACA	360
CTCCCCCCA CGCCGCGCGC GCCTCGGCCC GGTCGCTGCC GGTCGCCGCGT CGCCTTCGCC	420
AACTITICCC GAAGCCGATC CCAGCCCTCG GACCCAAACT TGTCGCGCGT CGCCTTCGCC	
GGGAGCCGTC CGCGCAGAGC GTGCACTTCT CGGGCGAG ATG TCG GAG CGC AGA Met Ser Glu Arg Arg 1 5	475
GAA GGC AAA GGC AAG GGG AAG GGC GGC AAG AAG	523
AAG AAG CCC GTG CCC GCG GCT GGC GGC CCG AGC CCA GCC TTG CCT CCC Lys Lys Pro Val Pro Ala Ala Gly Gly Pro Ser Pro Ala Leu Pro Pro 25	571
CGC TTG AAA GAG ATG AAG ATG CAG GAG TCT GTG GCA GGT TCC AAA CTA Arg Leu Lys Glu Met Lys Ser Gln Glu Ser Val Ala Gly Ser Lys Leu 40 45	619
GTG CTT CGG TGC GAG ACC AGT TCT GAA TAC TCC TCT CTC AAG TTC AAG Val Leu Arg Cys Glu Thr Ser Ser Glu Tyr Ser Ser Leu Lys Phe Lys 55 60 65	667
TGG TTC AAG AAT GGG AGT GAA TTA AGC CGA AAG AAC AAA CCA CAA AAC Trp Phe Lys Asn Gly Ser Glu Leu Ser Arg Lys Asn Lys Pro Gln Asn 70 75 80 85	715
ATC AAG ATA CAG AAA AGG CCG GGG AAG TCA GAA CTT CGC ATT AGC AAA Ile Lys Ile Gln Lys Arg Pro Gly Lys Ser Glu Leu Arg Ile Ser Lys 90 95	763
GCG TCA CTG GCT GAT TCT GGA GAA TAT ATG TGC AAA GTG ATC AGC AAA Ala Ser Leu Ala Asp Ser Gly Glu Tyr Met Cys Lys Val Ile Ser Lys 105	811

Figure 14 B

	GGA		Asp													859
			ACT Thr													907
			CCC Pro													955
			ACA Thr													1003
			AAG Lys 185													1051
			GAC Asp												CCA Pro	1099
			ACT Thr													1147
			TCC Ser									TAG	GCGC <i>I</i>	ATG		1193
CTC	AGTCO	GT (CCGC	TTTC	T TO	TTGC	CGC	A TCT	rccc	TCA	GATT	CAAC	CT A	AGAGO	TAGAT	1253
GCG	TTT	ACC A	GGTC	CTAAC	CA TT	GACT	GCC1	CIC	CCTC	TCG	CATO	SAGA	ACA 1	MAATT	CACAAC	1313
CGAT	TGT	ATG A	CTTC	CTCI	rg Tc	CGTC	SACTA	GTC	GGC1	CTG	AGC	CACT	GT A	AGGT	GCGTA!	1373
GGC1	CCAG	etg 1	TTCI	GAAA	T TO	ATCI	TGA	TT	CTGI	GAT	ACG!	CATO	SAT A	AGTC	CTCTC	1433
ACCO	AGTO	CA A	TGAC	AATA	A AG	GCCI	TGA	AAC	TCTC	CACT	TTT	ATTG/	AGA A	TAAL	AAAAA?	r 1493
CGTI	CCAC	CGG G	ACAG	TCCC	T CI	TCTI	TAT	AA.	ATGAC	CCT	ATC	TTG/	AAA A	AGGA	GTGT	1553
TTA	GTTC	STA A	CCAG	TACA	C AC	TTGA	LAAT(ATC	GTA	GTT	CGC	TCG	TT (CAGA	ATGTG:	r 1613
TCTI	TCTC	AC A	АТА	AACA	GAD	TAAZ	AAA			444	Δ					1654

Figure 15 A

CAT His 1	CAN Gln	GTG Val	TGG Trp	GCG Ala 5	GCG Ala	AAA Lys	GCC Ala	GGG Gly	GGC Gly 10	TTG Leu	AAG Lys	AAG Lys	GAC Asp	TCG Ser 15	CTG Leu		48
CTC Leu	ACC Thr	GTG Val	CGC Arg 20	CTG Leu	GGC Gly	GCC Ala	TGG Trp	GGC Gly 25	CAC His	CCC Pro	GCC Ala	TTC Phe	CCC Pro 30	TCC Ser	TGC Cys		96
Gly	Arg	Leu 35	Lys	Glu	Asp	AGC Ser	Arg 40	туг	116	FIIC	FIIC	45	020				144
GCC Ala	AAC Asn 50	AGC Ser	AGC Ser	GGC Gly	GGG Gly	CCC Pro 55	GGC Gly	CGC Arg	CTT Leu	CCG Pro	AGC Ser 60	CTC Leu	CTT	CCC Pro	CCC Pro		192
TCT Ser 65	CGA Arg	GAC Asp	GGG Gly	CCG Pro	GAA Glu 70	CCT	CAA Gln	GAA Glu	GGA Gly	GGT Gly 75	CAG Gln	CCG	GGT	GCT Ala	GTG Val 80		240
CAA Gln	CGG Arg	TGC Cys	GCC Ala	TTG Leu 85	CCT Pro	CCC Pro	CGC Arg	TTG Leu	AAA Lys 90	GAG Glu	ATG Met	AAC Lys	AGT Ser	CAG Glr 95	GAG Glu		288
TCI Ser	GTC Val	GCA Ala	A GG	y Sex	Lys	CTA Leu	GTG Val	CTI Leu 105	Arg	TGC Cys	GAC Glu	ACC Thi	AG? Sei 110		GAA Glu		336
TAC Tyr	TCC Sei	TC: Sei	Le	C AAC	TTO Phe	AAG Lys	TGG Tri	Phe	AAC Lys	AA' Asi	r GG(n Gl)	G AG' Y Set 12	. 61	TTI Le	A AGC 1 Ser		384
CG/ Arg	A AA(g Ly: 13(s As	c AA n Ly	A CC	A GAI	A AAC 1 Asi 135	3 TT€	C AAG e Ly:	S ATZ	A CA	a AA n Ly 14	ים כ	g CC	G GG o Gl	G AAG y Lys		432
TC: Se: 14	r Gl	A CT u Le	T CG u Ar	C AT	T AGG e Se: 15	r Ly:	A GCC	G TC. a Se	A CT r Le	G GC u Al 15	a no	T TC p Se	T GG r Gl	A GA y Gl	A TAT u Tyr 160		480
AT Me	G TG t Cy	C AA s Ly	A GI s Va	G AT	e Se	C AA	A CT. s Le	A GG u Gl	A AA y As 17	II AS	C AG	T GC	C TC	T GC er Al 17	C AAC a Asr	:	528

Figure 15 B

ATC Ile	ACC Thr	ATT Ile	GTG Val 180	GAG Glu	TCA Ser	AAC Asn	GCC Ala	ACA Thr 185	TCC Ser	ACA Thr	TCT Ser	ACA Thr	GCT Ala 190	GGG Gly	ACA Thr		576
AGC Ser	CAT His	CTT Leu 195	GTC Val	AAG Lys	TGT Cys	GCA Ala	GAG Glu 200	AAG Lys	GAG Glu	AAA Lys	ACT Thr	TTC Phe 205	TGT Cys	GTG Val	AAT Asn		624
GGA Gly	GGC Gly 210	GAG Glu	TGC Cys	TTC Phe	ATG Met	GTG Val 215	AAA Lys	GAC Asp	CTT Leu	TCA Ser	AAT Asn 220	CCC Pro	TCA Ser	AGA Arg	TAC Tyr		672
TTG Leu 225	Cys	AAG Lys	TGC Cys	CAA Gln	CCT Pro 230	GGA Gly	TTC Phe	ACT Thr	GGA Gly	GCG Ala 235	AGA Arg	TGT Cys	ACT Thr	GAG Glu	AAT Asn 240		720
GTG Val	CCC Pro	ATG Met	AAA Lys	GTC Val 245	CAA Gln	ACC Thr	CAA Gln	GAA Glu	AAG Lys 250	Cys	CCA Pro	AAT Asn	GAG Glu	TTT Phe 255	ACT Thr		768
GGT Gly	GAT Asp	CGC Arg	TGC Cys 260	Gln	AAC Asn	TAC Tyr	GTA Val	ATG Met 265	Ala	AGC Ser	TTC Phe	TAC Tyr	AGT Ser 270	THE	TCC		816
ACT Thr	CCC	TTT Phe 275	Leu	TCT Ser	CTG Leu	CCT Pro	GAA Glu 280		CGCA	TCT	CAGT	CGGI	GC C	GCTI	TCTIG		870
TTG	CCGC	ATC	TCCC	CTCA	GA T	TCCN	CCTA	G AG	CTAG	ATGO	GTI	TTAC	CAG	GTCT	AACAT	T	930
GAC	TGCC	TCT	GCCT	GTCG	CA T	GAGA	ACAT	T AA	CACA	AGCG	ATI	'GTA'	GAC	TTCC	TCTGT	c	990
CGT	GACT	AGT	GGGC	TCTG	AG C	TACT	CGTA	G GI	GCG1	'AAGC	CTC	CAGI	GTT	TCTG	SAAATT	.c	1050
ATC	TTGA	TTA	ACTG	TGAT	AC G	ACAT	GATA	G TC	CCTC	TCAC	CCA	GTGC	TAAC	GACA	AAATA J	.G	1110
			GTCA														1140

Figure 16 A

G AA Ly 1	rs Se	r Gl	u Le	u Ar	g Il	e Se	r Ly	rs Al	a Se	r Le	u Al	a As	sp Se	r Gl	y Glu	49
TAT Tyr	ATG Met	TGC Cys	AAA Lys 20	GTG Val	ATC Ile	AGC Ser	AAA Lys	CTA Leu 25	GGA Gly	AAT Asn	GAC Asp	AGT Ser	GCC Ala 30	TCT Ser	GCC Ala	97
AAC Asn	ATC Ile	ACC Thr 35	ATT Ile	GTG Val	GAG Glu	TCA Ser	AAC Asn 40	GCC Ala	ACA Thr	TCC Ser	ACA Thr	TCT Ser 45	ACA Thr	GCT Ala	GGG Gly	145
												ACT Thr				193
AAT Asn 65	GGA Gly	GGC Gly	GAC Asp	TGC Cys	TTC Phe 70	ATG Met	GTG Val	AAA Lys	GAC Asp	CTT Leu 75	TCA Ser	AAT Asn	CCC Pro	TCA Ser	AGA Arg 80	241
												AGA Arg			GAG Glu	289
AAT Asn	GTG Val	CCC Pro	ATG Met 100	AAA Lys	GTC Val	CAA Gln	Thr	CAA Gln 105	GAA Glu	AAA Lys	GCG Ala	GAG Glu	GAG Glu 110	CTC Leu	TAC Tyr	337
												GCG Ala 125				385
												AAG Lys			CGG Arg	433
															AAC Asn 160	481
															CCC Pro	529
		Val	Gln	Leu	Val	Asn	Gln	Tyr	Val	Ser	Lys		Val	Ile	TCT	577

Figure 16 B

										AGC Ser						625
CAC His	TAC Tyr 210	ACT Thr	TCG Ser	ACA Thr	GCT Ala	CAT His 215	CAT His	TCC Ser	ACT Thr	ACT Thr	GTC Val 220	ACT Thr	CAG Gln	ACT Thr	CCC Pro	673
										AGC Ser 235						721
CAC His	TCT Ser	GTC Val	ATC Ile	GTG Val 245	ATG Met	TCA Ser	TCC Ser	GTA Val	GAA Glu 250	AAC Asn	AGT Ser	AGG Arg	CAC His	AGC Ser 255	AGC Ser	769
										GGC Gly						817
										GAA Glu						865
										CTT Leu						913
										CAG Gln 315						961
										TGG Trp						1009
										GCA Ala					GCT Ala	1057
										AGC Ser					CCC Pro	1105
															CCC Pro	1153

Figure 16 C

TCC Ser 385	ATG Met	GCG Ala	GTC Val	AGT Ser	CCC Pro 390	TTC Phe	GTG Val	GAA Glu	GAG Glu	GAG Glu 395	AGA Arg	CCC Pro	CTG Leu	CTC Leu	CTT Leu 400	1201
GTG Val	ACG Thr	CCA Pro	CCA Pro	CGG Arg 405	CTG Leu	CGG Arg	GAG Glu	AAG Lys	TAT Tyr 410	GAC Asp	CAC His	CAC His	GCC Ala	CAG Gln 415	CAA Gln	1249
TTC Phe	AAC Asn	TCG Ser	TTC Phe 420	CAC His	TGC Cys	AAC Asn	CCC Pro	GCG Ala 425	CAT His	GAG Glu	AGC Ser	AAC Asn	AGC Ser 430	CTG Leu	CCC Pro	1297
CCC Pro	AGC Ser	CCC Pro 435	TTG Leu	AGG Arg	ATA Ile	GTG Val	GAG Glu 440	GAT Asp	GAG Glu	GAA Glu	ТАТ Тут	GAA Glu 445	ACG Thr	ACC Thr	CAG Gln	1345
GAG Glu	TAC Tyr 450	Glu	CCA Pro	GCT Ala	CAA Gln	GAG Glu 455	CCG Pro	GTT Val	AAG Lys	AAA Lys	CTC Leu 460	THE	AAC Asn	AGC Ser	AGC Ser	1393
CGG Arg 465	CGG Arg	GCC Ala	AAA Lys	AGA Arg	ACC Thr 470	Lys	CCC Pro	AAT Asn	GGT Gly	CAC His 475	ATT	GCC	CAC His	AGG Arg	TTG Leu 480	1441
GAA Glu	ATG Met	GAC Asp	AAC Asn	AAC Asn 485	Thr	GGC Gly	GCT Ala	GAC Asp	AGC Ser 490	Ser	AAC Asn	TCA Ser	GAG Glu	AGC Ser 495	GAA Glu	1489
ACA Thr	GAG Glu	GAT Asp	GAA Glu 500	Arg	GTA Val	GGA Gly	GAA Glu	GAT Asp 505	Tnr	CCT Pro	TTC Phe	CTG Lev	GCC Ala 510	TTE	A CAG	1537
AAC Asn	CCC	CTO Lev 515	Ala	GCC Ala	AGT Ser	CTC Leu	GAG Glu 520	ı Ala	GCC Als	CCI Pro	GCC Ala	TTO Phe 525	e Arg	CTC Lev	G GTC 1 Val	1585
GAC Asp	AGC Ser 530	Arg	ACI Thi	AAC Asr	C CCA	ACA Thr 535	G17	GGG Gly	TTC Phe	TC1	CCC Pro 540	O GTI	G GAI	A GAL	A TTG u Leu	1633
CAG Gln 545	Ala	AGO Arg	G CTO	TCC 1 Sei	GG1 G13 550	/ Val	ATY L Ile	C GC' e Ala	AAC a Asi	CAM 1 Gl: 55!	a Asj	p Pr	T ATO	C GC e Al	T GTC a Val 560	1681
TAP	LAAC	GAA	ATA	CACC	CAT A	AGAT	CAC	CT G	TAAA	ACTT	r at	TTTA	ATAT	ATA	AAGTATT	1741
CCA	CCT	AAA1	TTA	AACA	AAA A	AAA										176

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Figure 17 A

F-B-A'

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F-E-B-A'

F-B-A-C-C/D-D F-E-B-A-C-C/D-D F-B-A-C-C/D-H F-E-B-A-C-C/D-H F-B-A-C-C/D-H-L F-E-B-A-C-C/D-H-L F-B-A-C-C/D-H-K-L F-E-B-A-C-C/D-H-K-L F-B-A-C-C/D-D'-H F-E-B-A-C-C/D-D'-H F-B-A-C-C/D-D'-H-L F-E-B-A-C-C/D-D'-H-L F-B-A-C-C/D-D'-H-K-L F-E-B-A-C-C/D-D'-H-K-L F-B-A-C-C/D'-D F-E-B-A-C-C/D'-D F-B-A-C-C/D'-H F-E-B-A-C-C/D'-H F-B-A-C-C/D'-H-L F-E-B-A-C-C/D'-H-L F-B-A-C-C/D'-H-K-L F-E-B-A-C-C/D'-H-K-L F-B-A-C-C/D'-D'-H F-E-B-A-C-C/D'-D'-H F-B-A-C-C/D'-D'-H-L F-E-B-A-C-C/D'-D'-H-L F-B-A-C-C/D'-D'-'H-K-L F-E-B-A-C-C/D'-D'-'H-K-L F-B-A-C-C/D-C/D'-D F-E-B-A-C-C/D-C/D'-D F-B-A-C-C/D-C/D'-H F-E-B-A-C-C/D-C/D'-H F-B-A-C-C/D-C/D'-H-L F-E-B-A-C-C/D-C/D'-H-L F-B-A-C-C/D-C/D'-H-K-L F-E-B-A-C-C/D-C/D'-H-K-L F-B-A-C-C/D-C/D'-D'-H F-E-B-A-C-C/D-C/D'-D'-H F-B-A-C-C/D-C/D'-D'-H-L F-E-B-A-C-C/D-C/D'-D'-H-L F-B-A-C-C/D-C/D'-D'-H-K-L F-E-B-A-C-C/D-C/D'-D'-H-K-L

F-B-A-G-C-C/D-D F-B-A-G-C-C/D-H F-B-A-G-C-C/D-H-L F-B-A-G-C-C/D-H-K-L F-B-A-G-C-C/D-D'-H F-B-A-G-C-C/D-D'-H-L F-B-A-G-C-C/D-D'-H-K-L F-B-A-G-C-C/D'-D F-B-A-G-C-C/D'-H F-B-A-G-C-C/D'-H-L F-B-A-G-C-C/D'-H-K-L F-B-A-G-C-C/D'-D'-H F-B-A-G-C-C/D'-D'-H-L F-B-A-G-C-C/D'-D'-'H-K-L F-B-A-G-C-C/D-C/D'-D F-B-A-G-C-C/D-C/D'-H F-B-A-G-C-C/D-C/D'-H-L F-B-A-G-C-C/D-C/D'-H-K-L F-B-A-G-C-C/D-C/D'-D'-H F-B-A-G-C-C/D-C/D'-D'-H-L F-B-A-G-C-C/D-C/D'-D'-H-K-L F-E-B-A-G-C-C/D-C/D'-D'-H-K-L

F-E-B-A-G-C-C/D-D F-E-B-A-G-C-C/D-H F-E-B-A-G-C-C/D-H-L F-E-B-A-G-C-C/D-H-K-L F-E-B-A-G-C-C/D-D'-H F-E-B-A-G-C-C/D-D'-H-L F-E-B-A-G-C-C/D-D'-H-K-L F-E-B-A-G-C-C/D'-D F-E-B-A-G-C-C/D'-H F-E-B-A-G-C-C/D'-H-L F-E-B-A-G-C-C/D'-H-K-L F-E-B-A-G-C-C/D'-D'-H F-E-B-A-G-C-C/D'-D'-H-L F-E-B-A-G-C-C/D'-D'-'H-K-L F-E-B-A-G-C-C/D-C/D'-D F-E-B-A-G-C-C/D-C/D'-H F-E-B-A-G-C-C/D-C/D'-H-L F-E-B-A-G-C-C/D-C/D'-H-K-L F-E-B-A-G-C-C/D-C/D'-D'-H F-E-B-A-G-C-C/D-C/D'-D'-H-L

S

Figure 17 B

E-B-A'

E-B-A-C-C/D-D E-B-A-C-C/D-H E-B-A-C-C/D-H-L E-B-A-C-C/D-H-K-L E-B-A-C-C/D-D'-H E-B-A-C-C/D-D'-H-L E-B-A-C-C/D-D'-H-K-L E-B-A-C-C/D'-D E-B-A-C-C/D'-H E-B-A-C-C/D'-H-L E-B-A-C-C/D'-H-K-L E-B-A-C-C/D'-D'-H E-B-A-C-C/D'-D'-H-L E-B-A-C-C/D'-D'-H-K-L E-B-A-C-C/D-C/D'-D E-B-A-C-C/D-C/D'-H E-B-A-C-C/D-C/D'-H-L E-B-A-C-C/D-C/D'-H-K-L E-B-A-C-C/D-C/D'-D'-H E-B-A-C-C/D-C/D'-D'-H-L E-B-A-C-C/D-C/D'-D'-H-K-L

E-B-A-G-C-C/D-D E-B-A-G-C-C/D-H E-B-A-G-C-C/D-H-L E-B-A-G-C-C/D-H-K-L E-B-A-G-C-C/D-D'-H E-B-A-G-C-C/D-D'-H-L E-B-A-G-C-C/D-D'-H-K-L E-B-A-G-C-C/D'-D E-B-A-G-C-C/D'-H E-B-A-G-C-C/D'-H-L E-B-A-G-C-C/D'-H-K-L E-B-A-G-C-C/D'-D'-H E-B-A-G-C-C/D'-D'-H-L E-B-A-G-C-C/D'-D'-H-K-L E-B-A-G-C-C/D-C/D'-D E-B-A-G-C-C/D-C/D'-H E-B-A-G-C-C/D-C/D'-H-L E-B-A-G-C-C/D-C/D'-H-K-L E-B-A-G-C-C/D-C/D'-D'-H E-B-A-G-C-C/D-C/D'-D'-H-L E-B-A-G-C-C/D-C/D'-D'-H-K-L

								Val 15		48
								AGA Arg		96
								AAC Asn		144
		 	 	 	 	 	 	CTG Leu		192
GAA Glu 65	TAG									198

AGC Ser 1	CAT His	CTT Leu	GTC Val	AAG Lys 5	TGT Cys	GCA Ala	GAG Glu	AAG Lys	GAG Glu 10	AAA Lys	ACT Thr	TTC Phe	TGT Cys	GTG Val 15	AAT Asn	48
GGA Gly	GGC Gly	GAG Glu	TGC Cys 20	TTC Phe	ATG Met	GTG Val	AAA Lys	GAC Asp 25	CTT	TCA Ser	AAT Asn	CCC Pro	TCA Ser 30	AGA Arg	TAC Tyr	96
TTG Leu	TGC Cys	AAG Lys 35	TGC Cys	CAA Gln	CCT Pro	GGA Gly	TTC Phe 40	ACT Thr	GGA Gly	GCG Ala	AGA Arg	TGT Cys 45	ACT Thr	GAG Glu	AAT Asn	144
GTG Val	CCC Pro 50	ATG Met	AAA Lys	GTC Val	CAA Gln	ACC Thr 55	CAA Gln	GAA Glu	AAA Lys	GCG Ala	GAG Glu 60	GAG Glu	CTC	TAC Tyr	TAA	192

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 His	 	Lys 5				_			 48
 		TTC Phe							96
 	 	CCA Pro							144
 	 	TTC Phe	 	 	 	-	TAA		183

AGC Ser	CAT His	CTT Leu	GTC Val	AAG Lys 5	TGT Cys	GCA Ala	GAG Glu	AAG Lys	GAG Glu 10	AAA Lys	ACT Thr	TTC Phe	TGT Cys	GTG Val 15	AAT Asn	48
GGA Gly	GGC Gly	GAG Glu	TGC Cys 20	TTC Phe	ATG Met	GTG Val	AAA Lys	GAC Asp 25	CTT Leu	TCA Ser	AAT Asn	CCC	TCA Ser 30	AGA Arg	TAC Tyr	96
TTG Leu	TGC Cys	AAG Lys 35	TGC Cys	CCA Pro	AAT Asn	GAG Glu	TTT Phe 40	ACT Thr	GGT Gly	GAT Asp	CGC Arg	TGC Cys 45	CAA Gln	AAC Asn	TAC Tyr	144
GTA Val	ATG Met 50	GCC Ala	AGC Ser	TTC Phe	TAC Tyr	AAG Lys 55	CAT His	CTT	GGG Gly	ATT	GAA Glu 60	TTT Phe	ATG Met	GAG Glu	AAA Lys	192
GCG Ala	GAG Glu	GAG Glu	CTC Lev	TAC Tyr	TAA	•										210

AGC Ser 1	CAT His	CTT Leu	GTC Val	AAG Lys 5	TGT Cys	GCA Ala	GAG Glu	AAG Lys	GAG Glu 10	AAA Lys	ACT Thr	TTC Phe	TGT Cys	GTG Val 15	AAT Asn	48
												CCC Pro				96
TTG Leu	TGC Cys	AAG Lys 35	TGC Cys	CAA Gln	CCT Pro	GGA Gly	TTC Phe 40	ACT Thr	GGA Gly	GCG Ala	AGA Arg	TGT Cys 45	ACT Thr	GAG Glu	AAT Asn	144
GTG Val	CCC Pro 50	ATG Met	AAA Lys	GTC Val	CAA Gln	ACC Thr 55	CAA Gln	GAA Glu	AAG Lys	TGC Cys	CCA Pro 60	AAT Asn	GAG Glu	TTT Phe	ACT Thr	192
GGT Gly 65	GAT Asp	CGC Arg	TGC Cys	CAA Gln	AAC Asn 70	TAC Tyr	GTA Val	ATG Met	GCC Ala	AGC Ser 75	TTC Phe	TAC Tyr	AGT Ser	ACG Thr	TCC Ser 80	240
						CCT Pro	_	TAG								267

AGC Ser 1	CAT His	CTT Leu	GTC Val	AAG Lys 5	TGT Cys	GCA Ala	GAG Glu	AAG Lys	GAG Glu 10	AAA Lys	ACT	TTC Phe	TGT Cys	Val 15	Asn	•	•
GGA Gly	GGC Gly	GAG Glu	TGC Cys 20	TTC Phe	ATG Met	GTG Val	AAA Lys	GAC Asp 25	CTT Leu	TCA Ser	AAT Asn	CCC Pro	TCA Ser 30	AGA Arg	TAC Tyr	9	16
TTG Leu	TGC Cys	AAG Lys 35	TGC Cys	CAA Gln	CCT Pro	GGA Gly	TTC Phe 40	Thr	GGA Gly	GCG Ala	AGA Arg	TGT Cys 45	ACT	GAG Glu	AAT Asn	14	.4
GTG Val	CCC Pro 50	ATG Met	AAA Lys	GTC Val	CAA Gln	ACC Thr 55	CAA Gln	GAA Glu	AAG Lys	TGC Cys	CCA Pro 60	Wali	GAG Glu	TTT Phe	ACT Thr	19	12
GGT Gly 65	Asp	CGC Arg	TGC Cys	CAA Gln	AAC Asn 70	Tyr	GTA Val	ATG Met	GCC	AGC Ser 75	Pne	TAC	AAA Lys	GCG Ala	GAG Glu 80		40
	CTC Leu		TAA	,					•							2	52

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Figure 24 A

GGAATICCTT 111	TITITIT TITITIC	TI NINTITITI	TGCCCTTATA CCTCTTCGC	.C 60
TTTCTGTGGT TCC	ATCCACT TCTTCCCC	CT CCTCCTCCCA	TAAACAACTC TCCTACCCC	T 120
GCACCCCAA TAA	ATAAATA AAAGGAGG	AG GGCAAGGGGG	GAGGAGGAGG AGTGGTGCT	'G 180
CGAGGGGAAG GAA	AAGGGAG GCAGCGCG	AAGAGCCGGG	CAGAGTCCGA ACCGACAGC	C 240
AGAAGCCCGC ACG			GC GCC CCG CGC CGC rg Ala Pro Arg Arg 5	291
			GGC TCC GCC GCC CGC Gly Ser Ala Ala Arg 25	339
		eu Pro Leu Leu	CTG CTG CTG GGG ACC Leu Leu Leu Gly Thr Cys Leu Leu Thr Val GGF II 09	387
Ala Ala Leu Al Ala Ala Leu Pr	G CCG GGG GCG GC a Pro Gly Ala Al	G GCC GGC AAC	GAG GCG GCT CCC GCG Glu Ala Ala Pro Ala 55	435
GGG GCC TCG GT Gly Ala Ser Va 60	l Cys Tyr Ser Se Ala Se	r Pro Pro Ser	GTG GGA TCG GTG CAG Val Gly Ser Val Gln Val Gly Ser Val Gln GGF II 08	. 483
80	•	5	70	
Glu Leu Ala Gl Glu Leu Val Gl	n Arg Ala Ala n Arg Trp Phe	Val Val Ile Gl	G GGA AAG GTG CAC CC u Gly Lys Val His Pr u Gly Lys	G 531
75	90		05	

Figure 24 B

CAG Gln 90	CGG Arg	CGG Arg	CAG Gln	CAG Gln	GGG Gly 95	GCA Ala	CTC Leu	GAC Asp	AGG Arg	AAG Lys 100	GCG Ala	GCG Ala	GCG Ala	GCG Ala	GCG Ala 105	579
GGC Gly	GAG Glu	GCA Ala	GGG Gly	GCG Ala 110	TGG Trp	Gly	GGC Gly	GAT Asp	CGC Arg 115	GAG Glu	CCG Pro	CCA Pro	GCC Ala	GCG Ala 120	GGC Gly	627
CCA Pro	CGG	GCG Ala	CTG Leu 125	GGG Gly	CCG Pro	CCC Pro	GCC Ala	GAG Glu 130	GAG Glu	CCG Pro	CTG Leu	CTC Leu	GCC Ala 135	GCC Ala	AAC Asn	675
GGG Gly	ACC Thr	GTG Val 140	CCC Pro	TCT Ser	TGG Trp	CCC Pro	ACC Thr 145	GCC Ala	CCG Pro	GTG Val	CCC Pro	AGC Ser 150	GCC Ala	GGC Gly	GAG Glu	723
CCC Pro	GGG Gly	GAG Glu	GAG Glu	GCG Ala	CCC Pro	Tyr	Leu	GTG Val	LVS	Val Val	nis His F II	Glu 01	Val	Trp	GCG Ala Ala 11	771
	155					160					165	'				
Val	AAA Lys Lys	Ala	GGG Gly	GGC Gly	TTG Leu	AAG Lys	AAG Lys	ASD	TCG Ser Leu	Leu GGF	Leu	Xaa	Val	. Ary	Leu Leu	819
170	ı				175	1				180	ı				185	
Gla	The w	Trans	Gly Gly	CAC His Pro	Pro	Ala	Phe	Pro) Ser	: Cys	: GIJ	ATS	G CIO	ı răs	GAG Glu	867
				190)				195	i				200)	
GAC Asi	AGC Ser	AGC Arg	T327	- Tle	• Phe	• Phe	e Met	: Gl:	ı Pro) Ast	Ala Ala	a Asi	n se	r m	AGC Ser Gly	
			205	5				210					21	5		

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Figure 24 C

CGC Arg	GCG Ala	CCG Pro 220	GCC Ala	GCC Ala	TTC Phe	CGA Arg	GCC Ala 225	TCT Ser	TTC Phe	CCC Pro	CCT Pro	CTG Leu 230	GAG Glu	ACG Thr	GGC Gly	963
CGG Arg	AAC Asn 235	CTC Leu	AAG Lys	AAG Lys	GAG Glu	GTC Val 240	AGC Ser	CGG Arg	GTG Val	CTG Leu	TGC Cys 245	AAG Lys	CGG Arg	TGC Cys	GCC Ala	1011
TTG Leu 250	CCT Pro	CCC Pro	CAA Gln	TTG Leu	AAA Lys 255	GAG Glu	ATG Met	AAA Lys	AGC Ser	CAG Gln 260	GAA Glu	TCG Ser	GCT Ala	GCA Ala	GGT Gly 265	1059
TCC Ser	AAA Lys	Leu	Val Val	CTT Leu Leu II	Arg Arg	TGT Cys	GAA Glu	ACC Thr	AGT Ser	TCT Ser	GAA Glu	TAC Tyr	TCC Ser	TCT Ser	CTC Leu	1107
			GGF	270					175					180		
AGA Arg	TTC Phe	AAG Lys	TGG Trp 185	TTC Phe	AAG Lys	AAT Asn	GGG Gly	AAT Asn 190	GAA Glu	TTG Leu	AAT Asn	CGA Arg	AAA Lys 195	AAC Asn	AAA Lys	1155
CCA Pro	CAA Gln	AAT Asn 200	ATC Ile	AAG Lys	ATA Ile	CAA Gln	AAA Lys 205	AAG Lys	CCA Pro	GGG Gly	AAG Lys	TCA Ser 210	GAA Glu	CTT Leu	CGC Arg	1203
ATT Ile	AAC Asn	Lys	Ala	Ser	Leu	Ala	Asp Asp	TCT Ser Ser GF I	Gly Gly	Glu	Tyr	Met	Cys	Lys	GTG Val	1251
	215					220					225					
ATC Ile 230	AGC Ser	AAA Lys	TTA Leu	GGA Gly	AAT Asn 235	GAC Asp	AGT Ser	GCC Ala	TCT Ser	GCC Ala 240	AAT Asn	ATC	ACC Thr	ATC Ile	GTG Val 245	1299
								ACC Thr							GTA Val	1347

Figure 24 D

AAA TGT GCG GAG AAG GAG AAA ACT TTC TGT GTG AAT GGA GGG GAG TGC Lys Cys Ala Glu Lys Glu Lys Thr Phe Cys Val Asn Gly Gly Glu Cys 275	1395
TTC ATG GTG AAA GAC CTT TCA AAC CCC TCG AGA TAC TTG TGC AAG TGC Phe Met Val Lys Asp Leu Ser Asn Pro Ser Arg Tyr Leu Cys Lys Cys 280 285	1443
CCA AAT GAG TTT ACT GGT GAT CGC TGC CAA AAC TAC GTA ATG GCC AGC Pro Asn Glu Phe Thr Gly Asp Arg Cys Gln Asn Tyr Val Met Ala Ser 295 300	1491
TTC TAC AGT ACG TCC ACT CCC TTT CTG TCT CTG CCT GAA Phe Tyr Ser Thr Ser Thr Pro Phe Leu Ser Leu Pro Glu 400 405	1530
TAGGAGCATG CTCAGTTGGT GCTGCTTTCT TGTTGCTGCA TCTCCCCTCA GATTCCACCT	1590
AGAGCTAGAT GTGTCTTACC AGATCTAATA TTGACTGCCT CTGCCTGTCG CATGAGAACA	1650
TTAACAAAAG CAATTGTATT ACTTCCTCTG TTCGCGACTA GTTGGCTCTG AGATACTAAT	
AGGTGTGTGA GGCTCCGGAT GTTTCTGGAA TTGATATTGA ATGATGTGAT ACAAATTGAT	1770
AGTCAATATC AAGCAGTGAA ATATGATAAT AAAGGCATTT CAAAGTCTCA CTTTTATTGA	
TAAAATAAAA ATCATTCTAC TGAACAGTCC ATCTTCTTTA TACAATGACC ACATCCTGAA	1890
AAGGGTGTTG CTAAGCTGTA ACCGATATGC ACTTGAAATG ATGGTAAGTT AATTTTGATT	1950
CACAA MOODOO MANTACACA AAATAAACAT AATAAAAGGA AAAAAAAAAAA	2003

-	MRMRRAPRRSGRPGPRAORPGSAARSSPPLPLLPLLFLFGTAALAPGAAAGNEAAPAGAS
61	II-8 II-4 VCYSSPPSVGSVOELAORAAVVIEGKVHPORROXGALDRKAAAAAGFAGAWGGDREPPAA
	0 11-11
121	VKAGGLKR
•	11-3
181	LTVRLGTWGHPAFPSCGRLKEDSRYIFFMEPDANSTSRAPAAFRASFPPLETGRNLKKEV O
241 1	SRVLCKRCALPPQLKEMKSQESAAGSK O OMSERKEGRGKGKKKERGSGKKPESAAGSQSPRRN R K G D VP GP R
	II-14 II-12,
268	II-6 II-18 —— III-13
	LVLRCETSSEYSSLRFKNFKNGNELNRKNKPQNIKIQKKPGKSELRINKASLADSGEYMC
	S & S S
328	4 II-12 S
113	KVISKLGNDSASANITIVESNATSTS
1	T T
	6 II-15 8 6
354	TTGTSHLVKCAEKEKTFCVNGGECFMVKDLSNPSRYLCKCPNEFTGDRCQNYVMASFYST
173	<i>A</i>
	6
413	STPFLSLPE*
232	

Met 1	Arg	Tr	pλ	Arg	Arc	j A	la 1	Pro	Arg	g A	rg	Se:	c G	Sly	Arg	Pr	·o (Gly	Pr 1	5	Arg	ī
Ala	Gln	Ar	g	Pro 20	Gly	y S	er i	Ala	Ala	a A	arg 25	Se	r S	Ser	Pro	Pr	:o	Leu 30	Pr	· o	Leı	1
Leu	Pro	L€	eu 85	Leu	Le	u I	eu	Leu	G1;	y :	Thr	Al	a l	Ala	Leu	. A]	la 15	Pro	G]	L y	Ala	a
Ala	Ala		lγ	Asn	Gl	u A	lla	Ala 55	Pr	0	Ala	Gl	у	Ala	Ser 60	Vá	al	Cys	T	γr	Se	r
65							10															
	Va]				8	5						-										
	Ası			100							100											
	As _]	1	15						1.	20												
	Gl ⁻	0						13:	,							-						
145							120								•							
	ı Va				1	65						_	, ,									
	s As			180)						10.	,										
	e Pr		195)						.00												
	t G1 21	0						21	2													
22	a Se						230	J						23	_							
	er Ai				4	34J								_								
	et Ly			26	0						20	, ,										
			27	5						20	U											Asn
	2	90						2.	9)													Gln
3	05						31	·U						-								Ala 320
A	sp S	er	Gl	.у G	lu '	Ту: 32:	. M∈	t C	ys	Ly	s V	al	11 33	.e S	er :	Ļys	L	eu	Gly	, A 3	sn 35	Asp

Figure 26 (cont.)

Ser Ala Ser Ala Asn Ile Thr Ile Val Glu Ser Asn Ala Thr Ser Thr 340 345 350

Ser Thr Thr Gly Thr Ser His Leu Val Lys Cys Ala Glu Lys Glu Lys 355 360 365

Thr Phe Cys Val Asn Gly Gly Glu Cys Phe Met Val Lys Asp Leu Ser 370 380

Asn Pro Ser Arg Tyr Leu Cys Lys Cys Pro Asn Glu Phe Thr Gly Asp 385 390 395

Arg Cys Gln Asn Tyr Val Met Ala Ser Phe Tyr Ser Thr Ser Thr Pro 405 410 415

Phe Leu Ser Leu Pro Glu * 420

TCTAA A	AC TA sn Ty 1	C AG	A GA g As	C TG	T AT s Il 5	T TT e Ph	C AT e Me	G AT	e 11	C AT e Il	A GT e Va	T CT 1 Le	G TG u Xa	A AAT a Asn 15	ATA Ile	53
CTT AAA Leu Lys	Pro	CTT Leu 20	TGG Trp	TCC Ser	TGA Xaa	TCT Ser	TGT Cys 25	AGG Arg	AAG Lys	TCA Ser	GAA Glu	CTT Leu 30	CGC Arg	ATT Ile		101
AGC AAA Ser Lys	GCG Ala 35	TCA Ser	CTG Leu	GCT Ala	GAT Asp	TCT Ser 40	GGA Gly	GAA Glu	TAT Ser	ATG Met	TGC Cys 45	AAA Lys	GTG Val	ATC Ile		149
AGC AAA Ser Lys 50	CTA Leu	GGA Gly	AAT Asn	GAC Asp	AGT Ser 55	GCC Ala	TCT Ser	GCC Ala	AAC Asn	ATC Ile 60	ACC Arg	ATT Ile	GTG Val	GAG Glu		197
TCA AAC Ser Asn 65	GGT Gly	AAG Lys	AGA Arg	TGC Cys 70	CTA Leu	CTG Leu	CGT Arg	GCT Ala	ATT Ile 75	TCT Ser	CAG Gln	TCT Ser	CTA Leu	AGA Arg 80		245
GGA GTO Gly Val	ATC Ile	AAG Lys	GTA Val 85	TGT Cys	GGT Gly	CAC His	ACT Thr	TGA Xaa 90	ATC Ile	ACG Thr	CAG Gln	GTG Val	TGT Cys 95	GAA Glu		293
ATC TCA	TTG Cys	TGA Xaa 100	ACA Thr	AAT Asn	AAA Lys	AAT Asn	CAT His 105	GIU	AGG Arg	AAA Lys	ACT Thr	CTA Leu 110	Cys	TTG. Leu		341
AAA TA' Lys Tyr	r CTT Leu 115	ATG Met	GGT Gly	CCT	CCT Pro	GTA Val 120	Lys	CTC	TTC Phe	ACT Thr	CCA Pro 125	Ada	GGT Gly	GAA Glu		389
ATA GAG Ile Asp 130	, Leu	AAA Lys	TAT Tyr	ATA Ile	TAG Xaa 135	Ile	ATT	T								417